



L;33=00E1Na

N57

4045

CFTRI-MYSORE

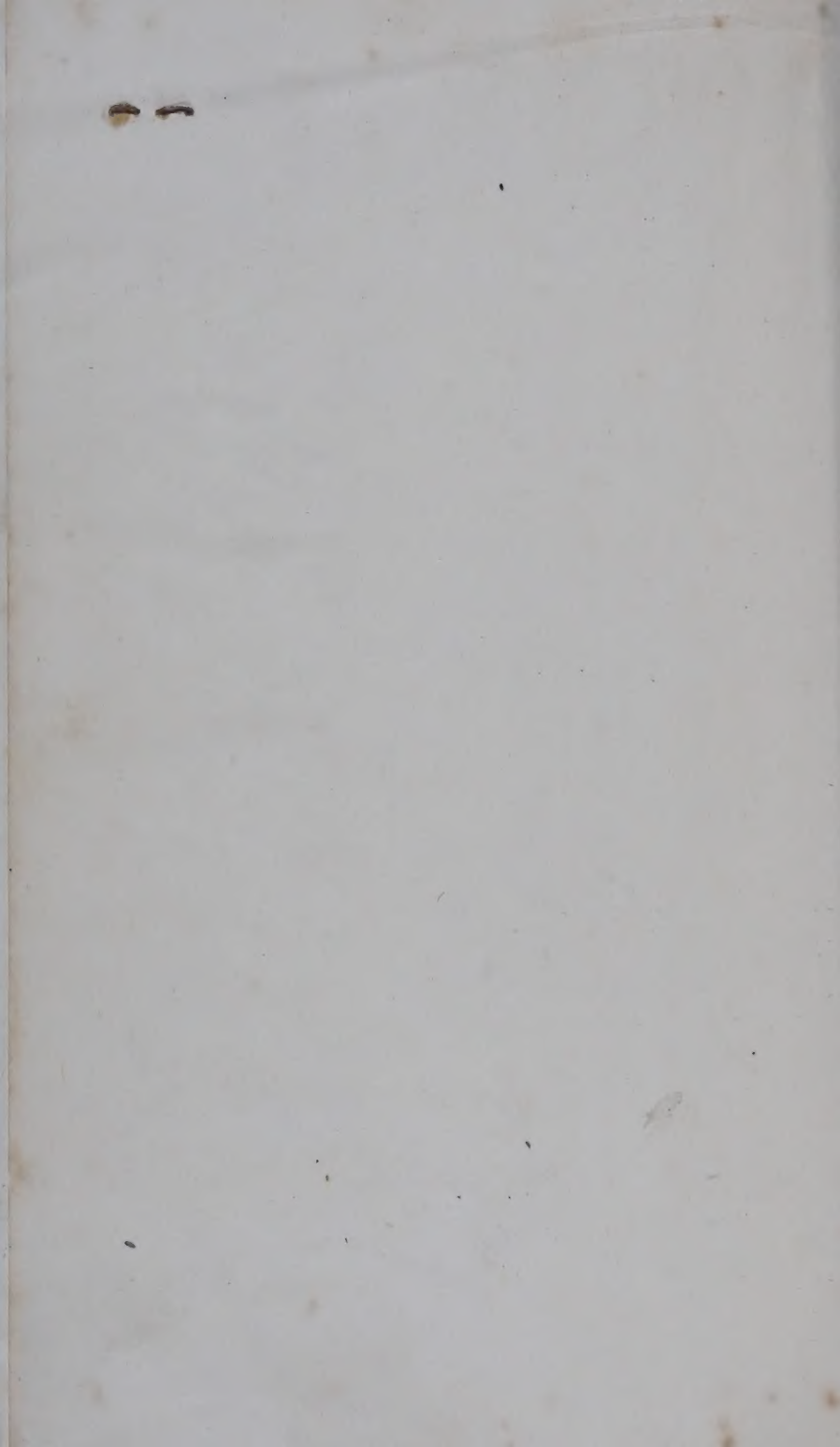


4045

Calcium metaboli..



✓  
T  
P.C.









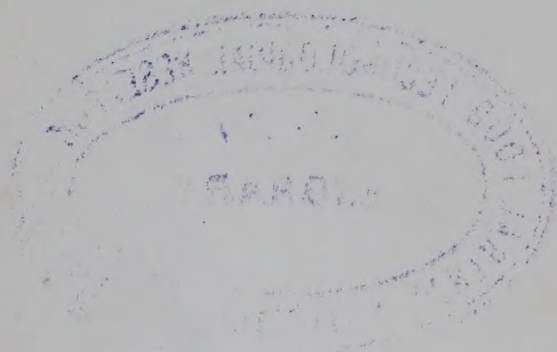
METHUEN'S  
MONOGRAPHS ON  
BIOCHEMICAL SUBJECTS

---

*General Editors*

IR RUDOLPH PETERS, F.R.S. & F. G. YOUNG, F.R.S.

CALCIUM  
METABOLISM



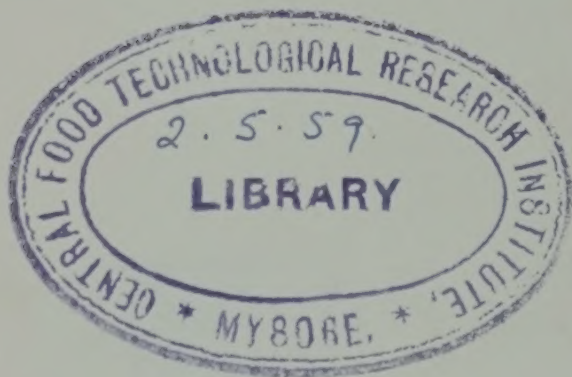
# CALCIUM METABOLISM

---

J. T. IRVING

M.A., Ph.D., M.D., F. Odont. Soc. S.A.

*Professor of Experimental Odontology; Director  
of the Joint Dental Research Unit of the Council  
for Scientific and Industrial Research, and of the  
University of the Witwatersrand, Johannesburg,  
South Africa; Hon. Member, New York Academy  
of Sciences*



LONDON: METHUEN & CO LTD

NEW YORK: JOHN WILEY & SONS INC



*First published in 1957*

4045. ✓

L; 33 = 0061 Na

N 57

CFTRI-MYSORE



4045

Calcium metabolism

CATALOGUE NO. 4146/U

PRINTED IN GREAT BRITAIN  
BY BUTLER & TANNER LTD, FROME

TO JANET



# Contents

Preface	<i>page xi</i>
I Introduction	1
II Content and Availability of <u>Calcium</u> in <u>Foods</u>	5
III Intestinal Absorption of Calcium	12
IV Intestinal Absorption of Calcium ( <i>continued</i> ) Influence of <u>Phytic Acid</u>	21
V Intestinal Absorption of Calcium ( <i>continued</i> ) The Action of <u>Vitamin D</u>	30
VI Calcium Content of the Body and of Various Tissues	36
VII Utilization and Retention of Calcium in the Body	49
VIII The <u>Dietary Requirement</u> of Calcium	71
IX The Calcium of the Blood	82
X Bone Formation and the Influence of Various Factors upon this	95
XI Bone - <u>Chemistry</u> and <u>Physics</u>	112
XII <u>Enzymes</u> and Bone Formation	123
XIII The Calcium Metabolism of the Teeth	135
XIV The Excretion of Calcium	147
XV Extra-skeletal Functions of Calcium	153
Summary	155
Index	159





## CHAPTER I

### Introduction

CALCIUM is the most important inorganic element in the body, and occurs in it in the highest amount. It is somewhat surprising that in human diets, at any rate, the supply is so meagre. The highest Ca content of foods is of course in bone, and it is a regular agricultural practice to feed farm animals on bone-meal. This dietary practice has never appealed to civilized man, who has to rely on dairy products and certain green vegetables for his supply of Ca. In spite of this small distribution in the diet, Ca deficiency is not common, and the reason for this will be explained in more detail later.

The form in which Ca occurs in foods has not been much investigated. In milk it is probably associated in part with the protein fractions, and in vegetables it must in part be in a simple soluble form since some of it is easily leached out by water during cooking.

In the intestine an acid reaction is necessary for the solution and absorption of Ca. Thus  $\text{Ca}_3(\text{PO}_4)_2$ , insoluble at neutral reaction, is readily available for utilization in the body. However, in human studies there is little description of Ca deficiency due to achlorhydria, though this has been caused experimentally. The degree of absorption of Ca is difficult to determine, owing to the concurrent processes of secretion of digestive juices containing Ca, and the possible active excretion of Ca in addition. Taking the total balance of intake and output into account, one finds a very low figure of retention in humans, but a considerably higher one in animals like rats.

The absorption of Ca is governed by a number of factors, the chief of which is vitamin D. It seems probable that the

degree of absorption is also controlled by the needs of the moment, especially during the growing period, though this has not yet been proved.

After absorption, Ca can exist in at least three forms in the body—Ca ions in solution, Ca bound to organic compounds, and Ca present in a crystal lattice as in bone. A close equilibrium exists between these three forms, governed by the endocrine glands and the dietary status, and obeying the rules of physical chemistry as closely as most biological reactions.

The excretion of Ca is controlled by factors still incompletely understood. The faecal Ca in man consists largely of unabsorbed dietary Ca, together with an unknown amount of secreted and actively excreted Ca. The kidney excretes Ca continually, even during starvation, but this may be due to the fact that the kidney tubule can deal with only certain types of Ca compounds. The urinary production of Ca is controlled by several endocrine glands.

The time-honoured method of investigating Ca metabolism is by the metabolic balance experiment. This involves an accurate knowledge of Ca intake and excretion and is tedious and often unsatisfactory to carry out. Errors can easily creep in and the full co-operation of the subject is essential. When errors do occur, they usually cause a fictitiously high positive balance of the element under consideration. As a result many writers have condemned balance methods, but unfortunately they have substituted nothing better in their place. The more recent use of radioactive isotopes has given us another tool which can be used freely in animal experiments, but which has naturally been sparingly used with human subjects.

Tetany was first described by Steinheim over 100 years ago and Gley [1] in 1893 recognized the role of the parathyroids as regulators of Ca metabolism. In 1918 Osborne and Mendel [2], working with rats, found that while the body could adapt to deprivation of most mineral elements, it was unable to adapt, either by substituting other elements or conserving what it had, to absolute Ca deficiency. The question of dietary Ca requirements was first put on a quantitative basis by Sherman

1920 [3], and while his premisses were not in all cases correct, he started an aspect of the problem which is not yet solved. H. H. Mitchell and his school have thoroughly probed the question of utilization and retention of Ca and the effects of the body stores upon these [4].

The role of Ca in bone structure has been investigated with success by biologists working in collaboration with crystallographers, especially by Hodge and his colleagues and by Pallemagne. During the last two decades our knowledge of the physiology and biochemistry of bone formation has greatly advanced; the metabolic changes in calcification appear to follow the carbohydrate phosphorylytic cycle found in other parts of the body [5]. Much remains to be sorted out, especially the function of phosphatase. Bone formation has also been studied from the point of view of both endocrine and dietary influences [6, 7].

The teeth, until recently the Cinderellas of the calcified tissues, are now receiving their due share of attention, largely in an attempt to solve the caries problem. Apart from this, much fundamental work has appeared from Schour and his colleagues in Chicago, from the Bethesda group, and from other workers such as Bevelander, Armstrong and Sognnaes. These aspects will be expanded in the following pages. And further it will be seen that Ca, although an apparently inert mineral element, undergoes some surprising adventures between its advent in the mouth and its final excretion.

## REFERENCES

- GLEY, E. *Arch. de Physiol.*, 5 Série, **5**, 766 (1893)  
OSBORNE, T. B. and MENDEL, L. B. *J. biol. Chem.*, **34**, 131 (1918)  
SHERMAN, H. C. *J. biol. Chem.*, **44**, 21 (1920)  
STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **21**, 577 (1941)  
GUTMAN, A. B. and YU, T. F. *A concept of the role of enzymes*

- in endochondral calcification*. Trans. 2nd Conf. med. Interr. New York. Josiah Macy Jr. Found. (1950)
6. BECKS, H. and EVANS, H. M. *Atlas of the skeletal development of the rat, normal and hypophysectomised*. An. Inst. of dental Med. (1953)
7. MCLEAN, F. C. and URIST, M. R. *Bone. An introduction to the physiology of skeletal tissue*. Univ. of Chicago P. (1955)

## CHAPTER II

# Content and Availability of Calcium in Foods

THE Ca content of common foodstuffs has now been widely determined and many tables giving the composition of foods have been published. The best source of Ca in human diets is milk or milk products excluding butter. Cows' milk contains about 0.12 per cent. of Ca and goats' milk has slightly more. Skimmed milk, liquid or dried, is just as good a source of Ca as whole milk. Cheddar cheese contains about 800 mg. Ca per 100 g. and thus is a very good source of Ca, but acid types of cottage cheese have only about 1/9th of the Ca content of cheddar cheese. Human milk contains far less Ca than cows' milk. Leitch [1] has surveyed the literature on this and reported figures ranging from 0.013 to 0.04 per cent., the Ca content being to some extent modified by the diet of the mother. It is an interesting coincidence, as pointed out by Bunge [2], that the Ca content of the milk has a relationship with the rate of growth of the young, as these figures show:

<i>Species</i>	<i>Days to double birth weight</i>	<i>Ca % in mother's milk</i>
Man	180	0.02
Cow	47	0.12
Dog	7	0.32

The other main dietary source of Ca is green vegetables, and especially the outer green leaves, which may contain as much as twenty to thirty times as much Ca as the inner



ones [3]. The methods of growing vegetables can influence their composition, artificial fertilizers tending to lower the C and raise the K content [4], the Ca possibly being replaced by some other element.

McCance and his colleagues [5] have analysed a number of vegetable foodstuffs. The most interesting conclusions are those relating to the effects of methods of preparation and cooking on the final Ca content of the food. They investigated the effects of steaming, boiling, baking, frying and roasting on the common constituents of vegetables. The only procedure affecting Ca is boiling. Boiling scarlet runner beans for 120 minutes and carrots for 120 minutes caused a loss of between 12 and 20 per cent. of the Ca in the cooking water. This was much less than the loss of, for example, Cl and K, of which over 60 per cent. were dissolved in the case of beans. Addition of alkali to the water made no difference to the loss. A more recent American study [6] showed that cooking cabbage in water resulted in a loss of over 20 per cent. of Ca by leaching, but with pressure cooking only 9 per cent. was lost. Averaging the results from eleven vegetables investigated, the same was found, nearly 25 per cent. of Ca being lost if the vegetable was covered with cooking water, and much less when the water was considerably reduced. It must be admitted that the Ca content of many of these vegetables is so low that these losses make little difference to the daily dietary intake. The outer leaves of cabbage contain 0.429 per cent., but carrots have only 0.045 per cent. and beans 0.055 per cent.

Several foods now have Ca added to them, the commonest being bread. In England, chalk is added to the flour to overcome the action of phytic acid, and in the U.S.A. dried milk and improvers are added, considerably increasing the Ca content. Prouty and Cathcart [7] found that the Ca content of fresh bread in the U.S. might vary from 0.065 per cent. to 0.141 per cent., six slices of bread giving 30 per cent. of the daily requirement, supposing all the Ca to be available.

One can summarize by saying that the bulk of our Ca intake comes from milk or milk products, taken either as such or

various cooked foods. The Ca from vegetable sources provides only a small fraction of the total daily intake.

*Availability of Ca in foodstuffs.* A knowledge of this is more important biologically than that of the Ca content in total amount, since it may be that the Ca in the given foodstuff is not in a condition to be nutritionally useful. This is especially true in the case of spinach and other chenopodiaceae, the Ca of which is in the form of oxalate and quite unavailable for use in the body. Spinach contains so much oxalate that it may so precipitate and render useless Ca in other constituents of the diet. In fact, Sherman [8] says that 'spinach was a mistaken choice for popularization as a typical green-leaf vegetable'. It will take some time before the feats of the seaman associated with this vegetable are forgotten.

The dipyrindyl method can be used for testing the availability of iron, but no such procedure is available for Ca and rather tedious metabolic balance experiments have to be employed. Many experiments have been done on animals, especially calves and rats. Milk Ca is taken as the reference standard and the availability of other forms of Ca compared with it. In most all inorganic forms of Ca seem to be available to some degree, such as carbonate, sulphate, phosphate and even lactate [9]. Ca lactate is also a good form of Ca and supported biological functions in rats, except growth, as effectively as the Ca of milk [10].

As already stated, the Ca of spinach is not available, and leaching removed very little of it, the Ca being quite insoluble and not extractable. The Ca of cauliflower and broccoli is not available as that of milk [11], nor is that of carrots, fresh lettuce or beans. Cooking of carrots or beans did not alter the availability [12].

These animal experiments have given useful information, but owing to species differences cannot necessarily be directly transferred to man. In general it has been found that Ca from organic sources is used as effectively by humans as that from milk, and the same applies to Ca gluconate or lactate [13, 14]. Kroyd and Krishnan [15] gave Ca lactate in a large-scale

experiment to school children in India and reported a significant improvement in weight and growth. The availability of inorganic salts appears to depend on their solubility. Steggerda and Mitchell [16] found that the availability of Ca to man from dried-milk solids or homogenized milk was the same as that from whole milk. Cheese Ca was as good as that from milk [17].

As far as vegetable Ca is concerned, as a rule this Ca is not so available as milk Ca. Breiter *et al.* [18] found that the utilization of carrot Ca was on an average 13.4 per cent, which is lower than the usual figure for milk (about 20 per cent.). The figures they give also illustrate the drawbacks of relying on vegetables as the main source of Ca. To obtain 202 mg. of Ca, 700 g. of carrots had to be eaten, this giving only about one-quarter of the daily requirement. On the other hand, lettuce Ca was even better than milk Ca when fed in amounts giving the same Ca intake, but here again the Ca content is so low (33 mg. per 100 g.) that, like most vegetables, lettuce could only be used as a subsidiary source of dietary Ca.

*Pasteurization of milk.* A great deal of controversy has raged over this process, and unfortunately considerations other than scientific ones have at times been used as arguments. As far as Ca is concerned, its availability is not very much affected. Kramer *et al.* [19] found in children and adults that the Ca in raw milk was better retained than that of pasteurized milk. Ellis and Mitchell [20] put rats on to diets of suboptimal Ca content and found that 98 per cent. of the Ca of raw milk was retained, and 92 per cent. of that of pasteurized milk. On the other hand, Henry and Kon [21] and Auchinachie [22] carried out simultaneously and independently balance experiments with rats similar to those of Ellis and Mitchell and found no differences in the availability of Ca from these two sources. Experiments undertaken by Crichton and Biggar [23] showed no differences whatever in the nutritional status of calves fed either type of milk. Steggerda and Mitchell found in man [24] that heating milk to 160° F. for 30 minutes did not affect the utilization of the Ca. It seems therefore unlikely that pasteur-

ation materially affects the nutritional value of Ca in milk, and if any slight change occurs, this is more than counterbalanced by the general improvement in the milk.

*Water as a source of Ca.* Hardness in water is expressed in a variety of ways, usually as parts of  $\text{CaCO}_3$  per 100,000 or 1,000,000. This assessment makes no distinction between the various salts which can cause hardness, and may be bicarbonates, sulphates or chlorides of Ca, Mg and other metals. For this reason it is very difficult to assess water as a source of Ca. Soft water with a hardness of 15 parts of  $\text{CaCO}_3$  per million is obviously a poor source of Ca since it contains only 6 mg. per 100 ml. (if the hardness is all Ca), and a daily intake of, say, 4 pints would give only 14 mg. Fairly hard waters would give on this basis a daily intake of about 200 mg. [25]. Really hard waters such as are found in the Kenhardt district of South Africa [26], with a total hardness of 1,000 parts of  $\text{CaCO}_3$  per million, would give 900 mg. per day if 4 pints were consumed. Walker [27] has calculated from figures available in South Africa that the Ca contributed by the drinking water is negligible in the case of one-third of the population; that half of the population get about 60 mg. daily from water; and that with less than one-sixth of the population the Ca ingested from the drinking water is from 100 to 300 mg. per day. Except in the case of the latter group, the Ca from drinking water makes an insignificant contribution to the daily Ca requirement, and in the case of the last group it is not known if the hardness is all due to Ca or how available this Ca is. It thus appears wise to disregard water as a source of Ca till more knowledge is forthcoming. Hutchison [28] in his well-known textbook considers water to be negligible in this respect.

## REFERENCES

1. LEITCH, I. *Nutr. Abst. Rev.*, **6**, 553 (1936-7)
2. BUNGE, G., *Lehrbuch der physiologischen Chemie*. 4th ed., p. 118. Leipzig (1898)



3. COWELL, S. J. *Biochem. J.*, **26**, 1422 (1932)
4. ELMENDORF, E. and PIERCE, H. B. *J. Nutr.*, **20**, 243 (1940)
5. MCCANCE, R. A., WIDDOWSON, E. M. and SHACKLETON, L. R. B. *Spec. Rep. Ser. med. Res. Council. Lond.*, **2** (1936)
6. KREHL, W. A. and WINTERS, R. W. *J. Amer. diet. Assoc.*, **26**, 996 (1950)
7. PROUTY, W. W. and CATHCART, W. H. *J. Nutr.*, **18**, 2 (1939)
8. SHERMAN, H. C. *Calcium and phosphorus in foods and nutrition*. New York: Columbia Univ. Press (1947)
9. STEENBOCK, H., HART, E. B., SELL, M. T. and JONES, J. H. *J. biol. Chem.*, **56**, 375 (1923)
10. GAUNT, W. E., IRVING, J. T. and THOMSON, W. J. *Hyg. Camb.*, **39**, 91 (1939)
11. FINCKE, M. L. *J. Nutr.*, **22**, 477 (1941)
12. SHIELDS, J. B., FAIRBANKS, B. W., BERRYMAN, G. H. and MITCHELL, H. H. *J. Nutr.*, **20**, 263 (1940)
13. PATTON, M. B. and SUTTON, T. S. *J. Nutr.*, **48**, 443 (1953)
14. PATTON, M. B. *J. Nutr.*, **55**, 519 (1955)
15. AYKROYD, W. R. and KRISHNAN, B. G. *Ind. J. med. Res.*, **27**, 409 (1939)
16. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **21**, 5 (1941)
17. MALLON, M. G., JOHNSON, L. M. and DARBY, C. *J. Nutr.*, **5**, 121 (1932)
18. BREITER, H., MILLS, R., RUTHERFORD, E., ARMSTRONG, W. and OUTHOUSE, J. *J. Nutr.*, **23**, 1 (1942)
19. KRAMER, M. M., LATZKE, E. and SHAW, M. M. *J. biol. Chem.*, **79**, 283 (1928)
20. ELLIS, M. and MITCHELL, H. H. *Amer. J. Physiol.*, **10**, 1 (1933)
21. HENRY, K. M. and KON, S. K. *Milk and nutrition*, Part 9-31. Poynder, Reading (1937)
22. AUCHINACHIE, D. W. *ibid.*, 32-6
23. CRICHTON, J. A. and BIGGAR, W. A. *ibid.*, Pt. III, (1938)



4. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **31**, 407 (1946)
5. WIDDOWSON, E. M. *Brit. med. Bull.*, **10**, 219 (1944)
6. OCKERSE, T. *Report on the incidence of dental caries among school children in South Africa*. Union of South Africa: Dept. of Public Health (1944)
7. WALKER, A. R. P. *J. S. Afr. chem. Instit.*, **24**, 20 (1947)
8. *Hutchison's Food and the principles of dietetics*. Revised by V. H. Mottram and G. Graham. 10th ed. London: Edward Arnold & Co. (1948)

## CHAPTER III

# Intestinal Absorption of Calcium

As indicated in the introduction, a number of factors are involved in the absorption of Ca in the intestine. We are still uncertain of the quantitative aspect of absorption, although some ingenious attempts to determine it have been described. It seems quite clear that Ca absorption is not intake minus faecal excretion. This assumption has been made by McCann and his school [e.g. 1], and in spite of the criticism of Stegger and Mitchell [2] has been repeated in a more recent publication [3]. Such a concept is quite untenable. Bergeim [4] showed that Ca and P entered and left the intestine at various points and Nicolaysen *et al.* [5] considered that 0.5 g. to 1 g. of Ca might be secreted daily in the digestive juices, and if this is so it can be seen that such Ca could exceed the daily intake and vitiate any intake minus excretion calculations.

Nicolaysen *et al.* [5] made a series of interesting calculations of the endogenous Ca loss in the faeces, by which is meant the loss which does not arise from unabsorbed dietary Ca. They assumed various levels of secreted Ca and also that the percentage absorption of this Ca was the same as that of food Ca. Whether this latter assumption is correct is possibly open to doubt, but their figures for secreted Ca agreed well with those calculated by Logan [6]. The results showed that at levels of Ca intake between 400 and 600 mg. daily, the absorption of food Ca varied between 52 and 84 per cent. and naturally, the figure for secreted Ca rose, so did the apparent efficiency of absorption of food Ca. Blau *et al.* [7] using the isotope dilution method of Vissek *et al.* [8] also found the absorption of ingested Ca in man to lie between 44 and 67 per cent., and

ssek *et al.* using the same method found the 'true digestibility' of Ca in cattle to be from 2 to 56 per cent. Geissberger [4] has reported similar figures in humans using  $\text{Ca}^{45}$ . These figures are surprisingly high since, as will be seen, the final utilization of ingested Ca in man seldom exceeds 30 per cent. The calculations of Nicolaysen *et al.* were based on the findings of three adults aged 29 to 58 years. Figures for children and babies are badly required since it is difficult to see how they store Ca on their known Ca intakes unless Ca absorption is extremely efficient.

It would be expected that Ca absorption would take place chiefly high up in the small intestine where the reaction was more acid. Bergeim [4] and Adolph and Liang [10] found in rats by direct analysis that Ca was absorbed in the small intestine, and Bergeim commented on the finding that Ca absorption occurred chiefly where P secretion was greatest, and considered this to be an important relationship. Both Nicolaysen [11] and Robinson *et al.* [12], working with loops of intestine or Thiry-Vella fistulae, found that the rate of Ca absorption was increased with increased concentrations of Ca in the intestine. Innes and Nicolaysen [13] found that the removal of the caecum did not affect Ca absorption. Jansen [14] observed that urinary excretion and intestinal absorption bore a relationship to each other, which McCance and Widdowson [15] confirmed, finding that the urinary excretion of Ca followed the intestinal absorption (intake minus faecal excretion), but not rising or falling to the same extent. They suggested that changes in urinary excretion might be regarded as a valuable index of changes in intestinal absorption.

Nicolaysen [16] advanced a novel and attractive hypothesis which unfortunately has not yet been confirmed [Henry and Brown, 17]. He suggested that, at any rate in rats, the intestinal absorption of Ca was a function of the Ca saturation of the body, and this was governed by an endogenous factor which needed vitamin D for it to act. The factor was present only in young animals and this was why vitamin D had little action on the absorption of Ca in older rats. There is no doubt, as

will be seen again later, that the retention of Ca is governed in some way by the level of the body stores and the utilization of Ca is more efficient with lower body stores, but no one has far has demonstrated the mechanism of this action.

*Reaction of the intestinal contents.* On purely chemical grounds it might be expected that Ca absorption would be seriously interfered with if the reaction of the intestine became alkaline. McGowan [18], on the basis of balance experiments, has stressed the importance of the gastric HCl in the absorption of Ca and considered it the chief single factor in the process. The absorption of Fe is greatly reduced in cases of achylia gastrica, and Bussaberger *et al.* [19] found that gastrectomy in puppies caused severe osteoporosis and even spontaneous fractures. The animals were in negative Ca balance. They attributed these effects to the absence of gastric HCl. To keep Ca salts in solution, the rapid transfer of food through the intestines in the absence of the stomach and also to possible cecal acidosis which might mobilize Ca from the bones.

The addition of acid to the diet, or giving an acidic type of diet, may apparently increase Ca absorption, and alkali in the diet may have the reverse effect. This has been claimed by a number of workers but mostly in acute experiments [e.g., Irving and Ferguson (20) or by the use of Thiry-Vella fistula (12, from  $\text{CaCl}_2$ , but not Ca lactate solutions)]. However, while intestinal absorption may be improved by giving acid, in many cases the urinary excretion of Ca was also increased, causing in the end a greater total loss of Ca from the body. On the other hand, it has been stated that the reaction of the diet has no effect on Ca absorption, and that alkaline solutions introduced into the intestine rapidly become acid. Similarly, it has been stated that acid solutions actually depress Ca absorption.

It seems almost impossible to draw any conclusions from the very contradictory evidence that exists. While acid solutions may for the moment increase Ca absorption, it seems probable that the contents of the intestine are well buffered so that such changes in pH are rapidly compensated for, and the

excretion in the urine is increased, the extra Ca being possibly mobilized as base to neutralize the acid introduced to the body with the food. An exception to this statement regards pH may exist in the case of fat.

*Fats and soaps.* Unabsorbed split fat in the intestine will form insoluble Ca soaps and thus may interfere with Ca absorption. Such conditions may happen when the flow of bile is obstructed. Fats have an antirachitic effect when diets high in Ca and low in phosphorus are used (like the well-known Steenbock-Black rachitogenic diet), but not with diets high in P and low in Ca [21]. This is almost certainly due to the precipitation of Ca as Ca soaps, which would equalize the Ca : P ratio with Steenbock-Black types of diets, but make the state of affairs worse with diets of low Ca : P ratio. Jones [22] investigated the effect of various substances upon the pH of the intestinal contents, since Boyd *et al.* [23] had suggested that fats exerted their antirachitic action by lowering the pH. Jones found that while lard, oleic acid and vitamin D lowered the pH to the same degree, vitamin D was much more antirachitic. Lactose also lowered the pH to the same degree as lard, but had no antirachitic action under his experimental conditions, and phosphate, also antirachitic, had no effect on intestinal pH. On the other hand, French [24], working with rats, used diets containing from 5 to 45 per cent. fat and found that the utilization of Ca fell consistently as the fat content of the diet was raised above 5 per cent., which he attributed to changes in the reaction of the intestinal contents, acidity being highest with the 5 per cent. fat diet. He thought that besides the reaction of the gut, absorption of Ca was governed by the formation of readily absorbed bile-fatty acid-complexes. Thus under normal conditions it appears that fat in the diet has within certain limits of concentration an auxiliary action on Ca absorption, the optimal fat : CaO ratio, the diet having been stated as lying between 1 : 0.045 and 0.06 [25]. From the point of view of practical nutrition in man the question is probably an academic one since Mallon *et al.* [26] and Steggerda and Mitchell [27] found that dietary



fat had, within a wide range, no effect upon Ca retention in humans. Humans do not normally eat fat-free diets or diets with a fat content above about 20 per cent.

*Protein.* Many workers have claimed that there was an association between protein and Ca in the diet, Ca absorption and utilization being better with high protein intakes. Sherman and his colleagues [28] found with rats that the total body Ca was raised during growth by increased protein in the diet, but this difference disappeared as the animals grew older. In the Chinese [29] it was found that a higher intake of protein facilitated the attainment of Ca equilibrium of intake and output, which is probably of importance in peoples habitually subsisting on low Ca intakes. McCance *et al.* [30] have produced evidence that Ca absorption (intake Ca minus faecal Ca) is dependent on the protein intake, and that such absorption may fall to very low levels in the absence of dietary protein. It has been suggested [31] that amino acids increased the solubility of the phosphates and carbonates of Ca. Hall and Lehmann [32] found that after ingestion of a powder made by mixing peptone and  $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$ , urinary and serum Ca values rose higher than if the Ca salt was taken alone and suggested the use of this powder in aiding Ca absorption. Stenger and Mitchell [2] have given reasons for doubting the interpretation that McCance *et al.* put upon their results, since, unlike these workers, Mitchell and his colleagues did not think that Ca absorption could be simply calculated by subtracting faecal Ca from that in the dietary intake.

*Fibre, bulk of stools, &c.* In general it can be said that the bulk or fluidity of the diet has no effect upon the faecal excretion of Ca, nor does constipation [33]. Excess fibre in the diet had no effect upon Ca retention in rats, but in dogs the absorption of Ca was lowered. Johnson [34] studied three patients with ileostomies and colostomies and found that the normal reaction of the terminal ileum was acid, but an excess of fibre in the diet changed the reaction to neutrality, which might possibly lower Ca absorption.

*Appetite.* A lack of Ca in the diet depresses the appetite, and



e versa [35]. This is well known to all working on Ca metabolism studies in animals, so that when comparisons of the effects of additional Ca in the diet are being investigated, it is usual to use the paired-feeding technique so that the intakes are at least isocaloric between the various animals.

**CaCl<sub>2</sub> acidosis.** It has long been known that the ingestion of CaCl<sub>2</sub> causes a profound acidosis, and this has been explained as due to a selective absorption in the intestine, the Cl ion being absorbed and the Ca excreted in the faeces as CaCO<sub>3</sub> [36]. More recently Robinson *et al.* [12], working with Thiry-Vella fistulae in dogs, have reported that the acidosis caused by CaCl<sub>2</sub> is not due to selective absorption, but to the displacement of other cations in the body by Ca, whose subsequent excretion produces a deficiency of total cations.

**Phosphate.** Under experimental conditions excess P in the diet, associated with a low dietary Ca, greatly interferes with Ca absorption and the blood Ca level falls, the animals may have tetany and bone formation is severely interfered with, giving the typical low Ca form of rickets [37]. This condition is alleviated by vitamin D dosage even when the Ca and P contents of the diet remain unchanged. This action of P is easily produced in rats and other experimental animals, but is less likely that it occurs in humans. A similar clinical picture, but for quite different reasons, is seen in the condition of renal rickets, when the kidney is unable to excrete phosphate [38], and in women with severe Ca and P deprivation, after multiple pregnancies and lactations [39]. Human bones almost without exception contain much more P than Ca, but this imbalance does not affect Ca metabolism. Only Schenring *et al.* [40] have reported a marked depression of Ca retention when phosphate was added to the diet. Two recent studies have shown that the ingestion of large amounts of P as phosphate or as H<sub>3</sub>PO<sub>4</sub> have within wide limits no effect upon the Ca balance. Malm [41] gave adult men phosphoric acid at levels of from 0.8 to 3.16 g. (250 to 1000 mg. P) daily for several weeks. The Ca intake varied from 450 to 1000 mg. per day. No effect at all was seen on the Ca balance.

The same was found when the P was given as a neutral Na-mixture to increase the level of intake of P from 1.4 to 2.4 g. daily, the Ca intake being 0.5 g. per day. Lauersen [19] (private communication) tested the effects of the daily ingestion of 5 g. of  $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$  or 2 g. of  $\text{H}_3\text{PO}_4$  on human adults. In both cases the same was found, no effect upon Ca excretion being noted. Baylor *et al.* [42] found the same with a woman who was given 2,000 mg. P as phosphate daily. Nicolay *et al.* [5] quote from their extensive unpublished material which supports the findings of Malm. It may be concluded on the balance of evidence that quite large increases in P ingestion, either as phosphoric acid or neutral phosphate, do not affect Ca metabolism in man to any significant extent.

## REFERENCES

1. MCCANCE, R. A. and WIDDOWSON, E. M. *J. Physiol.* **101**, 44 (1942)
2. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **423** (1946)
3. WIDDOWSON, E. M. and THRUSSEL, L. A. *Spec. Rpt. Ser. med. Res. Counc. Lond.*, **275**, 296 (1951)
4. BERGEIM, O. *J. biol. Chem.*, **70**, 51 (1926)
5. NICOLAYSEN, R., EEG-LARSEN, N. and MALM, O. *Physiol. Rev.*, **33**, 424 (1953)
6. LOGAN, M. A. *Physiol. Rev.*, **20**, 522 (1940)
7. BLAU, M., SPENCER, H., SWERNOW, J. and LASZLO, J. *Science*, **120**, 1029 (1954)
8. VISSEK, W. J., MONROE, R. A., SWANSON, E. W. and COMAR, C. L. *J. Nutr.*, **50**, 23 (1953)
9. GEISSBERGER, W. *Helv. med. Acta*, **18**, 461 (1951)
10. ADOLPH, W. H. and LIANG, C-C. *J. biol. Chem.*, **157**, 517 (1941)
11. NICOLAYSEN, R. *Biochem. J.*, **31**, 323 (1937)
12. ROBINSON, C. S., STEWART, D. E. and LUCKEY, F. J. *J. biol. Chem.*, **137**, 283 (1941)

- ANNES, J. M. R. and NICOLAYSEN, R. *Biochem J.*, **31**, 101 (1937)
- JANSEN, W. H. *Klin. Woch.*, **1**, 715 (1924)
- MCCANCE, R. A. and WIDDOWSON, E. M. *J. Physiol.*, **101**, 350 (1942)
- NICOLAYSEN, R. *Acta physiol. Scand.*, **5**, 200 (1943)
- HENRY, K. M. and KON, S. K. *Brit. J. Nutr.*, **1**, 147 (1953)
- MCGOWAN, J. P. *Biochem. J.*, **27**, 934 (1933)
- BUSSABERGER, P. A., FREEMAN, S. and IVY, A. C. *Amer. J. Physiol.*, **121**, 137 (1938)
- IRVING, L. and FERGUSON, J. *Proc. Soc. exp. Biol. N.Y.*, **22**, 527 (1925)
- BOOTH, R. G., HENRY, K. M. and KON, S. K. *Biochem. J.*, **36**, 445 (1942)
- JONES, J. H. *J. biol. Chem.*, **142**, 557 (1942)
- BOYD, O. F., CRUM, C. L. and LYMAN, J. F. *J. bio. Chem.*, **95**, 29 (1932)
- FRENCH, C. E. *J. Nutr.*, **23**, 375 (1942)
- HOLT, L. E., COURTNEY, A. M. and FALES, H. L. *Amer. J. Dis. Child.*, **19**, 97 (1920)
- MALLON, M. G., JORDON, R. and JOHNSON, M. *J. biol. Chem.*, **88**, 163 (1930)
- STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **45**, 201 (1951)
- CONNER, R. T. and SHERMAN, H. C. *J. biol. Chem.*, **115**, 695 (1936)
- CONNER, R. T., KAO, H-C and SHERMAN, H. C. *J. biol. Chem.*, **139**, 835 (1941)
- ADOLPH, W. H. and CHEN, S-C. *J. Nutr.*, **5**, 379 (1932)
- MCCANCE, R. A., WIDDOWSON, E. M. and LEHMANN, H. *Biochem. J.*, **36**, 686 (1942)
- LEHMANN, H. and POLLAK, L. *J. Physiol.*, **100**, 17P (1942)
- HALL, T. C. and LEHMANN, H. *Biochem. J.*, **38**, 117 (1944)
- AUB, J. C., TIBBETTS, D. M. and MCLEAN, R. *J. Nutr.*, **13**, 635 (1937)
- WATSON, E. K., MCGUIRE, E. W., MEYER, F. L. and HATHAWAY, M. L. *J. Nutr.*, **30**, 259 (1945)

34. JOHNSON, R. M. *J. clin. Invest.*, **16**, 223 (1937)
35. EPPRIGHT, E. S. and SMITH, A. H. *J. Nutr.*, **14**, 21 (1937)  
ROTTENSTEN, K. V. *Biochem J.*, **32**, 1285 (1938)
36. HALDANE, J. B. S., HILL, R. and LUCK, J. M. *J. Physiol.*, **57**, 301 (1923)  
SALVESEN, H., HASTINGS, A. B. and MCINTOSH, J. *J. biol. Chem.*, **60**, 311 (1924)
37. PARK, E. A., GUY, R. A. and POWERS, G. F. *Amer. J. Child.*, **26**, 103 (1923)  
SHOHL, A. T. and WOLBACH, S. B. *J. Nutr.*, **11**, 1 (1936)  
IRVING, J. T. *J. Physiol.*, **105**, 16 (1946)
38. MCLEAN, F. C. and URIST, M. R. *Bone. An introduction to the physiology of skeletal tissue*. Chicago: University of Chicago Press (1955)
39. ANDERSON, A. B. and BROWN, A. *Lancet*, **2**, 482 (1937)
40. LEICHSENRING, J. M., NORRIS, L. M., LAMISON, S. W., WILSON, E. D. and PATTON, M. B. *J. Nutr.*, **45**, 1 (1951)
41. MALM, O. J. *Scand. J. clin. Lab. Invest.*, **5**, 75 (1952)
42. BAYLOR, C. H., VAN ALSTINE, H. E., KEUTMAN, J. and BASSETT, S. H. *J. clin. Invest.*, **29**, 1167 (1950)

## CHAPTER IV

# Intestinal Absorption of Calcium (continued)

## Influence of Phytic Acid

This substance is inositol hexaphosphoric acid and occurs as a large part of the organic phosphorus of cereals and seeds, partly in the free state and partly as phytin, the CaMg salt. Its significance as far as Ca metabolism is concerned is that it forms an insoluble salt with Ca and thus interferes with Ca absorption in the intestine. Quite frankly the present writer is of the opinion that as stated by Horder *et al.* [1] 'the antagonism of phytin to the absorption of calcium is not highly significant' the whole question merits a fairly lengthy description as it has caused much stir in certain circles.

Mellanby [2] originally noted that certain forms of cereals have an anti-calcifying and rachitogenic power when fed to rats. This finding was confirmed by Bruce and Callow [3], who found that the agent responsible was phytin or phytic acid. They explained its action with high Ca diets by showing that the P of phytin was not available, and with low Ca diets by showing that phytic acid had a Ca salt at least as insoluble as Ca phosphate. In the latter case they thought that excess phosphate and excess of phytate acted similarly and equally on the diet in anti-calcifying power. In 1936 Lowe and Steenblock [4] confirmed these findings and demonstrated that the rachitogenic power of cereals was lessened by the hydrolysis of the phytate contained.

McCance and Widdowson [5] analysed 64 different cereal



foodstuffs for phytin and found the highest values in whole wheat meal. The content tended to vary with the total P content of the foodstuffs high in total P usually containing more phytin, but this was by no means invariable. Horder *et al.* [1] reported that whole wheat meal contained about 380 mg. P per 100 g. and white flour about 127 mg. P. In 70 per cent. extraction flour only about 15 per cent. of the total P was present as phytic acid, but in 90 per cent. extraction flour, 55 per cent. was present as phytic acid P. Thus the phytic acid content increased sharply as the extraction rate of the flour was increased. Yip and Dju [6] analysed various components of Chinese diets and stated that 25 to 50 per cent. of the P of a typical Shanghai diet might be in the form of phytin.

Following the report of Bruce and Callow, work was done to see how phytic acid acts in the diet and what practical consequences may follow from this. Harrison and Mellanby found that phytic acid had a powerful rachitogenic action equivalent to that of oatmeal on the basis of its P content. Phytic acid P was not unavailable, but acted by precipitating Ca in the gut, oatmeal containing enough to render unavailable twice the Ca in the meal, or in other words, Ca present in other constituents of the diet. This action could be overcome by adding Ca to the diet. Krieger *et al.* [8] found that the Ca in Ca phytate was as available as that of  $\text{CaCO}_3$ , but the Ca in phytic acid was not so easily used as that of phosphate [9].

These results would have had but an academic interest if it had not been for the use of high extraction flour in the United Kingdom during the recent war. This change, when 85 per cent. extraction National wheatmeal was introduced, increased the phytic acid content of the flour to virtually double and there appeared to be grave danger of slow widespread decalcification of the skeleton, especially among those habitually eating large amounts of bread.

Experiments to investigate this possibility were conducted by McCance and Widdowson in 1942 [10]. These workers carried out balance experiments on 10 subjects, 5 healthy men and 5 healthy women. They found that the absorption of



gauged by the intake compared with the faecal excretion, less if the subjects ate bread made from 92 per cent. extraction flour than if they took bread made from 69 per cent. extraction flour. The bread provided 40 to 50 per cent. of the total calories. The subjects were studied at two levels of Ca intake, about 500 mg. per day, and about 1,200 mg. On the low level of Ca intake, the subjects were in negative Ca balance on both the white and brown breads, but less so on the white bread. The data for the high level of Ca intake did not indicate the state of balance the subjects were in. This effect of brown bread could be reproduced by the addition of appropriate amounts of sodium phytate to white bread. When Ca as carbonate or phosphate was added to the 92 per cent. extraction flour to bring the total Ca intake to about 1 g. per day, all the subjects but two went into equilibrium or positive Ca balance on the fortified brown bread. One point of criticism of these experiments as reported is that one does not know the daily Ca intake of the subjects before the experiments began, but presumably it was higher than the 500 mg. or so of the first experimental period; the transfer from a high to a low Ca diet produces complications which will be mentioned elsewhere. In addition, it might be stated that the balance periods were short. McCance and Widdowson made suggestions as to the amount of Ca which should be added to flours of different extraction. They favoured  $\text{CaCO}_3$ , as  $\text{CaHPO}_4$  was visible as white specks in the baked bread, though both appeared equally effective in aiding Ca absorption. Following on this work, Ca was compulsorily added to the National wheatmeal in the amount of 7 oz. of chalk per 280-lb. bag.

This work was repeated in part by Krebs and Mellanby [11] on human subjects, using bread made from 85 per cent. and 69 per cent. extraction flours. On a Ca intake of 550 mg. per day the subjects were in negative balance on both breads, but on that made from 75 per cent. flour. With a Ca intake of 1,200 mg. per day, Ca equilibrium was attained with the brown bread. No details were given of the subjects' normal diet, nor the time allowed for the adjustment of Ca metabolism to

a possibly altered intake, but McCance and Widdowson's main conclusions were confirmed. Hoff-Jørgensen *et al.* [12] likewise showed in children that high cereal diets interfere with Ca absorption. Henry and Kon [13] repeated the work using rats. The dietary conditions were very stringent, the animals getting only white or wholemeal bread with a vitamin supplement. In spite of this, the retention of Ca from wholemeal bread was only slightly lower than that from white bread, and the addition of bone-meal or creta preparata to the wholemeal bread made surprisingly little difference to the percent retentions, a rise of 4 per cent. at most. The authors stated that their results supported the findings of McCance and Widdowson, which was true, but it was slight support.

Early in the late war the Government of South Africa introduced a National wheatmeal of higher extraction (96 per cent.) than that used in the United Kingdom. In view of the findings in the United Kingdom, the National Nutrition Council in South Africa instituted experiments to test the effects of bread on Ca metabolism. The results of these experiments, carried out on three human subjects for 19, 13 and 6 weeks respectively, showed that changing from a diet containing 1,000 mg. Ca and a small amount of bread per day, to one of about 500 mg. Ca and containing 1 lb. of standard white bread per day, caused a negative Ca balance for some weeks. This balance became increasingly less negative, and finally the subjects went into equilibrium or positive balance and easily made good their previous Ca losses on this low level of intake. The phytic acid content of the 96 per cent. extracted meal used in making the bread was 145 mg. per 100 g. weight [Walker *et al.*, 14].

The reasons for this adaptation were discussed by the writers, who pointed out that the hydrolysis of phytate was almost complete when the subjects were on the low Ca diet. This relationship, an inverse one, between the Ca level of the diet and the extent of hydrolysis of phytate, has also been reported by Cruickshank *et al.* [15]. The difficult question is where this hydrolysis occurs. Rats have a phytase in the

intestinal secretions which is a real intestinal enzyme and not of plant origin [Marcy, 9]. The same is true of guinea-pigs and rabbits, but in man no such enzyme has been demonstrated. In fact, the degree of hydrolysis in man appears to vary considerably, as judged from various reports on this, McCance and Widdowson [5] finding much less hydrolysis than Walker *et al.* [14] and Cruickshank *et al.* [15]. However, hydrolysis undoubtedly decreases the interference with Ca absorption, for McCance and Widdowson [16] found that previous hydrolysis of the phytic acid of the flour before baking the bread improved Ca absorption, and that removal of the products of hydrolysis improved Ca absorption still more. Thus on this score it must be concluded that humans are able somehow to hydrolyse phytic acid in the intestine when the Ca content of the diet is low.

Another aspect of the matter is the finding that substantial amounts of the phytic acid are hydrolysed by phytase in the flour during baking. McCance and Widdowson [17] found that the phytase of wheat flour was resistant to dry heat, and baking at 260°–360° F. did not destroy all of it. Mellanby [18] pointed out that not only the wheat but also the yeast used in the dough contained phytase and that the phytic acid content of the loaf could be reduced from an initial value of 100 mg. phytic acid P per 100 g. to one of 77 mg. while the dough was standing. The effect was more marked as the pH was lowered towards 4.5. Walker [19] likewise found that 40–49 per cent. of the phytic acid was destroyed during baking, depending on the process used. Thus it is evident that a great deal of phytate of flour is destroyed in bread-making and that the phytic acid content in flour is no criterion of that in the final product. The other point that the South African workers stressed was the ability of the body to adapt to low Ca intake, which was clearly evident from their figures. This has been noted by other workers [e.g. Steggerda and Mitchell, 20] and will be discussed in more detail later (pp. 54–56).

Field evidence in this connexion may also be considered. As Hicks [21] has pointed out that until eighty years ago,

all Europeans consumed wholemeal flour, a number still and it seemed unreasonable to assume that these ancestors suffered from an adverse Ca balance or had tetany, and the history carried more weight than laboratory experiments on a few humans for short periods. Walker *et al.* [14] cited evidence from the literature of people subsisting on high cereal diets with low Ca intakes of about 300 mg. per day, who were either in positive balance or equilibrium and who should have been ideal candidates for phytate action.

The chief shot in the locker of the protagonists of the deleterious action of phytic acid is the evidence from England when almost straight-run flour was adopted during the recent war, and an increase in the incidence of rickets actually occurred. The sequence of events is well summarized by Jessop [22]. Between September 1940 and February 1942 the extraction rate of the flour was increased from 70–72 per cent. to 100 per cent., and in addition the importation of cod-liver oil and other sources of vitamin D fell considerably because of the war. No reliable figures exist for the incidence of rickets before the question arose. The records of the radiology departments of children's hospitals showed that the incidence of rickets for August 1941 to July 1942 was almost double that of the period August 1939 to July 1940, and the mothers of the children concerned were emphatic that no change in diet or habits had occurred, the staple diet being bread and butter.

A survey was then carried out in the spring months of each year from 1943 to 1948. The incidence of rickets in children up to 4 years of age rose to its highest figure in 1944 and then declined. The figure for children 3 to 6 months of age and also that for those between 7 and 12 months also reached its maximum in 1944. The extraction rate of the flour was reduced to 85 per cent. in December 1943.

These results can be interpreted in many ways. As Davidson [23] has pointed out, there was no control experiment. The extraction rate was reduced in December 1943, but in spite of that the rate of rickets was highest in 1944 and was still high in 1945 and 1946. Furthermore, statistical analysis showed that



incidence of rickets was related to an inadequate intake of vitamin D and of milk. The fact that all the children, including twins, were affected implicates some other common factor in bread. Walker [24] has also pointed out the lack of correlation between the introduction of high extraction bread and the incidence of rickets, the time lag being  $1\frac{1}{2}$  to 2 years. He concluded his article as follows:

Though the increase in incidence [of rickets] may have been influenced by the change in the composition of the staple bread, it is certain that other factors also operated. Saunders [25], for example, considers the increase to have been largely due to the greater scarcity of foods and concentrates containing vitamin D. Whatever factors were responsible, the phenomenon can hardly be used as unequivocal evidence in support of the phytic-acid hypothesis.

The chief contention of the South African workers is that many of the investigations demonstrating the deleterious action of phytic acid have been of too short duration. Thus the more recent experiments of McCance and his school [McCance and Isham, 26; Widdowson and Thrussel, 27] were at most carried out for only 14 days. In fact, had Walker *et al.* [14] prolonged their experiments at 4 weeks, they would have agreed with McCance and Widdowson [10].

On the other hand, it must be admitted that the supply of vitamin D is much greater in South Africa than in the United Kingdom, and this may well be an explanation of the differences observed, especially since McCance and Widdowson [10]

Widdowson and Thrussel [27] have denied that any adaptation to low Ca intakes occurred in their subjects. Lanby [28] has stated that a high phytate diet demands a high vitamin D intake, and Spitzer *et al.* [29] showed that vitamin D made the P of phytate almost as available as inorganic P. In any case it is not considered in South Africa that phytic acid in the flour is of the slightest importance, any Ca added to flour being included merely to increase the daily intake; and it appears also that in the U.S.A. it is not

considered that there is any danger from anti-calcifying factors in cereals [30].

## REFERENCES

1. HORDER, LORD, DODDS, C. and MORAN, T. *Bread: chemistry and nutrition of flour and bread with an introduction to their history and technology*. London: Cassell (1954)
2. MELLANBY, E. Spec. Rep. Ser. med. Res. Council. London, **93** (1925)
3. BRUCE, H. M. and CALLOW, R. K. *Biochem. J.*, **28**, 1 (1934)
4. LOWE, J. T. and STEENBOCK, H. *Biochem. J.*, **30**, 1 (1936)
5. MCCANCE, R. A. and WIDDOWSON, E. M. *Biochem. J.*, **29**, 2694 (1935)
6. YANG, E. F. and DJU, M. Y. *Chin. J. Physiol.*, **14**, 1 (1939)
7. HARRISON, D. C. and MELLANBY, E. *Biochem. J.*, **16**, 1660 (1939)
8. KRIEGER, C. H., BUNKFELDT, R. and STEENBOCK, H. *J. Nutr.*, **20**, 15 (1940)
9. KRIEGER, C. H., BUNKFELDT, R. and STEENBOCK, H. *J. Nutr.*, **20**, 7 (1940)
10. KRIEGER, C. H., BUNKFELDT, R., THOMPSON, C. R. and STEENBOCK, H. *J. Nutr.*, **21**, 213 (1941)
11. MARCY, L. F. *J. Nutr.*, **28**, 17 (1944)
12. MCCANCE, R. A. and WIDDOWSON, E. M. *J. Physiol.*, **44**, 44 (1942)
13. KREBS, A. and MELLANBY, E. *Biochem. J.*, **37**, 466 (1943)
14. HOFF-JØRGENSEN, E., ANDERSEN, O. and NIELSEN, E. *Biochem. J.*, **40**, 555 (1945)
15. HENRY, K. M. and KON, S. K. *Biochem. J.*, **39**, 117 (1945)
16. WALKER, A. R. P., FOX, F. W. and IRVING, J. T. *Biochem. J.*, **42**, 452 (1948)



- CRUICKSHANK, E. W. H., DUCKWORTH, J., KOSTERLITZ, H. W. and WARNOCK, G. M. *J. Physiol.*, **104**, 41 (1945)
- MCCANCE, R. A. and WIDDOWSON, E. M. *J. Physiol.*, **101**, 304 (1942)
- MCCANCE, R. A. and WIDDOWSON, E. M. *Nature, Lond.*, **153**, 650 (1944)
- MELLANBY, E. *Nature, Lond.*, **154**, 394 (1944)
- WALKER, A. R. P. *J. S. Afr. chem. Inst.*, May, p. 9 (1947)
- STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **21**, 577 (1941)
- HICKS, C. S. *Med. J. Aust.*, **1**, 251 (1949)
- JESSOP, W. J. E. *Brit. J. Nutr.*, **4**, 289 (1950)
- DAVIDSON, L. S. P. *Proc. Nutr. Soc.*, **4**, 47 (1946)
- WALKER, A. R. P. *Lancet*, **2**, 244 (1951)
- SAUNDERS, J. C., *Lancet*, **2**, 580 (1944)
- MCCANCE, R. A. and WALSHAM, C. M. *Brit. J. Nutr.*, **2**, 26 (1948)
- WIDDOWSON, E. M. and THRUSSEL, L. A. Spec. Rep. Ser. med. Res. Counc. Lond., **275**, 296 (1951)
- MELLANBY, E. *J. Physiol.*, **109**, 488 (1949)
- SPITZER, R. R., MARUYAMA, G., MICHAUD, L. and PHILLIPS, P. H. *J. Nutr.*, **35**, 185 (1948)
- BRONNER, F., HARRIS, R. C., MALETSKOS, C. J. and BENDA, C. E. *J. Nutr.*, **54**, 523 (1954)

## CHAPTER V

# Intestinal Absorption of Calcium (continued)

## The Action of Vitamin D

THIS vitamin is the most important single factor in the absorption of Ca. It is formed from a sterol precursor by ultra-violet irradiation and was originally held to be part of the group of fat-soluble vitamins, later called vitamin A [Mellanby, McCollum *et al.* [2] were able to separate the anti-xerophthalmic and antirachitic properties of cod-liver oil by appropriate treatments, and later it was found that irradiation of certain foodstuffs increased their antirachitic but not their anti-xerophthalmic properties.

Steenbock and Black [3], Hess *et al.* [4] and Rosenheim and Webster [5] reported independently and simultaneously that the irradiation of cholesterol or phytosterol with ultra-violet rays converted these substances, previously inactive, into powerful antirachitic agents. It was soon clear, however, that the active substance was not cholesterol, but a contaminant which subsequent work proved to be ergosterol [Rosenheim and Webster, 6; Windaus and Hess, 7]. The active substance, called calciferol, differs from ergosterol in having the second sterol ring opened between C 9 and C 10, and is called D<sub>2</sub>, ergocalciferol, D<sub>1</sub> being a name previously given to a mixture of D<sub>2</sub> and inactive isomers and now no longer used.

It was at this time supposed that calciferol was the natural vitamin D, but it was found that it had much less activity than an equivalent amount of cod-liver oil when tested on chicks.

ss and Supplee, 8; Massingale and Nussmeier, 9], and this to the isolation of the vitamin present in cod-liver oil. This is closely related chemically to  $D_2$ . The precursor is 7-dehydrocholesterol, which after irradiation becomes the active vitamin with the opening of the second sterol ring. The active compound is known as vitamin  $D_3$  or cholecalciferol. Both  $D_2$  and  $D_3$  have an equal activity for mammals [ $40 \times 10^6$  i.u. per mg., Smith *et al.*, 10].  $D_3$  is also equivalent to 40,000 i.u. per mg. in the chicken, but  $D_2$  has a value of only 1,000 i.u. per mg. [11]. Russell *et al.* [12] and McChesney [13] found that the difference was not due to destruction of  $D_2$  in the intestine since the same difference was seen if the vitamins were given parenterally. Since most research on vitamin D has been done on mammals, ergocalciferol is the vitamin usually employed. A number of other sterols with activity similar to that of vitamin D have been described.

Irradiation with sunlight has long been known to have an antirachitic effect in animals. In fact, since vitamin D does not occur to any significant extent in foodstuffs other than fish-liver oils, humans depend to a considerable extent on sunlight for their supply of it. Children require the vitamin during the period of active bone formation, but it is still doubtful if it is needed by adults. The altitude of the sun is the governing factor for its antirachitic power [Irving and Schwartz, 14], and thus children at higher latitudes should be given supplements of a fish-liver oil during the winter.

Sunlight activates an antirachitic precursor in the sebaceous secretions of the skin [Hou and Tso, 15], which can be removed with ether. This activated precursor is subsequently absorbed into the body. Bourne [16] has suggested that birds preen themselves from the preen gland in order to obtain vitamin D which is present in the preen-gland oil, though Henniken [17] considers that in birds, too, the activated vitamin is absorbed through the skin. 7-dehydrocholesterol has been isolated from pigskin by Windaus and Bock [18], and Helmer and Jansen [19] have found that irradiated human surface skin and also the surface skin fat from irradiated human subjects

possess antirachitic activity. Mackenna *et al.* [20], however, reported that there was no provitamin D<sub>2</sub> or D<sub>3</sub> in human sebum and concluded that the provitamin was another, as yet unidentified, substance. It is possible that provitamin D<sub>2</sub> is present elsewhere in the body. Scott *et al.* [21] have demonstrated that it is made from cholesterol by a cholesterol hydrogenase, an enzyme in the intestinal wall, the small intestine and especially the duodenum having a high content of 7-dehydrocholesterol in the mucosa. Possibly ingested cholesterol is changed to 7-dehydrocholesterol in the intestine and then carried to the skin, where it is converted into the active vitamin.

In the absence of vitamin D influence the condition rickets develops. This is described in more detail in a later chapter but it can be stated at this point that it is a bone disease characterized by certain histological changes, and can be caused in rats by drastically altering the Ca : P ratio of the diet. In clinical medicine the condition is almost invariably due to lack of vitamin D, since alterations in the Ca and P contents of the diet equivalent to those produced experimentally are never met with in ordinary human dietaries.

Vitamin D has a number of activities, one of which is its effect on intestinal absorption. Schabado [22] as long ago as 1910 found that during rickets there was a big loss of Ca in the faeces, and during healing or treatment this loss was greatly reduced. Harris and Innes [23] first postulated that the action of vitamin D was to increase the net absorption of Ca and P in the intestine. This view would presuppose that rachitic cartilage would calcify properly if the supply of Ca and P were adequate. That this will happen has been demonstrated by Robison and Soames, and Shipley *et al.* [24], who showed that rachitic epiphyses would calcify *in vitro* if immersed in fluid of appropriate composition.

Smith and Spector [25] found that mineral oil interfered with the action of the vitamin if given simultaneously by mouth, the same interference occurring if the animal was irradiated; since the oil was not absorbed, it presumably



erted its antirachitic action in the gut. Nicolaysen [26] provided evidence that the vitamin acted specifically on Ca absorption, P absorption being secondarily improved, and es [27] found that a hypercalcaemia followed vitamin D dication to rats on a high Ca : P ratio diet and also considered the action of the vitamin to be on Ca absorption. He nd that the degree of healing of rickets bore a relationship he blood Ca level. Work using radio-Ca also shows that vitamin increases intestinal Ca absorption in rats [28] and ckens [29]. The exact mechanism of this action is not understood. Harrison and Harrison [30] have adduced evidence, ng  $\text{Ca}^{45}$ , that vitamin D acts on Ca absorption in the lower t of the small intestine. Nicolaysen [31], as already mened, considered vitamin D to be associated with an endous factor which controlled Ca absorption in young mals. Zetterström and Ljunggren [32] have claimed that the min is an activator of the enzyme alkaline phosphatase, ch is widespread in the body and is found in the intestinal cosa. But phosphatase has not yet been implicated in Ca orption.

Apparently there is no evidence that vitamin D acts on P orption. While it is true that addition of P to a low P diet ses rapid healing of experimental rickets [33], this healing dissimilar to that caused by the vitamin. This has been wn by Irving [33], who stated that the histological changes he epiphyses and also in the teeth differed in many respects n the two treatments. Experiments with radio-P have also n undertaken. Dols *et al.* [34] found no change in  $\text{P}^{32}$  abption when vitamin D was given to normal or rachitic rats. n and Greenberg [35] and Morgareidge and Manly [36] o found the absorption of  $\text{P}^{32}$  to be little or not at all affected he simultaneous administration of the vitamin to rachitic s. Thus the evidence seems conclusive that P absorption is governed by vitamin D.

## REFERENCES

1. MELLANBY, E. Spec. Rep. Ser. med. Res. Counc. Lon  
61 (1921)
2. MCCOLLUM, E. V., SIMMONDS, N., BECKER, J. E. and  
SHIPLEY, P. G. *J. biol. Chem.*, **53**, 293 (1922)
3. STEENBOCK, H. and BLACK, A. *J. biol. Chem.*, **64**,  
(1925)
4. HESS, A. F., WEINSTOCK, M. and HELMAN, F. D. *J. b  
Chem.*, **63**, 305 (1925)
5. ROSENHEIM, O. and WEBSTER, T. A. *Lancet*, **1**, 1025 (19
6. ROSENHEIM, O. and WEBSTER, T. A. *Biochem. J.*,  
389 (1927); *Lancet*, **1**, 306 (1927)
7. WINDAUS, A. and HESS, A. *Nachr. ges. Wiss. Götting  
Math.-physik. Klasse*, **2**, 175 (1926)
8. HESS, A. F. and SUPPLEE, G. C. *Proc. Soc. exp. Biol. N*  
**27**, 609 (1930)
9. MASSINGALE, O. N. and NUSSMEIER, M. *J. biol. Che*  
**87**, 423 (1930)
10. SMITH, M. C., GLYNN, H. E., WILKINSON, P. A. and  
PEEVERS, R. W. *Nature, Lond.*, **152**, 385 (1943)
11. BILLS, C. E. *Physiol. Rev.*, **15**, 1 (1935)
12. RUSSELL, W. C., TAYLOR, M. W. and WILCOX, D.  
*J. biol. Chem.*, **99**, 109 (1932); **107**, 735 (1934)
13. MCCHESENEY, E. W. *J. Nutr.*, **26**, 81, 487 (1943)
14. IRVING, J. T. and SCHWARTZ, H. M. *Clin. Proc. (C*  
Town), **4**, 260 (1945)
15. HOU, H. C. and TSO, E. *Chin. J. Physiol.*, **4**, 93 (19
16. BOURNE, G. H. *Nature, Lond.*, **163**, 257 (1949)
17. HANNKEN, P. B. *Nature, Lond.*, **165**, 451 (1950)
18. WINDAUS, D. and BOCK, F. *Hoppe-Seyl. Z.*, **245**,  
(1937)
19. HELMER, A. C. and JANSEN, C. H. *Stud. Inst. Divi Thom*  
**1**, 207 (1937)
20. MACKENNA, R. M. B., WHEATLEY, V. R. and WORMA  
*A. Biochem. J.*, **52**, 161 (1952)



- SCOTT, M., GLOVER, J. and MORTON, R. A. *Nature, Lond.*, **163**, 530 (1949)
- SCHABAD, J. A. *Arch. Kinderheilk.*, **53**, 380 (1910)
- HARRIS, L. J. and INNES, J. R. M. *Biochem. J.*, **25**, 367 (1931)
- ROBISON, R. and SOAMES, K. M. *Biochem. J.*, **18**, 740 (1924)
- SHIPLEY, P. G., KRAMER, B. and HOWLAND, J. *Biochem. J.*, **20**, 379 (1926)
- SMITH, M. C. and SPECTOR, H. *J. Nutr.*, **20**, 19 (1940)
- NICOLAYSEN, R. *Biochem. J.*, **31**, 107, 122 (1937); *Proc. 1st internat. Congr. Biochem.*, 167 (1949)
- JONES, J. H. *J. Nutr.*, **28**, 7 (1944)
- GREENBERG, D. M. *J. biol. Chem.*, **157**, 99 (1945)
- UNDERWOOD, E., FISCH, S. and HODGE, H. C. *Amer. J. Physiol.*, **166**, 387 (1951)
- MIGICOVSKY, B. B. and EMSLIE, A. R. G. *Arch. Biochem.*, **28**, 324 (1950)
- HARRISON, H. E. and HARRISON, H. C. *J. biol. Chem.*, **188**, 83 (1951)
- NICOLAYSEN, R. *Acta Physiol., Scand.*, **5**, 200 (1943)
- ZETTERSTRÖM, R. and LJUNGGREN, M. *Acta chem. Scand.*, **5**, 283 (1951)
- KARELITZ, S. and SHOHL, A. T. *J. biol. Chem.*, **73**, 665 (1927)
- KRAMER, B., SHEAR, M. J. and SIEGEL, J. *J. biol. Chem.*, **91**, 271 (1931)
- KEY, K. M. and MORGAN, B. G. E. *Biochem. J.*, **26**, 196 (1932)
- IRVING, J. T. *J. Physiol.*, **104**, 253 (1946)
- DOLS, M. J. L., JANSEN, B. C. P., SIZOO, G. J. and DE VRIES, J. *Proc. K. Akad. Wetensch. Amsterdam*, **40**, 547 (1937)
- COHN, W. E. and GREENBERG, D. M. *J. biol. Chem.*, **130**, 625 (1939)
- MORGAREIDGE, K. and MANLEY, M. L. *J. Nutr.*, **18**, 411 (1939)

## CHAPTER VI

# Calcium Content of the Body and of Various Tissues

**Total Ca.** This has been investigated in animals, chiefly by a number of workers. In the case of man, unfortunately and discrepant figures are available, very few indeed in the of children, and many unreliable ones in the case of adult

*Animals.* The new-born rat is virtually Ca-free. Cox and Imboden [1] found that a new-born rat weighing 5 g. contained about 14 mg. Ca. At weaning (21 days) and weighing 48 g. it contained 313 mg. Ca. The mineral content at weaning was fairly constant and females contained more Ca than males. Bessey *et al.* [2] found that 99.06 per cent. of the total Ca was in the skeleton and teeth at 2 months of age and 99.3 per cent. at 4 months. The Ca : P ratio of the body changed with the increase in weight, Ca and P contents increasing in the proportion of growth  $\times 5$ , P  $\times 7$  and Ca  $\times 12$  [3]. After 6 months of age the Ca and body-weights increased in a parallel manner [Sherman and Pappenheimer, 4]. The atomic ratio of Ca to P in the new-born was 3.5; during calcification it rose to 1 : 1, and as calcification became complete it became 6 : 5, the final value agreeing with the concept of 99 per cent. of the Ca and five-fifths of the P in the skeleton, where the atomic ratio was 3

The most recent study of the Ca content of the animal body is by Spray and Widdowson [5]. In the case of rats, they found that a large increase in Ca content occurred during the suckling period, after which the rate of accretion gradually fell until the mature percentage was reached, which occurred in the rat at about 100 days. A kink in the curve of accretion at ab-

days was presumably due to new feeding conditions following weaning. The Ca : P ratio of the body was 0.91 for the rat at birth and rose gradually to 1.75 at 246 days of age.

Ca content of adult male rats was about 1.1 per cent. and females about 1.3 per cent. in whole fat-free body tissues. These figures are considerably lower than those for man calculated on the same basis [Widdowson *et al.*, 6].

Gray and Widdowson also gave figures for the rabbit, cat, and mouse. The accretion figures for rat, mouse and rabbit are similar and all begin life with a very low Ca content. The difference in the rat is not seen in the rabbit. The pig starts with a high Ca content, but this subsequently falls in percentage till the 16th week, after which it rises to figures comparable to those of adult rats and rabbits. These authors also give useful information on the Ca : P ratio of the bodies of these animals.

A great deal of work has been done on the effect of dietary and other factors upon the Ca content of the body—most of the work being done on rats. The composition of the new-born rat is remarkably constant despite changes in the mother's diet [7], but if the mother's diet is poor, many more young are born dead [8]. With drastic changes in the mother, such as giving her a very high Ca : P ratio diet or after parathyroidectomy, the foetal Ca and P are lowered, while giving vitamin D to the mother improved the foetal storage of Ca and P [9]. Most work on the effects of the Ca content of the diet upon the body has been done by Sherman and his colleagues almost entirely on rats. Animals on low Ca diets, and retarded in growth, had total Ca contents higher than that of animals of corresponding weight but lower than that of animals of corresponding age [10]. If the Ca intake was very low, the body Ca remained stationary and the animals lived for about 6 weeks, showing at the end of that time multiple Ca deficiency but not rickets. Addition of Ca to such a diet increased the body percentage to the right figure, though the absolute amount was too low and the animals now had rickets. Campbell *et al.* [11] found that the second generation of rats on a

poor Ca diet (0.094 per cent.) could survive but could not reproduce and had only three-quarters to four-fifths of normal total Ca content of the body. The addition of potassium phosphate to a low Ca diet accelerated Ca uptake so that it was actually higher in percentage than the normal figure for that age though still low in absolute amount. With a fixed dietary P content of 0.43 per cent., increasing Ca up to 1.04 per cent. has no effect upon the body Ca; but when the P was fixed at 0.73 per cent., the Ca content of the body was increased by extra dietary Ca [12]. Shohl *et al.* [13] found in much the same way that the P intake was a limiting factor in the control of the Ca content of the body in the rat.

Sherman and Booher [14] investigated growing animal diets containing 0.16 to 0.50 per cent. Ca. Although all animals grew at the same rate and looked equally healthy, those on the higher Ca intake contained more Ca during the growth period, the difference gradually disappearing as the rats became middle-aged. In a later paper [15], it was shown that rats on a diet containing 0.64 per cent. Ca reached a plateau of total body Ca which was not attained by rats on a diet containing 0.35 per cent. Ca till 50 days later. Fairbanks and Mitchell [16] also found that the carcass Ca increased with high Ca content of the diet.

It thus appears that the total Ca content of the body can be increased by extra Ca in the diet during growth even though the growth rate and appearance of the animals are unaffected. This matter will be discussed subsequently in another connection, namely, as to whether such an increased Ca content is essential and desirable.

### Ca content of the human body during growth

*In utero figures.* Fehling [17] was the first to analyse foetal tissue. Since then many more analyses have been made and recent figures are reported by Coons [18], Iob and Swanson [19], Givens and Macy [20]. The last authors found the Ca content to rise in apparently an exponential way when plotted against age, to a final Ca content of 21 g. at term, the greatest



and coming in the last 3 months of gestation. The most important and authoritative findings are those of Widdowson and Boylston [21], who analysed fetuses and still-born children weighing from 200 g. to 4.4 kg. The total Ca rose continually from about 2 g. at the first weight to 36 g. at 4.4 kg. body-weight, the rise being linear when plotted against body-weight. At the time the fetus weighed 0.4 kg. The percentage of Ca likewise rose, but more slowly, from 0.4 per cent. to 1.0 per cent. on a whole body basis. The Ca : P ratio rose from 1.4 to 1.7-1.8, due to progressive calcification, and stayed at this latter figure until the fetus weighed about 2.0 kg. With one exception, the placenta contained negligible amounts of Ca, 29 mg. at 17 weeks and 280 mg. at term. The total Ca figures found in the literature for fetuses at term vary considerably since fetuses of different weight were analysed, but there is general agreement that the Ca percentage is 0.8 on a whole body basis.

*Ca content of the body of growing children.* Knowledge of the Ca content of the body of the child during growth is of importance in assessing the dietary Ca requirements of children, but no direct information exists on this subject. Many attempts have been made to compute figures, based on the Ca content of the new-born child, that of the adult, and various assumptions on the rate of accretion of Ca during various phases of the growth process.

The first attempt at this was made by Leitch [22]. Her assumptions led her astray, as she arrived at an adult figure of 1.6 per cent. Ca, which is double the figure found on analysis. Nicholls and Nimalasuriya [23] calculated the Ca contents of the bodies of Ceylonese children partly by analysis of bones and partly by assumptions of growth, and got values much less than those of Leitch.

Since then an analysis has been made by Holmes [24] of four different methods of computation. These were the figures of Leitch [25], Mitchell and Curzon [26], data based on the findings of Venar and Todd [27] and Holmes's own calculations. A comparison of the final figures shows how the various



assumptions made can produce different results, and incidentally how little is known of the essential data. Shohl assumed that the weight of the skeleton, as a percentage of the body-weight, rose up to 6 years of age and then remained constant, the percentage of Ca in the skeleton gradually rising as age increased. Mitchell and Curzon assumed that the gains in Ca were a constantly rising percentage of the gains in body-weight and that the weight of the fresh skeleton was a constant percentage of the body-weight. Venar and Todd based their figures upon the weights of fresh skeletons dissected from children, and proposed that there was a rise in proportionate weight of skeleton up to 6 years of age and then a decline. Holmes herself also worked out the Ca accretion on the basis that these were almost in the same proportion to those of total body-weight. In the case of Holmes's figures the Ca gains added up to a figure of 1.6 per cent. in the adult.

Holmes has pointed out that Venar and Todd's figures are quite out of keeping with what might be expected, since the greatest accretion was during the period of least growth. It has been suggested that the children they dissected were not normal. The method of Shohl, while looking 'correct' in early years, took no account of pubertal growth and presumably increased Ca storage during this time. The methods of Mitchell and Curzon, and Holmes gave results which might have been expected, since the greatest accretions of Ca occurred during the fastest growth. The Ca storage figures obtained from balance experiments in the literature were all much higher than those worked out by any of these methods.

In a later paper Mitchell *et al.* [28] analysed a fresh human cadaver and obtained a figure for total body Ca of 1.596 per cent. On the basis of this more accurate figure for the adult Ca content, these workers reassessed the estimate of the Ca increments during growth. The growth data of Mollath [29] were used, and these assumptions were made:

(a) That the Ca content at birth is 0.8 per cent. (whole body basis).

(b) That that in the adult is 1.6 per cent.

) That the change from the infantile to the adult percentage occurs progressively during growth, but more rapidly in growth is more rapid, and thus the Ca gains were tested over the period of 14 to 18 years. This final assumption was based upon findings with growing rats, chickens and dogs, and it appeared valid to apply them to humans.

Some of the calculated figures for body Ca were as follows:

Age	Wt. Kg.	Ca content g.
1	10.6	100
5	19.1	219
10	33.3	396
15	55.0	806
20	67.0	1,078

Widdowson *et al.* [6] analysed the body of a boy aged 4½ years who died of tuberculous meningitis after a two-week illness. The body was moderately nourished but rather thin and weighed 14 kg. The Ca content was given as a percentage of the fat-free body and on recalculation the total Ca is found to be 228 gr. The weight of the boy was considerably less than that given by Mitchell *et al.*'s figures (17.6 kg. at 4 years and 19.1 kg. at 5 years), and the Ca content was higher than Mitchell's figure (219 g. at 5 years). It is unfortunately impossible to argue from one analysis and that of the body of a very young child.

Mitchell *et al.* [28] comment on the fact that balance experiment data in the literature showed much higher figures for Ca retention than those calculated by their method and these figures integrated to a final Ca content much higher than that actually found in the adult body. They gave other reasons for considering isolated balance experiment data to be of doubtful value in arriving at conclusions on this matter, especially since the gross errors of balance methods all tend to increase the estimated Ca retention.

It is evident from the foregoing discussion that the existing information on the Ca content of the growing human body is scanty and has mostly been arrived at by indirect methods.

Many more chemical analyses will have to be carried out on bodies at different ages before we are in a position to approach this information with confidence to the question of requirements.

**Ca content of the adult human body.** Here too it must be admitted that while figures from cadaver analyses exist, most of them are quite unreliable and the subjects analysed were not normal. The uncertainty about exact figures has been illustrated by Mitchell *et al.* [28], who quote those given in various editions of Sherman's *Chemistry of Food and Nutrition* the values varying from 1.15 to 2.2 per cent.

Nicholls and Nimalasuriya [23] analysed bones from Chinese skeletons and arrived at a body Ca figure of 1.65 per cent. for males. More recently three analyses have been carried out which are the most accurate to date. Mitchell *et al.* analysed the body of a man of 35 years, weighing 70.55 kg. He died of cardiac failure but appeared normal nutritionally. The skeleton was 14.84 per cent. of the body-weight and contained 99.0 per cent. of the total body Ca. The percentage content of the body was 1.596. The Ca : P ratio of the whole body was 2.07 : 1. Widdowson *et al.* [6] analysed the bodies of two men and one woman. Of the two males, one died of uraemia and one of infective endocarditis, and the female committed suicide. The Ca contents in all were about the same, 2.3 g. per 100 g. fat-free weight, Ca : P ratio, 1.77. In the latest paper from Mitchell's laboratory [Forbes *et al.*, 30], a man killed as the result of a fall, was examined. He weighed 53.8 kg but was 23 per cent. below average weight for his age and height. The Ca content of the body was 1.907 per cent. on a fresh weight basis, significantly higher than the result of the previous analysis, and 2.40 per cent. on a fat-free basis, which agrees fairly well with the figures of Widdowson *et al.* The skeleton was 17.58 per cent. of the body-weight and contained 99.58 per cent. of the total body Ca. The Ca : P ratio of the whole body was 2 : 1. They concluded that the percentage of Ca in the body (fat-free basis) ranges from 1.8 to 2.5 per cent.

The present writer has instanced these figures to show what work has been done in this field and the difficulties inherent in such investigations. All writers have emphasized the difficulty of getting fresh and normal bodies and also the importance of avoiding post-mortem changes, especially desiccation during the dissection and analysis of the tissues. Many more analyses must be undertaken before we have figures reliable from a statistical point of view.

*Ca content of individual tissues.* A bare recital of the Ca content of various tissues is not very informative although Ca plays an essential role in many tissue functions. It will be seen from the following figures that in most cases the Ca level appears to be related to that of the plasma but not to be affected much by fluctuations in the plasma level. The exact level of tissue Ca is not known but it would appear to be fixed in some fairly stable way [31].

Katz [32] was the first to give accurate figures for tissue Ca, 10 mg. per 100 g. for human muscle and 6.9 mg. for dog muscle. Burns [33] found that of rat muscle to be about 7 mg. per 100 g.; she also gave figures for other rat tissues and muscles of other animals. Rat fat has a very small content, but brain has values of 8-16 mg. per cent. Dog and cat muscle was of the same order as that of rat; frog muscle had 24 mg. per 100 g. The trachea contains 0.5 mg. to 4 mg. per cent., the value increasing with age due to calcification of cartilages [2]. The Ca content of skin has been determined by Brown [34]. The values per 100 g. dry skin were highest for the rabbit (50-86 mg.) and lowest for the dog and man (31-59 mg.). Bürger and Schlomka [35] found that the Ca content of the human aorta rose with age from 0.05 g. per 100 g. dry weight at 5-7 years to 1.6 g. per 100 g. at 76 years. Widdowson [36] in a recent study found that the Ca of human liver was about 6 mg. per 100 g. fresh weight and that of the spleen was about 9 mg. per 100 g. fresh weight. The significance of this difference is not clear.

The effect of various factors on the Ca content of tissues has been investigated by several workers. Burns [33] found no



change in muscle Ca after the animals had subsisted on Ca : P ratio diets, with and without vitamin D, or on Ca : P ratio diets. The blood Ca was lowered by a variety of means but the muscle Ca remained unchanged. Injections of parathormone may have raised the muscle Ca slightly. *et al.* [36] also found that parathyroid tetany did not lower the muscle Ca. Loughridge [37] stated that parathyroidectomy lowered the muscle Ca, but his normal values were higher than those found by other workers. Haury [38] found that the Ca content of muscle, calculated on a dry-weight basis, was lower in rachitic animals. Linder [39] reported in rats that a rachitic diet raised the brain Ca but left the liver Ca unchanged. Acute ergosterol poisoning raised both the liver and brain Ca.

The giving of Ca salts apparently has little effect on the Ca content of tissues [40]. Heubner and Rona [41] found that intramuscular injections of  $\text{CaCl}_2$  into cats in acute experiments increased only the kidney Ca. In chronic experiments the Ca of the kidney and liver possibly rose.

*Bone Ca.* Most animal work has been done on rats, Pettit [42] being one of the first to produce accurate ash figures. Gaunt and Irving [43] found the following for the combined analyses of one femur, tibia and fibula (dry, fat-free) of 70-day rats:

Ash %	Ca % in ash	P % in ash
62.77	37.14-38.09	18.09-18.78

The ash content of normal rat bones (dry, fat-free) rose with age as follows [44]:

Age (days)	Sex	Ash %
21	M	48.51
	F	47.06
40	M	54.75
	F	54.91
70	M	62.77
	F	64.65
100	M	64.84
	F	65.25

The adult figure is attained by 100 days.



The analysis of separate human bones has been carried out by a number of workers. Mitchell *et al.* [28] gave a review of older findings, and Booher and Hansman [45] reported figures for the Ca of the tibiae of normal full-term foetuses. Figures for the ash of bones of adults have been found by various workers to be as follows: Ca: 35.09 to 38.80 per cent., P ratio 2.19 : 1 to 2.35 : 1. Mitchell *et al.* [28] analysed the left tibia, ulna and ninth rib of their cadaver. The ash and Ca varied from bone to bone, the ash being highest in the tibia (66.34 per cent.) and lowest in the rib (57.54 per cent.). The Ca content of the ash was more constant, varying from 36.06 in the tibia to 39.37 in the ulna. These variations show the unreliability of drawing conclusions about the total Ca of the body from the analyses of one bone as Nicholls and Mahasuriya did.

Forbes *et al.* [30] have given the most recent figures for human bones. As in the previous report from this laboratory [28] the analyses of individual bones varied considerably. The following figures were obtained on fat-free bones:

<i>Bone</i>	<i>Ash %</i>	<i>Ca % of ash</i>
Tibia	69.30	37.56
Tenth rib	52.10	36.99
Whole skeleton	56.89	40.09

In spite of wide variations in ash content from bone to bone, the Ca content of the ash appeared to be fairly constant. But the ash content of the same bone in different bodies appears to be quite variable.

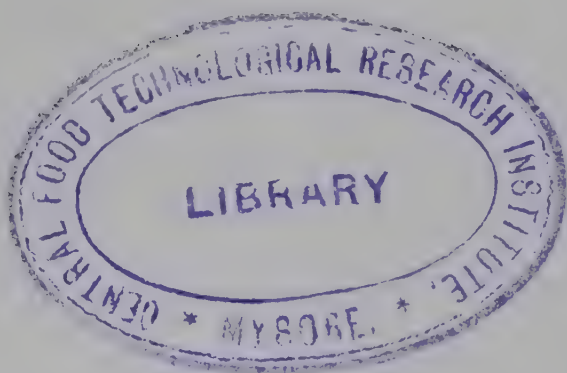
## REFERENCES

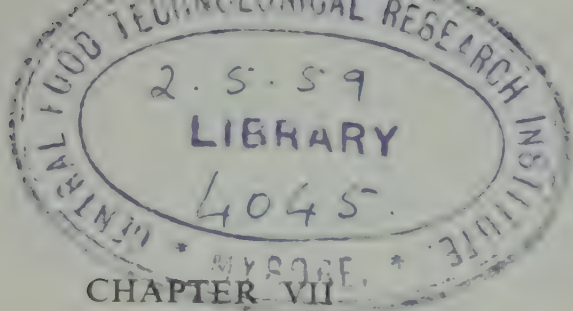
- COX, W. M. and IMBODEN, M. J. *Nutr.*, **11**, 177 (1936)  
 BESSEY, O. A., KING, C. G., QUINN, E. J. and SHERMAN, H. C. *J. biol. Chem.*, **111**, 115 (1935)  
 SHERMAN, H. C. and QUINN, E. J. *J. biol. Chem.*, **67**, 667 (1926)

4. SHERMAN, H. C. and PAPPENHEIMER, A. M. *Proc. S. exp. Biol. N.Y.*, **18**, 193 (1921)
5. SPRAY, C. M. and WIDDOWSON, E. M. *Brit. J. Nutr.*, **332** (1950)
6. WIDDOWSON, E. M., MCCANCE, R. A. and SPRAY, C. *Clin. Sci.*, **10**, 113 (1951)
7. GOSS, H. and SCHMIDT, C. L. A. *J. biol. Chem.*, **86**, 4 (1930)  
TOVERUD, G. *J. biol. Chem.*, **58**, 583 (1923-24)
8. KORENCHEVSKY, V. and CARR, M. *Biochem. J.*, **17**, 5 (1923)
9. BODANSKY, M. and DUFF, V. B. *J. Nutr.*, **22**, (1941)
10. SHERMAN, H. C. and MACLEOD, F. L. *J. biol. Chem.*, **429** (1925)
11. CAMPBELL, H. L., BESSEY, O. A. and SHERMAN, H. *J. biol. Chem.*, **110**, 703 (1935)
12. WHITCHER, L. B., BOOHER, L. E. and SHERMAN, H. *J. biol. Chem.*, **115**, 679 (1936)
13. SHOHL, A. T., BROWN, H. B., CHAPMAN, E. E., ROSE, C. and SAURWEIN, E. M. *J. Nutr.*, **6**, 271 (1933)
14. SHERMAN, H. C. and BOOHER, L. E. *J. biol. Chem.*, **93** (1931)
15. LANFORD, C. S., CAMPBELL, H. L. and SHERMAN, H. *J. biol. Chem.*, **137**, 627 (1941)
16. FAIRBANKS, B. W. and MITCHELL, H. H. *J. Nutr.*, **551** (1936)
17. FEHLING, H. *Arch Gynäk.*, **11**, 523 (1877)
18. COONS, C. M. *Oklahoma Agr. exp. Sta. Bull.*, **223**, 1 (1935)
19. IOB, V. and SWANSON, W. W. *Amer. J. Dis. Child.*, **302** (1934)
20. GIVENS, M. H. and MACY, I. G. *J. biol. Chem.*, **102** (1933)
21. WIDDOWSON, E. M. and SPRAY, C. M. *Arch Dis. Child.*, **26**, 205 (1951)
22. LEITCH, I. *Nutr. Abst. Rev.*, **6**, 553 (1936/7)

- NICHOLLS, L. and NIMALASURIYA, A. *J. Nutr.*, **18**, 563 (1939)
- HOLMES, J. O. *Nutr. Abst. Rev.*, **14**, 597 (1945)
- SHOHL, A. T. *Mineral metabolism*. Amer. Chem. Soc. Monogr. Ser. **82**, 50 (1939). New York: Reinhold Pub. Co.
- MITCHELL, H. H. and CURZON, E. G. *The dietary requirement of calcium and its significance*. Paris: Hermann et Cie (1939)
- VENAR, Y. A. and TODD, T. W. *White House Conf. Child Health Protect.*, **2**, *Anatomy and physiology*, p. 93. New York: Century Co. (1933)
- MITCHELL, H. H., HAMILTON, T. S., STEGGERDA, F. R. and BEAN, H. W. *J. biol. Chem.*, **158**, 625 (1945)
- MEREDITH, H. V. *Amer. J. phys. Anthropol.*, **28**, 1 (1941)
- FORBES, R. M., COOPER, A. R. and MITCHELL, H. H. *J. biol. Chem.*, **203**, 359 (1953)
- JOSEPH, N. P., ENGEL, M. B. and CATCHPOLE, H. R. *Fed. Proc.*, **12**, 227 (1953)
- KATZ, J. *Pflüg. Arch. ges. Physiol.*, **63**, 1 (1896)
- BURNS, C. M. *Biochem. J.*, **27**, 22 (1933)
- BROWN, H. *J. biol. Chem.*, **68**, 729 (1926)
- Quoted in COWDRY, E. V., *Problems of aging, biological and medical aspects*, p. 612. London: Baillière, Tindall and Cox (1939)
- DIXON, H. H., DAVENPORT, H. A. and RANSON, S. W. *J. biol. Chem.*, **83**, 739 (1929)
- LOUGHRIDGE, J. S. *Brit. J. exp. Path.*, **7**, 302 (1926)
- HAURY, V. G. *J. biol. Chem.*, **89**, 467 (1930)
- LINDER, G. C. *Biochem. J.*, **34**, 1574 (1940)
- DENIS, W. and CORLEY, R. C. *J. biol. Chem.*, **66**, 609 (1925)
- HARPUDER, K. *Z. ges. exp. Med.*, **76**, 709 (1931)
- HEUBNER, W. and RONA, R. *Biochem. Z.*, **135**, 248 (1923)
- HAMMETT, F. S. *J. biol. Chem.*, **64**, 409 (1925)
- GAUNT, W. E. and IRVING, J. T. Unpublished results (1940)

44. GAUNT, W. E., IRVING, J. T. and THOMSON, W. J. H.  
*Cambr.*, 39, 91 (1939)
45. BOOHER, L. E. and HANSMAN, G. H. *J. biol. Chem.*,  
195 (1931-2)





## CHAPTER VII

# Utilization and Retention of Calcium in the Body

Before going into the question of Ca retentions and requirements it is necessary to consider the endogenous Ca loss, the effect of the previous dietary Ca intake, and adaptation to low intakes.

The following definitions are usually conventionally employed in these fields. They have arisen largely as a result of the work of H. H. Mitchell and his school and have found general acceptance because of their convenience:

**Utilization.** This is applied to the Ca of one particular item in the diet, usually under test for availability, and is expressed as a percentage of the intake. It is calculated by a special formula.

**Retention.** This applies to all the Ca ingested compared to total excretion and is usually but not invariably expressed as a percentage of the intake.

**Endogenous Ca loss.** By this term is meant Ca arising from processes other than those involved in growth, and such a loss occurs in the faeces and also in the urine. It is sometimes referred to as the 'maintenance requirement'. Exactly how it varies is not yet completely understood. Possibly a constant turnover in bone as found by Smith and Smith [1] may contribute to it. It is well known that the kidney does not hold back Ca during periods of Ca lack as it does, e.g., Na [2]. Smith and Mitchell [3] considered it might be solely due to



unavoidable leakage through the kidneys and intestinal wall caused by physico-chemical processes.

Hamilton [4] first raised the question. He pointed out that normal infants excreted about 200 mg. of CaO per day in perspective of intake or weight. He therefore considered that the Ca must be used for some process of internal metabolism other than calcification, and if the intake fell below this level then calcification suffered, not the internal metabolism. Barr *et al* [5] investigated a number of subjects on low Ca intake and who were all but one in negative Ca balance. From the results obtained they suggested that Ca 'katabolized' was not available for use again and was excreted. Even in the case of a pregnant woman on a low Ca intake, the Ca loss was the same as if she had not been pregnant, suggesting that the Ca lost was not available to the foetus. Pugsley [6] found that the increased Ca excretion which followed weaning in the mother rat could be abolished if the mammary tissue was removed, and commented on the fact that the Ca absorbed from the involuting gland could not be immediately used in the animal's bone. Bell *et al.* [7] considered that the removal during growth of Ca from the inside of the bone-shaft and redistribution on the outside formed a kind of endogenous process going on continually.

Mitchell and his school have made an intensive study of this problem. Ellis and Mitchell [3] and Fairbanks and Mitchell [8] compared the Ca retentions of young rats on diets with different Ca contents. From their results they concluded that there was no Ca requirement for maintenance in growing animals, the Ca being all used in growth and calcification processes. The same conclusion was reached by Kinsman *et al.* [9] using an ingenious calculation. Their experiments were actually undertaken to test the utilization of the Ca of milk by pre-school children. Mitchell and McClure [10] had argued that if there were an endogenous loss of Ca this would appear to make the retention of Ca less at lower levels of intake compared with that at higher levels, since the Ca lost at lower levels would come partly from unabsorbed Ca and

tly from the body itself. To overcome this, Kinsman *et al.* used this formula to work out the degree of utilization of the milk Ca:

$$\frac{\text{Retention for period A} - \text{retention for period B}}{\text{Intake for period A} - \text{intake for period B}} \times 100 = \text{percentage utilization}$$

period A being that of higher Ca intake and B that of lower intake. This method enabled the authors to see if there was endogenous loss, provided that this loss was not affected by the Ca intake, and provided further that the Ca intakes in the two periods did not exceed the minimum requirement for maximum retention. The figures they gave showed that there is no endogenous loss of Ca or maintenance requirement in children, since the percentage retentions in both periods were the same and agreed with the percentage utilization figure of milk Ca. One example from their paper gave the retentions in periods A and B as 19 and 20 per cent., and the utilization of the extra Ca as 18 per cent. Had there been an endogenous loss of, say, 50 mg. per day, the figures would have been 11 per cent. for period A and 6 per cent. for period B. The utilization, of course, remaining at the same figure of 18 per cent. This demonstrates the validity of their formula and shows clearly that, as with growing rats, there is no endogenous loss of Ca in children. Ellis and Mitchell [3] say 'in a young growing animal these "used" ions as well as the calcium coming directly from the alimentary tract may be absorbed and retained by the developing bones and the growing tissues at such a rapid rate that, at low levels of Ca intake, the threshold of excretion by the kidneys and the intestines is never reached.' Kinsman *et al.* also discussed the whole question at some length.

In adults there is undoubtedly an endogenous loss of Ca and many methods have been used to calculate this. Mitchell and Curzon [11] reviewed the whole subject and compiled a graph from 139 average observations in the literature regarding intake and output of Ca. A straight line was fitted to

the data by the method of least squares and possessed formula:

$$y = 0.6826x + 3.0940$$

where  $y$  was the Ca output in milligrams per kilogram body weight per day, and  $x$  the intake expressed in the same units. The line intersected the Ca output abscissa at 3.0940, which represented the endogenous Ca excretion when there was no Ca intake. This figure would be about 200 mg. per day for a 70-kg. man. Steggerda and Mitchell in a more recent paper [12] found the endogenous loss to be 3.72 mg. per kg. body weight per day. The calculations of Nicolaysen *et al.* [11] have already been referred to. Using their assumption that Ca secreted and reabsorbed in the intestinal tract, total endogenous losses of from 261 to 499 mg. per day were obtained (faeces and urine), the figure rising with higher assumed figures for unabsorbed digestive juice Ca. Unfortunately the weights of the subjects were not given, but the present writer feels that these figures are on the high side. Blau *et al.* [14] used the isotope dilution method of Visek *et al.* [15], and assumed that the specific activity \* of faecal endogenous  $\text{Ca}^{45}$  is the same as that of the serum  $\text{Ca}^{45}$ . Visek *et al.* [15] worked on cattle and they found that the faecal endogenous Ca did not rise with increase of Ca intake but seemed to be lower in animals on a low Ca diet. Blau *et al.*, working with two human subjects, found a daily endogenous faecal loss of 91 mg. in one and 117 mg. in the other, figures which are the whole lower than those of Nicolaysen *et al.*

It is possible, by means of a back calculation using the formula of Kinsman *et al.*, to work out the endogenous Ca of adults from Ca balance figures. From findings of Steggerda and Mitchell [16] a figure of 163 mg. per day or 2 mg. per kg. body-weight is obtained. Calculations from the figure of Owen [17] and Owen *et al.* [18] gave similar results. In

---


$$\text{* } \frac{\text{Ca}^{45}}{\text{Ca}^{40} + \text{Ca}^{45}}$$

subject on a low Ca intake, the daily endogenous loss was 2 mg., or 1.8 mg. per kg., but on higher intakes this rose over 200 mg. in the same subject. On more liberal intakes the endogenous loss was higher still. If the assumptions made are valid, the calculations made from Steggerda and Mitchell's figures and those of Owen show that the endogenous loss is not independent of the Ca intake as had been thought. This is in fact implied in the statement in Holmes's article [19] that 'during periods of generous intake, endogenous losses would appear and a maintenance requirement for calcium, in the usual sense of the term, would develop'. One can thus conclude that the appearance of 'endogenously' derived Ca in the excreta depends largely on the avidity of the body for Ca. In growth, this demand prevents such an appearance, but in adults, when Ca is not so readily taken up by the body, the threshold of excretion is exceeded. The adult body seems, however, to have lost the ability of the growing body to react thus in emergencies, as seen, for example, in lactation, when much Ca may be lost. When the adult is in Ca equilibrium the daily Ca intake merely has to satisfy this endogenous loss.

*Effect of previous dietary intake.* It has not been sufficiently realized by workers using balance metabolic methods that the results of such methods must be carefully assessed before conclusions are drawn, and in particular the previous Ca intake must be known and the length of time the subject has been on this level. If this is not considered and a group of people are chosen at random and put on to the same Ca level of intake, it may well be that half will go into positive balance and half into negative balance, and it would be assumed that they had quite different Ca requirements.

This question was first raised by Fairbanks and Mitchell [8] and more convincingly by Rottensten [20], who found that the Ca retention of rats depended on their previous Ca level of intake. Rottensten had one group of animals on a level of 0.5 per cent. dietary Ca and another on 0.8 per cent. Ca. Their retentions were 85 per cent. and 50 per cent., respectively. Paired feeding was employed and growth was the same



in both groups. After 4 weeks, all rats were put on to a diet containing 0.4 per cent. Ca for 5 weeks. The animals previously on the low Ca diet now retained about 70 per cent. Ca and the retention of the others remained at 50 per cent. The animals were indistinguishable in weight and appearance and had they been investigated by someone ignorant of their previous dietary history it would have been concluded that they were two quite different types of animals. Similar findings have been reported by Carlsson [21] using  $\text{Ca}^{45}$ .

Experiments by Owen [17] and Owen *et al.* [18] have demonstrated the same in humans. Owen [17] studied a group of people who were in balance on 520 mg. Ca per day and who had been attending a diabetic and obesity clinic so that their previous dietary history was well known. Owen *et al.* [18] investigated indigent men whose dietary history was also known and who were found to be in equilibrium on a Ca intake of 240 mg. to 280 mg. per day. When their intakes were raised to 550 mg. per day they went into positive balance and stored Ca. Thus here too were two different groups of individuals whose metabolic balance behaviour was quite different and had been conditioned by their previous dietary history.

*Adaptation to low levels of intake of Ca.* It can be concluded that the body has considerable ability in this respect, in spite of statements to the contrary.

The first quantitative findings supporting this concept were those of Kelly and Henderson [22] and of Kung and Yeh [23] who reported that Africans and Chinese were in Ca equilibrium on extremely low intakes of Ca without apparent impairment of health. Nicholls and Nimalasuriya [24] found the same in Cingalese and suggested that an adaptation to a low Ca intake had occurred. That this adaptation was not unique to races other than white people had also been shown by Owen *et al.* [18] in the paper quoted above. Kraut and Wecker [25] emphasized the power of adaptation possessed by the body to low Ca intakes and Walker *et al.* [26] showed by balance experiments carried out over long periods that not only could adaptation occur but Ca losses could be maintained.



while the subject was still on the low Ca diet. The total loss while adaptation was occurring was in one subject only 2 g.

One of the most important recent papers in this field is by Hegsted and his colleagues [27], who studied prisoners at Lima, Peru. These writers found that intakes as low as 100 to 200 mg. Ca per day were adequate for equilibrium, the requirement being related to the Ca reserves and to the previous intake. Similarly, Nicolaysen *et al.* [13] in long-term experiments have shown that 13 out of 14 adult male subjects adapted to low Ca intakes in from 2 to 8 months.

The chief antagonists to the theory of adaptation are the McCance school. In their phytic acid paper [McCance and Fiddowson, 28] they denied that any adaptation had occurred, and in a later publication [29], in which some Germans had been studied on low Ca intakes, they virtually poured scorn on the whole concept, but did admit that 'it was possible that they [the subjects] had been eating at home amounts of milk and cheese, additional to the official rations, which they did not disclose'. As usual, the balance periods were short. More recently McCance [30] has reiterated his views, with little additional evidence, including an account of an attack of tetany which he claimed that he himself had after eating a brown-bread diet for a fortnight [31].

One can conclude that the vast majority of workers in this field feel with Steggerda and Mitchell [32] as follows:

It appears that in the presence of an inadequate supply of any nutrient, including calcium, the body can adjust itself to the situation, either by a more economical use of what little is available, or by a lowering of its own requirements, so that eventually it comes into equilibrium with the limited food supply. Only during this adjustment period can the body be adjudged undernourished, since only in this period is the body suffering a loss of nutriment. When adaptation is complete, the body replaces from its restricted supply all losses of the nutrient from its body, and unless some

subsidiary ill effects ensue, it may reasonably be regarded as adequately supplied with food.

In a classical paper dealing with the whole question of adaptation to undernutrition, Mitchell [33] added:

If I were to edit this statement to-day, it would be only as a safeguard against a possible implication that the body can adjust to any nutritive level, however low. Obviously, adjustment is possible only within certain limits at present unknown.

It seems clear that the mechanism of adjustment is largely a function of the level of the Ca stores in the body, Ca being utilized more efficiently as the stores become depleted [Rottensten, 20]. Rottensten himself pertinently stated:

In this connection one might naturally ask if it is desirable to keep the Ca stores filled. Our present knowledge is insufficient to answer this question, but it does seem likely that Ca stores may be considerably reduced, and Ca utilization consequently improved without any harmful effect on the individual.

*Lactose.* The interesting influence that this sugar has on the retention of Ca should be mentioned at this stage. This influence appears to be specific for this sugar and is not shared by any other carbohydrate.

Parathyroid tetany is alleviated in animals [34] and humans by lactose [McCullagh and McCullagh, 35]. Mills *et al.* found with children that lactose in the diet increased the retention to a marked degree (33.5 per cent. on an average) and these children excreted less Ca in the urine while taking lactose, and Mills *et al.* considered that lactose exerted its effect not by rendering the dietary Ca more absorbable in the intestinal tract but by making the absorbed Ca more utilized. This view is contrary to that of Bergeim [37], who attributed the action of lactose to an increased acidity of the intestinal contents. Jones [38], however, found that lactose had no

thitic effect in his experimental conditions and made the intestinal contents no more acid than did lard, and Outhouse *et al.* [39] found likewise that lactose had no effect on intestinal pH.

Not only is retention increased by lactose, but in addition the bone ash is increased [Outhouse *et al.*, 40], as is the body content of Ca and P [Mitchell *et al.*, 41]. French and Cowgill [42] found the retention effect to occur only in young rats and dogs.

Duncan [43] has drawn attention to some significant recent work by Fournier which he has interpreted to mean that lactose plays a specific role in the function of the bone cell. If this is so, the small amounts absorbed unchanged must be responsible, since galactose is apparently inactive in this respect. Fournier [44] quoted results showing that lactating rats getting 12 per cent. lactose in their diets stored Ca during the experimental period, while those with starch instead of lactose stored a large amount of Ca. Fournier suggests that lactose belongs to a group of 'structural' carbohydrates, in contradistinction to other sugars which he calls 'energetic' carbohydrates. As Duncan states, 'if the effects of lactose on the serum Ca in rats as well as those on bone composition be taken into account, a wider concept of its role than that suggested by Fournier emerges, namely, the possibility that lactose can take part in the biochemical processes fundamentally concerned in calcium metabolism'. Arguing teleologically, this may well be the reason for the presence of lactose and not some other sugar in milk, and when cows' milk is modified for infants lactose should be added to it.

*Ca utilization in adult humans.* Unless on a previously poor Ca intake, no Ca retention occurs in adults. Ca utilization has been studied by a number of investigators, especially Heggerda and Mitchell and their colleagues. Subjects with a previously normal Ca metabolic status were employed. Heggerda and Mitchell [16], working with a normal adult man, studied the utilization of Ca in milk and Ca gluconate and found it to be 20 per cent. in each case. In two later papers

working with male subjects [32, 45] utilizations of 18 to 49 per cent. and 16 to 45 per cent. were found, averaging 29 per cent. and 32 per cent.; in both papers the Ca of milk and various milk preparations or Ca salts were investigated. In still later paper they found in 13 subjects a much higher average for utilization of milk Ca, 50.2 per cent., but were unable to explain this difference [12]. Geissberger [46], using  $\text{Ca}^{45}$ , found a utilization of 30 per cent. when taken by mouth in the form of gluconate, but a much higher apparent utilization if the radio-Ca was given intravenously, due probably to a low level of intestinal absorption of the oral radio-Ca and to exchange of the parenteral radio-Ca in the bones. Patten and Sutton [47] compared the utilization of Ca in the form of lactate, gluconate, sulphate and carbonate by young college women. The utilization was the same in all, about 18 per cent. Breiter *et al.* [48] drew attention to what is a probable explanation of many discrepancies, that there are high and low Ca utilizers. They found among seven subjects that one group utilized 30 to 35 per cent. of milk Ca and the rest only 15 to 20 per cent. This aspect has also been stressed by Nicolaysen *et al.* [13], who reported that individuals on a constant Ca intake, and observed over long periods of time abruptly changed their retentions and went from no apparent reason from positive to negative balance and back again, the changes being chiefly caused by alterations in faecal excretion. Thus, as has been stressed above, in investigating this aspect of Ca metabolism, the need for long studies is imperative since the individual's Ca status is probably always changing. The reasons for these changes in retention are not yet known.

When subjects on insufficient Ca intakes are tested for the utilization of extra Ca, much higher utilization figures are found. Recalculation of the figures of Owen *et al.* [18] showed that with their subjects on low Ca intakes, the utilization of extra Ca varied from 44 per cent. to as high as 71 per cent.

*Ca metabolism in old people.* Senile osteoporosis is so well known that many investigations have been undertaken to determine its cause and possible prevention.



Owen *et al.* [18] found that old men who had been subsisting for a long time on Ca intakes of about 250 mg. per day, and some of whom had osteoporosis, were in Ca equilibrium, and when the Ca intake was increased, went into positive Ca balance, and thus had not lost their ability to store Ca. Unfortunately the experiment could not be conducted for a sufficiently long period to see if the osteoporosis disappeared. Similar findings have been reported by Nicolaysen *et al.* [13], who in long-term experiments found that elderly men stored Ca on an intake of 900 mg. per day, indicating a probable previous Ca deficiency, but also indicating that these old men could still retain Ca if they got enough. Vinther-Paulsen [9] likewise found in old patients the incidence of senile osteoporosis to be related to the level of Ca intake.

Cooke [50], however, concluded that it was doubtful if malnutrition was a common agent in the production of osteoporosis, since Ca or P or vitamin D deficiency should produce rickets or osteomalacia. There is no doubt that many other factors besides Ca deficiency can cause osteoporosis. Immobility is a well-known cause, producing a large Ca loss in the urine, irrespective of the Ca intake [Howard *et al.*, 51], and the inactivity of elderly people may cause the bony changes. Endocrine effects must also be considered, though little is known about this in old age. Albright *et al.* [52] reported postmenopausal osteoporosis in women, and from analogy of the action of oestrogens on bone in birds (see p. 106) tried oestrogen therapy. They found strongly positive Ca balances and after a long interval an improvement in the X-ray appearances of the bones [Albright, 53]. It should be pointed out that the X-ray diagnosis of bone changes is extremely crude, that at least 30 per cent. of bone salts must be lost before a change can be detected [Babaianetz, 54].

Thus it is not easy to conclude whether senile osteoporosis is due to Ca deficiency, or to an altered mode of life, or to an endocrine imbalance, or possibly to all of these combined. It is undoubtedly an increasingly important aspect of geriatrics.



### Ca retention and utilization in children

*Retention.* This whole subject has been well summarized by Holmes in her article on Ca requirements during growth [54]. The dietary Ca retentions of infants are relatively high. Aikman [55] concluded that the Ca of breast milk must be very nearly completely retained for growth, and Telfer [56] observed a retention of 60 per cent. of milk Ca by breast-fed infants. If this will be seen, a high retention must be expected for infants to get enough Ca to supply their daily requirements. In older children, the percentage retentions are seldom more than 20 per cent. and usually less. Kinsman *et al.* [9] and Outhwaite *et al.* [57], working with pre-school children, found figures of about 20 per cent. Watson *et al.* [58] studied pre-school children at two levels of Ca intake, 1,122 and 775 mg. per day. The retentions were higher in absolute amount on the higher level of intake, but the percentage retentions were about the same, being in both cases unusually low, around 10 per cent. Even with a poor previous dietary history, Danforth [59] found that 3 pre-school children retained only 17 to 18 per cent. when the Ca intake was about 1,150 mg. per day. Johnston *et al.* [60] found that girls aged 13–14, known to have been on high Ca intakes, retained 26 per cent. of the intake of Ca. McLean *et al.* [61], in a long study in which precautions were taken to see that the previous dietary history did not complicate the findings, found the average retention in boys aged  $2\frac{1}{2}$  to 6 years to be 17 per cent.

Calculated figures for the daily retention per kilogram body-weight are given by Mitchell and Curzon [11]. They show that the amount, which is very high in early life (100 mg. at birth), falls considerably in the first 6 years to 3–4 mg. and rises during the second period of rapid growth, from the 6th to the 16th year, to about 6 mg., finally falling as adulthood is reached. More recent calculations [Mitchell *et al.*, 62], based on the analysis of a male subject, were essentially the same. The figures integrated to the adult Ca content. Sherman and Hawley [63] had summarized the information in the literature

to 1922 and reported figures higher than those of Mitchell and his colleagues. Almost all the balance work on children has given results higher than Mitchell's figures. Thus Watson *et al.* [58] found in children between 38 and 55 months of age daily retentions of 4.6 to 10.8 mg. per kg., the values of Mitchell *et al.* [62] being between 2.5 and 4.2. Outhouse *et al.* [7] investigated children aged from  $3\frac{3}{4}$  to 6 years. By recalculation of their percentage retentions it is seen that the children retained from 6 to almost 9 mg. per kg. daily. Children with a poor previous dietary history would, as might be expected, retain more. Thus three children aged from  $3\frac{1}{4}$  to almost 5 years of age studied by Daniels [59], and who had retarded skeletal development, retained from 10 to 11 mg. per kg. A severely malnourished child of  $3\frac{1}{2}$ , investigated by Earne and Moore [64], retained from 50 to 68 mg. per kg. In the three periods of study when put on to an adequate diet. Daniels [59], from her studies of the three skeletally retarded children and the fact that they gained weight at a rate faster than normal, considered that her figures and those of similar magnitude in the literature were abnormally high and indicated that the subjects studied had suffered from a previous Ca lack. This would explain the findings of Hunscher *et al.* [65] and Jeans *et al.* [66], who found that as the Ca intake rose, the retention increased. Macy, in her well-known study [67], investigated two groups of boys. One group, which had been on a generous mixed diet with a quart of milk per day for three months before observation, retained 103 mg. of Ca (in absolute amount) per day, while a comparable group on the same diet, but without the extra milk, retained 473 mg. of Ca. The children studied by Macy [68] had in general low retentions of Ca in milligrams per kilogram body-weight, and they had had a generous Ca intake for the previous 5-6 years.

It has been suggested that other factors can vary the retention of Ca. Thus Johnston [69] found that the percentage retention of Ca could be increased in girls by increasing the vitamin D intake, and Holmes [19] has queried whether the

children studied by Kinsman *et al.* [9] got enough of vitamin.

Many of these findings illustrate the unreliability of isolation balance methods in children. The Ca retentions as calculated by Holmes were in general much less than those found in balance experiments, and as noted above, the figures from balance experiments often integrate to a final figure in excess of the adult Ca content. For this reason, many of the reported findings must be rejected.

*Utilization of dietary Ca.* As would be expected from what has been said above, the utilization of extra Ca is the same as the figure for the retention of total Ca [Kempster, Kinsman *et al.*, 9].

Thus in both children (except possibly infants) and adults Ca utilization is seen to be surprisingly inefficient, compared with rats who may retain 100 per cent. of ingested Ca. Mitchell and Curzon stated in 1939, 'quantitative information on the extent of these wastages of dietary calcium in both human and animal nutrition is woefully inadequate'. This statement is still true.

### **Pregnancy and lactation**

The combined processes of pregnancy and lactation impose a severe strain on the maternal organism, especially if the dietary intake is poor. This applies particularly to women owing to the Ca content of the maternal milk. Thus Goss [71] found that 60 to 70 per cent. of the total Ca of the bodies of rats could be withdrawn by repeated pregnancies and lactations coupled with a low Ca diet. The Ca of the human mother's milk may represent from 15 to 41 per cent. of the total intake [72] and repeated pregnancies and lactations may cause a lowering of the blood Ca to 6.9 mg. per 100 ml. serum, with positive Chvostek and Trousseau's signs [73].

*Pregnancy.* From the point of view of Ca metabolism, pregnancy has but a slight effect on the healthy organism. In the case of the human, calcification of the foetus demands 35 g. Ca at the very most, which means a drain of about 3.5 per cent.

from the maternal stores, supposing that no extra Ca was given in the diet. In the case of the rat, rabbit and mouse [Gray and Widdowson, 74], the newly born animal is virtually Ca-free and its calcification imposes little strain on the mother. The same generally applies to women if the intake is adequate, though great variations occur [75]. Left to themselves, women tended to choose a diet with a high Ca content [76].

The first and classical balance investigation was carried out by Hoffström [77], who studied a woman from the 17th week of term. The Ca balance was positive most of the time, just enough Ca being stored for the needs of the foetus. Many workers have reported, on the other hand, that at the end of gestation the maternal organism has stored far more Ca than needed for the products of conception. Thus Goss and Schmidt [78] found that rats stored during pregnancy four to five times as much Ca as was contained in the foetus, placenta and uterus. Even if the young were resorbed owing to vitamin E deficiency, this extra storage of Ca still occurred. Forbes *et al.* [79] found a rapid storage of Ca at the end of gestation in cows and Landsberg [80] has reported the same for women. Hummel *et al.* [81] carried out balance experiments on a primipara over the last 65 days of pregnancy, and who, on a normal Ca intake, retained more than twice the Ca needs of the foetus. In this case, the patient had an unsatisfactory dietary history and the authors were inclined to attribute the high Ca storage to replenishment of maternal stores. They contrasted this woman with a multipara they had previously investigated who retained, as in Hoffström's case, just enough Ca for the foetus. Coons and Coons [82] studied a case from the 31st to 38th weeks of pregnancy and also found Ca storage at this lower level. Thus there appears to be considerable variation in the storage of Ca at the end of pregnancy, and since the previous Ca intake has not always been considered, it is quite possible that this may have been insufficient, especially as the Ca intake is often deliberately increased during pregnancy. It seems safe to conclude that any extra storage of Ca in the mother's body during pregnancy is due to previous deficiency,



and if the Ca intake has been adequate, Ca storage does not occur.

It has been shown that  $\text{Ca}^{45}$  moves easily across the placenta and a higher percentage of  $\text{Ca}^{45}$  was found in the foetal than in the maternal bones [Shirley *et al.*, 83; Plumlee, 84].

The Ca intake is however important for the fertility and health of the mother, apart from the Ca requirement of the foetus. Thus rats on a low Ca intake were either infertile or ate their young, or the young died *in utero* [Toverud, 85; Macomber, 86]. Davidson [87] found with pigs that a low Ca diet greatly increased the number of piglets born dead. This is probably not a specific Ca effect, since any type of inadequate diet would presumably interfere with fertility and pregnancy.

*Lactation.* From the point of view of Ca metabolism, this imposes a severe strain on the mother. Most experimenters with rats have found them consistently in negative Ca balance. Sherman and Quinn [88] found that Ca and P were lost together in lactation and subsequently regained in the same ratio, suggesting that they were both mobilized from bone. Direct analysis by Warnock and Duckworth [89] showed that the 'ends' and cancellous parts of the bones contributed this Ca, the shafts being unaffected. Goss and Schmidt [78] found that the largest litters caused the greatest losses, as much as 20 per cent. of the body Ca being removed from the mother rat. They noted that while the animals appeared to get enough Ca in the diet, they were unable to absorb it fast enough. Kletzien *et al.* [90] found that with a diet containing 0.6 per cent. Ca, the blood Ca was lowered, and suggested that mobilization was unable to keep up with the demands of the mammary glands.

Cows can apparently be kept in Ca equilibrium during lactation if the intake is enough [91, 92], but as a rule, even with apparently adequate supplies, they are in negative Ca balance [93, 94]. Both Forbes *et al.* [80] and Huffman *et al.* [92] found that cows which were in negative Ca balance early in lactation, or during the height of lactation, could make this up afterwards, by retaining Ca again. Forbes *et al.* considered



The Ca of the bones was more available for milk than that of the diet and doubted whether supplementary diet was used at all for lactation purposes, though *Man et al.* showed that Ca utilization was more efficient during heavy milking.

Work on lactating women has shown that these findings also apply to them. Donelson *et al.* [95] and Hummel *et al.* [96] reported negative balances during lactation, but if the Ca intake is adequate, Ca can be retained [97], the retention being greatest with the greatest milk production. In this report the intakes were from 1.92 to 2.18 g. per day. Shukers *et al.* [76] found that lactating women, allowed to choose their own food, raised their Ca intake 62 to 88 per cent. over that during pregnancy, the daily intakes being from 2.03 to 5.3 g., far more than usually taken. Hunscher [98] investigated three lactating women and found that in spite of large intakes the activity of the Ca balance increased after 6 months' high milk production, the Ca being lost mainly in the faeces, and to such an extent that the faecal loss sometimes exceeded the whole intake. Here, too, the bone Ca was presumably more available than the food Ca. In late lactation, when less milk was secreted, Ca was again stored in the

These facts illustrate the remarkable elasticity of the body in dealing with emergencies in Ca metabolism. During periods of high demand, large amounts can be given up, to be regained when the need falls. But presumably the endogenous loss is not reduced. Whether it is physiological for such large losses to be sustained is another question. It must happen in the majority of cases that provided the stores are well filled at the end of pregnancy, and provided further that they will be replenished after lactation has ended, no harm ensues. In the case of women starting off with low reserves, on the other hand, it is obvious that serious results may follow, as they probably have little opportunity of refilling their stores, especially if subsequent pregnancies follow rapidly. It is thus of paramount importance that extra Ca be included in the diet

of poorly nourished lactating women, and a wise precaution in the case of those better nourished.

## REFERENCES

1. SMITH, A. H. and SMITH, P. K. *J. biol. Chem.*, **107**, 681 (1934)
2. BENEDICT, F. G. *Carn. Inst. Wash. Pub.*, **203**, 1 (1915)
3. ELLIS, M. and MITCHELL, H. H. *Amer. J. Physiol.*, **104**, 1 (1933)
4. HAMILTON, B. *Acta Paediat.*, **2**, 1 (1922-3)
5. BAUER, W., ALBRIGHT, F. and AUB, J. *J. clin. Invest.*, **7**, 75 (1929)
6. PUGSLEY, L. I. *Biochem. J.*, **30**, 1271 (1936)
7. BELL, G. H., CUTHBERTSON, D. P. and ORR, J. *J. Physiol.*, **100**, 298 (1941)
8. FAIRBANKS, B. W. and MITCHELL, H. H. *J. Nutr.*, **11**, 551 (1936)
9. KINSMAN, G., SHELDON, D., JENSEN, E., BERNDT, M., outhouse, J. and MITCHELL, H. H. *J. Nutr.*, **17**, 429 (1939)
10. MITCHELL, H. H. and MCCLURE, F. J. *Bull. nat. Res. Counc.*, **99**, 51 (1937)
11. MITCHELL, H. H. and CURZON, E. G. *The dietary requirements of calcium and its significance*. Paris: Hermann et Cie (1939)
12. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **45**, 201 (1951)
13. NICOLAYSEN, R., EEG-LARSEN, N. and MALM, O. J. *Physiol. Rev.*, **33**, 424 (1953)
14. BLAU, M., SPENCER, H., SWERNOW, J. and LASZLO, D. *Science*, **120**, 1029 (1954)
15. VISEK, W. J., MONROE, R. A., SWANSON, E. W. and COMAR, C. L. *J. Nutr.*, **50**, 23 (1953)
16. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **17**, 253 (1939)

- OWEN, E. C. *Biochem. J.*, **33**, 22 (1939)
- OWEN, E. C., IRVING, J. T. and LYALL, A. *Acta med. Scand.*, **103**, 235 (1939)
- HOLMES, J. O. *Nutr. Abst. Rev.*, **14**, 597 (1945)
- ROTTENSTEN, K. V. *Biochem. J.*, **32**, 1285 (1938)
- CARLSSON, A. *Acta pharm. tox. Kbh.*, **7**, Supp. 1 (1951)
- KELLY, F. C. and HENDERSON, J. M. *J. Hyg., Camb.*, **29**, 418 (1929-30)
- KUNG, L. C. and YEH, H. L. *Chin. J. Physiol.*, **12**, 139 (1937)
- NICHOLLS, L. and NIMALASURIYA, A. *J. Nutr.*, **18**, 563 (1939)
- KRAUT, H. and WECKER, H. *Biochem. Z.*, **315**, 329 (1943); **318**, 495 (1948)
- WALKER, A. R. P., FOX, F. W. and IRVING, J. T. *Biochem. J.*, **42**, 452 (1948)
- HEGSTED, D. M., MOSCOSO, I. and COLLAZOS, CH. C. *J. Nutr.*, **46**, 181 (1952)
- MCCANCE, R. A. and WIDDOWSON, E. M. *J. Physiol.*, **101**, 44 (1942)
- WIDDOWSON, E. M. and THRUSSEL, L. A. Spec. Rep. Ser. med. Res. Counc. Lond., **275**, 296 (1951)
- MCCANCE, R. A. Trans. 5th Conf. metab. Interr. New York, Josiah Macy Jr. Found. p. 166 (1954)
- MCCANCE, R. A. and WALSHAM, C. M. *Brit. J. Nutr.*, **2**, 26 (1948)
- STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **21**, 577 (1941)
- MITCHELL, H. H. *J. Amer. dietet. Assoc.*, **20**, 511 (1944)
- SALVESEN, H. A. *J. biol. Chem.*, **56**, 443 (1923)
- MCCULLAGH, E. P. and MCCULLAGH, D. R. *J. Lab. clin. Med.*, **17**, 754 (1932)
- MILLS, R., BREITER, H., KEMPSTER, E., MCKEY, B., PICKENS, M. and OUTHOUSE, J. *J. Nutr.* **20**, 467 (1940)
- BERGEIM, O. *J. biol. Chem.*, **70**, 35 (1926)
- JONES, J. H. *J. biol. Chem.*, **142**, 557 (1942)

39. OUTHOUSE, J., SMITH, J., TWOMEY, I. and THORP, M.  
*J. Home Econ.*, **27**, 541 (1935)
40. OUTHOUSE, J., SMITH, J., MERRITT, L. and WHITE, F. R.  
*J. Nutr.*, **14**, 579 (1937)
41. MITCHELL, H. H., HAMILTON, T. S. and BEADLES, J. R.  
*J. Nutr.*, **14**, 435 (1937)
42. FRENCH, R. B. and COWGILL, G. R. *J. Nutr.*, **14**, 38  
(1937)
43. DUNCAN, D. L. *Nutr. Abst. Rev.*, **25**, 309 (1955)
44. FOURNIER, P. *C.R. Acad. Sci.*, **238**, 509 (1954)
45. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **31**, 40  
(1946)
46. GEISSBERGER, W. *Helv. med. acta.*, **18**, 461 (1951)
47. PATTON, M. B. and SUTTON, T. S. *J. Nutr.*, **48**, 443 (1955)
48. BREITER, H., MILLS, R., DWIGHT, J., MCKEY, B.  
ARMSTRONG, W. and OUTHOUSE, J. *J. Nutr.*, **21**, 35  
(1941)
49. VINTHER-PAULSEN, N. *Geriatrics*, **8**, 76 (1953)
50. COOKE, A. M. *Lancet*, **1**, 877, 929 (1955)
51. HOWARD, J. E., PARSON, W. and BIGHAM, R. S. *Bull.*  
*Johns Hopkins Hosp.*, **77**, 291 (1945)
52. ALBRIGHT, F., BLOOMBERG, E. and SMITH, P. H. *Trans.*  
*Ass. Amer. Physns.*, **55**, 298 (1940)
53. ALBRIGHT, F. *Recent Prog. Hormone Res.*, **1**, 293 (1944)
54. BABAIANTZ, L. *Radiol. clin. Basel*, **16**, 291 (1947)
55. ARON, H. *Biochem. Z.*, **12**, 28 (1908)
56. TELFER, S. V. *Glasgow Med. J.*, **114**, 10 (1930)
57. OUTHOUSE, J., KINSMAN, G., SHELDON, D., TWOMEY, I.  
SMITH, J. and MITCHELL, H. H. *J. Nutr.*, **17**, 199 (1937)
58. WATSON, E. K., MCGUIRE, E. W., MEYER, F. L. and  
HATHAWAY, M. L. *J. Nutr.*, **30**, 259 (1945)
59. DANIELS, A. L. *Amer. J. Dis. Child.*, **62**, 279 (1941)
60. JOHNSTON, F. A., SCHLAPHOFF, D. and MCMILLAN, J.  
*J. Nutr.*, **41**, 137 (1950)
61. MCLEAN, D. S., LEWIS, G. K., JENSEN, E., HATHAWAY,  
M., BREITER, H. and HOLMES, J. O. *J. Nutr.*, **31**, 12  
(1946)

- MITCHELL, H. H., HAMILTON, T. S., STEGGERDA, F. R.  
and BEAN, H. W. *J. biol. Chem.*, **158**, 625 (1945)
- SHERMAN, H. C. and HAWLEY, E. *J. biol. Chem.*, **53**, 375  
(1922)
- STEARNS, G. and MOORE, D. L. R. *Amer. J. Dis. Child.*,  
**42**, 774 (1931)
- HUNSHER, H. A., COPE, F., NOLL, A. and MACY, I. G.  
*J. biol. Chem.*, **100**, lv (1933)
- JEANS, P. C., STEARNS, G., MCKINLEY, J. B., GOFF, E. A.  
and STINGER, D. *J. Pediat.*, **8**, 403 (1936)
- MACY, I. G. *Nutrition and chemical growth in childhood*.  
Springfield, Ill. **1**, 15 (1942)
- MACY, I. G. *ibid.*, **3**, 1784-1829 (1951)
- JOHNSTON, J. A. *Amer. J. Dis. Child.*, **67**, 265 (1944)
- KEMPSTER, E., BREITER, H., MILLS, R., MCKEY, B.,  
BERNDS, M. and OUTHOUSE, J. *J. Nutr.*, **20**, 279 (1940)
- GOSS. Unpublished experiments quoted by SCHMIDT,  
C. L. A. and GREENBERG, D. M. *Physiol. Rev.*, **15**, 297  
(1935)
- SHUKERS, C. F., MACY, I. G., NIMS, B., DONELSON, E.  
and HUNSCHER, H. A. *J. Nutr.*, **5**, 127 (1932)
- ANDERSON, A. B. and BROWN, A. *Lancet*, **2**, 482 (1941)
- SPRAY, C. M. and WIDDOWSON, E. M. *Brit. J. Nutr.*, **4**,  
332 (1950)
- MACY, I. G., HUNSCHER, H. A., NIMS, B. and MCCOSH,  
S. S. *J. biol. Chem.*, **86**, 17 (1930)
- SHUKERS, C. F., MACY, I. G., DONELSON, E., NIMS, B.  
and HUNSCHER, H. A. *J. Nutr.*, **4**, 399 (1931)
- HOFFSTRÖM, K. A. *Skand. Arch. Physiol.*, **23**, 326 (1910)
- GOSS, H. and SCHMIDT, C. L. A. *J. biol. Chem.*, **86**, 417  
(1930)
- FORBES, E. B., SCHULZ, J. A., HUNT, C. A., WINTER, A. R.  
and REMLER, R. F. *J. biol. Chem.*, **52**, 281 (1922)
- LANDSBERG, E. Z. *Geburts. Gynäk.*, **76**, 53 (1915)
- HUMMEL, F. C., HUNSCHER, H. A., BATES, M. F.,  
BONNER, P., MACY, I. G. and JOHNSTON, J. A. *J. Nutr.*,  
**13**, 263 (1937)



82. COONS, C. M. and COONS, R. R. *J. Nutr.*, **10**, 289 (1935)
83. SHIRLEY, R. L., JETER, M. A., FEASTER, J. P., MCCALL, J. T., OUTLER, J. C. and DAVIS, J. K. *J. Nutr.*, **54**, 59 (1954)
84. PLUMLEE, M. P., HANSARD, S. L., COMAR, C. L. and BEESON, W. M. *Amer. J. Physiol.*, **171**, 678 (1952)
85. TOVERUD, G. *J. biol. Chem.*, **58**, 583 (1923-4)
86. MACOMBER, D. *J. Amer. med. Assoc.*, **88**, 6 (1927)
87. DAVIDSON, H. R. *J. agric. Sci.*, **20**, 233 (1930)
88. SHERMAN, H. C. and QUINN, E. J. *J. biol. Chem.*, **67**, 667 (1926)
89. WARNOCK, G. M. and DUCKWORTH, J. *Biochem J.*, **38**, 220 (1944)
90. KLETZIEN, S. W. F., TEMPLIN, V. M., STEENBOCK, H. and THOMAS, B. H. *J. biol. Chem.*, **97**, 265 (1932)
91. HART, E. B., STEENBOCK, H., SCOTT, H. and HUMPHREY, G. C. *J. biol. Chem.*, **71**, 263 (1926-7)
92. HUFFMAN, C. F., ROBINSON, C. S. and WINTER, O. B. *J. Dairy Sci.*, **13**, 432 (1930)
93. HART, E. B., STEENBOCK, H., HOPPERT, C. A., BETHKE, R. M. and HUMPHREY, G. C. *J. biol. Chem.*, **54**, 75 (1922)
94. TURNER, W. A. and HARTMAN, A. M. *J. Nutr.*, **1**, 445 (1928-9)
95. DONELSON, E., NIMS, B., HUNSCHER, H. A. and MACY, I. G. *J. biol. Chem.*, **91**, 675 (1931)
96. HUMMEL, F. C., STERNBERGER, H. R., HUNSCHER, H. A. and MACY, I. G. *J. Nutr.*, **11**, 235 (1936)
97. OBERST, F. W. and PLASS, E. D. *Amer. J. Obst. Gyn.*, **40**, 399 (1940)
98. HUNSCHER, H. A. *J. biol. Chem.*, **86**, 37 (1930)

## CHAPTER VIII

# The Dietary Requirement of Calcium

THE calcium requirements of man have been the subject of a great deal of work and thought, and the subject has been recently discussed by Stearns [1]. The older investigators, who were ignorant of many of the peculiarities of Ca metabolism, arrived at figures which were in many cases incorrect. Sherman [2], studying balance figures from the literature and from his own observations, supposed that if the subject was in negative balance, the addition of the Ca lost would bring him to equilibrium, and if the Ca balance was positive, the requirement was equal to the intake minus the Ca stored. He was unaware that only part of the ingested Ca was retained, and since most of his subjects were in negative balance, the requirement figure he arrived at was too low.

It has now been realized that, unless very carefully controlled, the results of balance experiments can be misleading. In fact, Kraut and Wecker [3] regret there is not some method independent of metabolic experiments which could be used instead. In the case of children a new method has been suggested which will be described below.

Official organizations dealing with research into nutrition have now published Tables of recommended dietary standards, including Ca. The first to attempt this was the League of Nations Health Organization, which issued a report in 1935 [4]. The chief bodies which have since then recommended standards are the Food and Nutrition Board of the National Research Council of the U.S.A. [5]; the Committee on Nutrition of the British Medical Association [6]; the Canadian Council on Nutrition [7]; and the Publicity Office of the

Dutch Nutrition Council [8]. The Nutrition Council of South Africa have also recently published a Table of dietary standards [9].

The rationale of these various recommendations differs and makes comparison difficult. The resulting limitations have been well discussed by Pett [10].

The following quoted from the National Research Council Board's report published in 1953 [5] gives a good summary of the positions adopted by the various bodies. With respect to its own figures, the following is stated:

The recommendations are not intended to provide a basis for judging the nutritional status of population groups but rather to serve as a guide for planning food supplies for these groups. If these allowances are used in dietary evaluation, it is essential to appreciate that, while most persons whose consumption equals or exceeds the goal are presumably adequately nourished, not all persons who fail to reach these goals are malnourished.

They then go on to state:

The dietary standards proposed by the Committee on Nutrition of the British Medical Association are somewhat similar in philosophy and objective to the Recommended Dietary Allowances (of the National Research Council Board), but the caloric estimates and quantities of various nutrients recommended 'are believed to be sufficient to establish and maintain a good nutritional state in *representative individuals* of population groups'. The British standard thus differs in providing values which apply to an average person rather than values which will cover essentially all individuals, recognizing, however, that some members of every group may need more than the average.

The dietary standard proposed by the Canadian Council on Nutrition differs considerably in purpose and philosophy from the American and British recommendations. The Canadian standard represents 'a nutritional floor bene

which maintenance of the health of the people cannot be assumed'. It is designed not only as a basis for planning food supplies for individuals or groups, but also for use in assessing the amount of each nutrient provided by a diet for an individual or group in terms of *probable physiologic requirement*. Recommendations are based upon body size, and the influence of work on requirements is emphasized.

In view of the differences in underlying philosophy and purpose between the Canadian, British and American standards, it is obvious that specific recommendations will differ. Values proposed in the Canadian standard approach minimal requirements; allowances in the British standard are for the maintenance of good nutrition in the average person; allowances in the United States standard are recommended for the maintenance of good nutrition in substantially all normal persons.

The National Nutrition Council of South Africa [9] state in this connexion:

The standards proposed here should be regarded as adequate for the maintenance of health without allowing for a safety margin for ill health, or for great individual differences in absorption and metabolism.

The recommendations of these bodies for Ca are summarized in Tables I and II.

TABLE I  
RECOMMENDED Ca STANDARDS FOR ADULTS  
G. per day

	U.S.A.	B.M.A.	Canada	Dutch	South Africa
Man	0.8	0.8	0.75	1.0	0.7
Woman	0.8	0.8	0.6	1.0	0.6
Pregnancy	1.5 (3rd trimester)	1.5 (last half)	1.6 (last half)	2.0	1.5 (3rd trimester)
Lactation	2.0 (850 ml. milk per day)	2.0	1.6	2.0	0.12 more for each 100 ml. milk per day

TABLE II  
RECOMMENDED Ca STANDARDS FOR CHILDREN  
G. per day

*Infants*

	U.S.A.	B.M.A.	Canada	Dutch	South Africa		
Months							
1-3	0.6	} 1.0	—	} 0.8	} 0.8		
4-9	0.8		—				
10-12	1.0		—				
Years		Children					
1-3	} 1.0	} 1.0	} 1.0	} 0.8	0.6		
4-6							0.7
7-9							0.8
10	} 1.2	} B 1.3 G 1.5	} B 1.0, G 1.5	} 1.2	0.8		
11							B 0.8, G 1.0
12							
13	} B 1.4 G 1.3	} B 1.3, G 1.2	} 1.5				
14							
15							
16	} B 1.4 G 1.3	} B 1.4 G 1.1	} — — —	} 1.2	B 1.3		
17							G 1.2
18							
19							
20							

**Ca requirements of adults**

It will be noted that there is a certain variation mostly line with the philosophies of the various bodies. The figures have been arrived at mainly as a result of estimates of the total food consumption of the population, information derived from surveys, and balance experiments. The B.M.A. committee stated that they 'found no little difficulty in reaching decision regarding the desirable intake of calcium', largely owing to the ability of the body to adapt to lower intake. The N.R.C. of the U.S.A. had recommended 1.0 g. per day.



for adults in their 1948 publication, which the B.M.A. committee thought unjustified, and in the latest Table the standard was lowered to 0.8 g. The question of a 'margin of safety', raised originally by Sherman, does not appear to be now considered.

In men and non-pregnant and non-lactating women a figure of 10 mg. per kg. body-weight per day seems to the present writer to be adequate, and the South African figures were based on an average man weighing 73 kg. and woman of 59 kg. There is much evidence to support this point of view. Leitch [11] selected from the literature a large number of balance data on healthy women both in positive and negative balance. On plotting intake against output, the level of Ca intake above which losses and gains of Ca were equal was 550 mg. per day. This worked out at almost 10 mg. per kg. body-weight. Mitchell and Curzon [12] selected 139 observations from the literature on 107 subjects, of whom 18 were men. These results were plotted with respect to intake and output and a straight line fitted to the data. This line intersected the diagonal at 9.75 mg. per kg. body-weight per day, a figure which agrees well with that of Leitch obtained by an analogous method.

In a study of nine normal male subjects [Steggerda and Mitchell, 13], when the availability of milk Ca was being investigated, an average Ca requirement of 9.55 mg. per kg. body-weight per day was arrived at. In 1946 Steggerda and Mitchell [14] did balance experiments on 19 men, the diet periods being 20 days and 75 such periods being considered. In both these studies the subjects were first on a basal diet of very low Ca content (about 200-250 mg. per day); the Ca level was then increased by the addition of various milk products. From these figures they could calculate the percentage utilization of the extra Ca by the method of Kinsman *et al.* [15]. The negative balance was then divided by the percentage utilization and multiplied by 100, which gave the amount of Ca needed to wipe out the lost Ca, and on being added to the intake of Ca gave the total requirement. Thus in a subject with a utilization of 32 per cent., the total intake was 458 mg.

and the balance — 57 mg. This loss would be abolished by the addition to the diet of  $\frac{57}{0.32} = 178$  mg.:  $178 + 458 = 636$  mg.

which is considered the total daily Ca required. Averaging the results, the daily Ca requirement came to 9.21 mg. per kg. body-weight. In a later paper from the same laboratory Bricker *et al.* [16] found, working with 8 women, that the requirement was 11.8 mg. per kg. body-weight per day.

In their 1946 paper Steggerda and Mitchell [14] summarized their present and earlier findings, certain unpublished data and also calculations from the work of Breiter *et al.* [17] who conducted similar balance experiments. The mean of the daily Ca requirement per kilogram of body-weight varied from 9.21 to 11.61 mg., and averaged for 43 individuals 9.99 mg. Taken in conjunction with the suggestions, using different methods, by Leitch of 10.0 mg., and Mitchell and Curzon of 9.75 mg., it must be agreed that the correspondence is quite remarkable. In view of the findings of Steggerda and Mitchell, Walker *et al.* [18] adjusted the Ca intake of three of their subjects to a level of 10 mg. per kg. per day and found that they either were in equilibrium on this intake or else adapted to it without difficulty.

Steggerda and Mitchell [14] recommend a daily intake of 10 mg. per kg. body-weight per day, provided that an average proportion of Ca comes from dairy products. They do not feel that a 'margin of safety' is needed. In the case of the individual who has been accustomed to more, the adaptive mechanism will come into effect and, judging from the experiments quoted elsewhere, will cause no physiological disturbance at this level of intake.

The figures of the N.R.C. of the U.S.A., and the B.M.A. and those for Canada and South Africa are fairly closely in accord with the information available in the literature, but the present writer feels that the Dutch recommendations are unnecessarily high. No harm of course comes from this high consumption of Ca, but from an economic point of view when the supplies may be limited, and especially when the

a great deficiency in other parts of the world, unnecessarily high standards should be avoided. It should be recalled that Hegsted and his colleagues [19] found perfectly healthy people in equilibrium on intakes of 100 and 200 mg. per day, and stated, as has been emphasized above, that all considerations of Ca requirements are really studies of the previous Ca intake, and that Ca deficiency is unlikely in adults (except women in the child-bearing age) on most natural diets. Thus the reservation must be made that the recommended allowances of Ca are those for people accustomed to eating Ca at that level. If our dietary habits changed, it is possible that the Ca requirement would too.

*Pregnancy.* As has been stated above, pregnancy does not involve a great strain on the Ca metabolism of a woman previously adequately nourished in this regard and it is easy, knowing the Ca content of the foetus, to calculate the extra requirement.

According to figures published by Mitchell and Curzon [12], Ca accretion on any scale does not begin till the 7th month and most standards are applied only to the last half of pregnancy or to the last trimester. The Ca content of the foetus at term is usually about 25 g., the other products of conception being virtually Ca-free. If extra Ca is given over the last trimester (of 93 days) 270 mg. per day must be available to the foetus. Presupposing utilizations of 25 or 30 per cent., 1,000 or 800 mg. respectively must be added to the maternal diet. The figures recommended by Canada, Holland and South Africa allow extra Ca at these levels, but those for the U.S.A. and Great Britain are less. It must be admitted that the utilization of the extra Ca is not accurately known, and it is possible that it is more efficient in pregnancy. With a total body Ca of over a kilogram, it might be thought that the loss of a few grams would be immaterial. But with the strain of lactation coming soon, no drain of the body Ca stores should be allowed at this time.

*Lactation.* As with pregnancy, it should not be difficult to arrive at figures for the Ca supply to the mother's milk, which

on an average contains 0.03 per cent. The U.S.A. figures allow 1.2 g. extra Ca, supposing that 850 ml. of milk are produced daily. This milk would contain 255 mg. Ca and thus about 20 per cent. utilization of the extra Ca has been allowed for. The B.M.A., Canadian and Dutch figures allow 1 to 1.2 g. extra Ca per day and the South African tables state that 0.12 g. extra Ca should be taken for each 100 ml. of milk produced, the utilization being placed at 25 per cent. Whether these figures are mathematically correct, it is not infrequently found during lactation that bone Ca is used in preference to that from the food. This difficulty cannot be overcome, and the best is to give the mother the extra Ca she should get, some or probably a great deal of which she will use in lactation and with a proper diet after weaning any losses will soon be made good. If she had no extra Ca at all and produced 1,000 ml. of milk per day for 9 months (a peak output), she would lose altogether about 80 g. of Ca or about 8 per cent. of her stores.

### Ca requirements of children

*Infants.* This is probably the most difficult of all estimates to arrive at, mainly because the degree of utilization of the Ca of milk is not accurately known. It might be expected that some idea could be got from the consumption of mother's milk, a daily intake of 1,000 ml., which is a top figure, giving 300 mg., which presumably is about the maximum a baby needs till it is weaned.

Turning, on the other hand, to balance experiments and investigations such as those of Holmes [20], one finds that the calculated daily Ca retention of a baby in the first month is as high as 600 mg. There is some discrepancy here, since even if the baby's retention was 100 per cent. [and it is probably more than 60 per cent., Telfer, 21] it could not obtain this intake without imbibing 2 litres of human milk, and more if the retention was less. Even at 6 months the breast-fed baby would not be getting enough Ca. Most of the recommendations for Ca for infants in Table II pertain to cow's milk, p



ably on the supposition that the retention of breast milk is much higher than that from cow's milk. Even so, these figures are not very realistic. To get 1 g. of Ca, a baby would have to drink over 800 ml. of unmodified cow's milk. In a study of Ca requirements by the present writer published in 1950 [22] he stated that he did 'not feel competent, in the state of present knowledge . . . to recommend any figures for the first period of life'. His feeling is unchanged. Only accurate balance work can solve this problem.

*Children.* The figures in Table II show a general agreement that the Ca intake should be about 0.8 to 1.0 g. per day in extreme youth, rising to 1.2 to 1.5 g. during the period of adolescent growth. While these recommendations are undoubtedly adequate, some of them have a 'penny in the slot' look which would indicate that not a great deal of thought had been given to them as average or maintenance allowances. Thus it is hardly possible that the Ca requirement, which is to supply Ca for growth, would be the same between the ages of 1 and 11, or between 10 and 20.

To study the matter more objectively, the present writer [22] made calculations for boys based on the calculated Ca increments published by Mitchell *et al.* [23]. This method was one proposed by Terroine [24], who used it for determining the nitrogen requirements during growth. The information necessary is the daily Ca accretion in the body, and the degree of retention of the dietary Ca. The latter figure is the hardest to arrive at, and was placed arbitrarily at 25 per cent. for 1-2 years and 20 per cent. for the rest of the growth period. As pointed out earlier, the retained Ca is all used for growth and calcification processes, and no maintenance requirement has to be considered. The fraction

$$\frac{\text{Daily Ca accretion} \times 100}{\text{Percentage retention}}$$

gave the daily requirement. The figures arrived at were far lower than those recommended by any official body, being 0.65 g. for 1-2 years, 0.4 g. up to 7 years, then gradually rising



to a top figure of 1.3 g. for ages 14 and 15, after which the allowance gradually fell to the adult one of 0.7 g. when maintenance requirement appeared. The present writer still feels that these figures are a better representation of the Ca needs during growth. The South African Nutrition Council followed in general the arguments used by Irving, and their recommendations are mostly lower than those of the other bodies. When reliable figures, based on analyses of normal children, exist, this method will be the natural way of determining the Ca needed during growth.

### REFERENCES

1. STEARNS, G. J. *Amer. med. Assoc.*, **142**, 478 (1950)
2. SHERMAN, H. C. *J. biol. Chem.*, **44**, 21 (1920)
3. KRAUT, H. and WECKER, H. *Biochem. Z.*, **315**, 329 (1943)
4. League of Nations Health Organization. *Report on the physiological bases of nutrition by the Technical Commission appointed by the Health Committee*. Off. No. C.H. 1197 (1935)
5. National Academy of Sciences—National Research Council. Food and Nutrition Board. Publication 302 (1953)
6. *Report of the Committee on Nutrition*. British Medical Association, London (1950)
7. *Canad. Bull. Nutr.*, **3**, No. 2 (1953)
8. Nederlandse Voedingsmiddelen Tabel. Voorlichting bureau van de Voedingsraad (1951)
9. National Nutrition Council of South Africa. *Recommended minimum daily dietary standards*. *S. Afr. med. J.*, **30**, 10 (1956).
10. PETT, L. B. *J. Amer. dietet. Assoc.*, **27**, 28 (1951)
11. LEITCH, I. *Nutr. Abst. Rev.*, **6**, 553 (1936-7)
12. MITCHELL, H. H. and CURZON, E. G. *The dietary requirement of calcium and its significance*. Paris: Hermann & Cie (1939)
13. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **2**, 577 (1941)

5. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **31**, 407 (1946)
6. KINSMAN, G., SHELDON, D., JENSEN, E., BERNDT, M., OUTHOUSE, J. and MITCHELL, H. H. *J. Nutr.*, **17**, 429 (1939)
7. BRICKER, M. L., SMITH, J. M., HAMILTON, T. S. and MITCHELL, H. H. *J. Nutr.*, **39**, 445 (1949)
8. BREITER, H., MILLS, R., DWIGHT, J., MCKEY, B., ARMSTRONG, W. and OUTHOUSE, J. *J. Nutr.*, **21**, 351 (1941)
9. WALKER, A. R. P., FOX, F. W. and IRVING, J. T. *Biochem. J.*, **42**, 452 (1948)
10. HEGSTED, D. M., MOSCOSO, I. and CALLAZOS, CH. C. *J. Nutr.*, **46**, 181 (1952)
11. HOLMES, J. O. *Nutr. Abst. Rev.*, **14**, 597 (1945)
12. TELFER, S. V. *Glasgow med. J.*, **114**, 10 (1930)
13. IRVING, J. T. *S. Afr. med. J.*, **24**, 601 (1950)
14. MITCHELL, H. H., HAMILTON, T. S., STEGGERDA, F. R. and BEAN, H. W. *J. biol. Chem.*, **158**, 625 (1945)
15. TERROINE, E. F. *Quart. Bull. Hlth. Org., L.O.N.*, **5**, 427 (1936)

## CHAPTER IX

# The Calcium of the Blood

THE Ca content of blood in man is remarkably constant under physiological conditions and over all ages. It is virtually constant present in the plasma (or serum, where it is usually estimated) at a level of 9 to 11 mg. per 100 ml. This constancy is seen also in many other animals, but in the rabbit the plasma level is subject to great variations, rising as high as 19 mg. under abnormal normal conditions. The blood-cell of the ox, sheep and rabbit, unlike those of man and pig, contain variable amounts of Ca.

Experimental raising of the blood Ca to high figures is usually fatal. When the Ca falls in amount, the condition tetany supervenes when the level has reached about 6 mg. per 100 ml. This is characterized by typical muscle spasms, usually attributed to increased sensitivity of the neuro-muscular junction, though Greenberg *et al.* [1] think that a part of the central nervous system higher than the spinal cord is involved.

The Ca in the plasma is present in two forms, ionized and non-ionized, diffusible, and non-diffusible. Benjamin and Hess [2] suggested that the Ca was present in four fractions, but this concept has not been generally accepted.\*

The determination of the proportions of ionized and non-ionized diffusible Ca presents some difficulties; if simple dialysis is carried out, all the Ca will be removed from serum, since the two fractions are in equilibrium. A procedure known as compensation dialysis can be used instead, in which serum is dialysed against a number of solutions of different Ca content. The solution in which no change of Ca concentration occurs contains Ca ions at the same level as the serum. With this method, about 60 per cent. of the Ca of human serum is ionized.

\* There is a very small diffusible fraction of unionized Ca complexes in plasma.

und to be diffusible. Greene and Power [3], using the method of vividiffusion, in which the entire blood of a living animal is dialysed against a small volume of fluid, reported on an average 62 per cent. of the Ca to be diffusible. Ultrafiltration under pressure through a collodion membrane gives figures of much the same order [Greenberg and Larson, 4]. McLean and Hastings [5] made use of the sensitivity of the frog heart to Ca ions and found about half the total serum Ca to be diffusible and nearly all of this in an ionized state. Calculations based on the composition of cerebrospinal fluid, edema fluid, ascites or pleural effusions indicate that the diffusible Ca of normal serum is about 5 mg. per 100 ml. It thus appears fair to conclude from all these methods that this fraction lies between 5 and 7 mg. per 100 ml.

Turning to the nature of the non-diffusible fraction, there seems hardly any need to consider the older views, since all evidence now points to this being a Ca-protein complex. McLean and Hastings [5] showed beyond doubt that a relationship exists between the Ca ions and the protein of plasma and have calculated mass-action equations and an ionization constant. They have constructed a nomogram from which, knowing the values of total protein and total Ca, the Ca ion concentration can be derived. The mass-action considerations involved have been shown to be true, as regards the proportion of ionized and unionized Ca, over a wide range of values as in hypo- and hyperparathyroidism, provided the protein content of the plasma does not change.

The ability of blood protein to bind Ca has been demonstrated by a number of workers. McLean and Hastings [6] first attempted a quantitative study with purified albumins prepared from the sera of horses and oxen. Martin and Perkins [7] found the serum albumin of horse and ox to bind about 11 mg. per g.N. and in humans the corresponding figure was 4.7 to 4.8 mg. Repeated recrystallization could alter this binding power. In a subsequent paper [8] the binding power of human albumin was found to be very constant at 4.7 mg. or 0.9 mg. per g.N. over ages from 2 to 91, and the same in



both sexes. In hyperparathyroidism the combining power increased. Carr [9] reported that bovine albumin took a maximum number of 8 Ca ions per molecule of protein at pH of 7. With increased pH the binding power rose, and with decreased pH. The binding of Ca in this way must be of a loose type, since as stated above, all the Ca can be dialysed out of serum. Armstrong *et al.* [10] investigated *in vivo* using  $\text{Ca}^{45}$ , and reported that the protein binding did not impede the transcapillary movement of Ca and that the redistribution of Ca between the ionized and bound fractions was rapid. The transcapillary movement of  $\text{Ca}^{45}$  and  $\text{Ca}^{40}$  were the same.

Hopkins *et al.* [11] studied this problem in a different way. They added Ca to normal serum and found that as the concentration was increased up to 19 mg. per 100 ml., the ionized and bound Ca fractions rose together, in an extraordinarily fixed proportion, the serum having the capacity to 'carry' more than it normally did. The P up to this point was unchanged. When a figure of 20 mg. per 100 ml. was reached, the concentrations of filtrable Ca and of P fell, possibly due to the formation of a supersaturated solution. An important point made by Howard [12] was that there was no reciprocal relationship between Ca and P concentrations in the serum *in vivo*, and thus any such changes seen *in vivo* were the result of physiological mechanisms. This has been shown by McCluskey and Hinrichs, and Gersh [13], who found that the injection of phosphates caused the formation of a colloidal Ca phosphate complex in the blood, which was rapidly removed by the liver and spleen.

Another aspect of the matter has been studied, viz. blood-changes in hens during the egg-laying period. At this time both the total [14] and non-diffusible Ca in the plasma rise [15]. Clegg and Hein [16] found that after giving diethylstilboestrol the rise in non-diffusible Ca in chick serum was paralleled by a rise in concentration of two of the electrophoretic components, and in a later paper [17], after similar treatment found a 'new' protein which was rich in P.



identified with  $P^{32}$ . When the Ca in the serum was raised, the electrophoretic mobility of this protein was markedly reduced, but that of the others was not, and this was attributed to combination with Ca.

*Factors affecting the blood Ca level.* Starvation usually has little effect upon this. But Ca deficiency will in the end cause the blood Ca to fall. Greenberg and Miller [18] found that a Ca-deficient diet fed to growing rats caused a fall in the blood figure from an original level of 10–12.5 mg. per 100 ml. to one of 7 mg. after 40 days and to 6.1 at 68 days. By this time the body Ca stores had fallen to less than half and the authors considered that a significant reduction in the Ca stores must take place before the blood Ca is reduced. Interestingly enough, tetany was not seen, nor did Irving [19] observe it in rats on low Ca diets and with serum Ca values of 5 mg. The injection of NaF, which was carried out for another purpose, into such rats immediately precipitated violent tetanic seizures. Presumably the ionized Ca is not lowered to the same extent as the total Ca.

In animals with well-marked low P rickets, as caused by the Steenbock-Black diet, the blood Ca level is usually a high normal and the blood inorganic P greatly lowered. When such animals are starved, the blood inorganic P rises rapidly, due to mobilization of P from soft tissues, and the Ca simultaneously falls at a fast rate, the animals looking miserable and being liable to tetany [20]. These changes are presumably due to the formation of a colloidal Ca phosphate complex and its removal from the blood as described above. When animals on a low Ca diet are starved, the blood Ca remains at a constant low level for as long as 10 days [19], and is presumably kept depressed by the continual mobilization of phosphate. A rapid and fatal hypocalcaemia can be induced by the injection of ethylene diamine tetracetic acid (versene) [Popovici *et al.*, 21].

*Vitamins.* The only vitamin with profound effects upon the blood Ca is vitamin D and associated compounds. McLean [22] held that there were two actions of vitamin D, antirachitic

and calcaemic. The antirachitic effect is self-explanatory and caused by a low-dosage level, capable of curing rickets. According to the older observations of Kramer *et al.* [20], the  $\text{Ca} \times \text{P}$  content of the blood had to rise, under the action of vitamin D or other curative agents, to a certain height before calcification of rachitic cartilage occurred. The rationale of this concept, which though empirical has been most helpful, is explained elsewhere (see p. 117). This antirachitic dose level of vitamin D has no effect upon the blood Ca level of normal animals or humans, but raises the blood P in animals suffering from low P rickets, and the Ca in those with low Ca rickets largely due to its action on intestinal absorption and possibly also to its effects upon renal function. Vitamins D<sub>2</sub> and D<sub>3</sub> are equally effective in this respect in mammals.

The calcaemic effect of vitamin D has been used chiefly in the treatment of tetany and very large doses of the vitamin are used. Here the effect is caused by the destructive action upon bone [McLean, 22]. Other substances allied to vitamin D have been used, especially dihydrotachysterol or A.T.10. McChesney and Messer [23] found, using massive doses given to dogs, that D<sub>3</sub> caused a longer lasting hypercalcaemia than D<sub>2</sub>, taking up to 26 days to subside, and A.T.10 produced a quick rise and fall in blood Ca. Parathyroid extract has a very quick action, the hypercalcaemia being over in 36 hours [Collip and Clark, 24]. McLean considers that the effect of A.T.10 is, like that of vitamin D in large doses, directly upon bone. In addition it has marked effects upon kidney function, decreasing P reabsorption in the tubules [Harrison and Harrison, 25] and thus secondarily affecting bone, mobilizing bone salts and thus raising the blood Ca. A.T.10 can probably increase intestinal Ca absorption in the same way that vitamin D does, but Albright *et al.* [26] consider that any antirachitic action is largely overshadowed by its marked action on renal excretion. In contrast, parathyroid extract, which also has a calcaemic effect, has no action on intestinal Ca absorption and is therefore not antirachitic. Its effects elsewhere will be considered below.

It might be mentioned at this stage that suggestions have been made that vitamin D acts through the parathyroid glands. This was first suggested by Taylor *et al.* [27], who claimed that the toxic actions of vitamin D were not obtained after parathyroidectomy. This was not confirmed by Dale *et al.* [28]. Tweedy *et al.* [29] found it was difficult to raise the low blood Ca level after parathyroidectomy with vitamin D; but if the animals were put on to a high Ca : P ratio diet the blood Ca became higher and it was then possible to demonstrate the calcaemic action of the vitamin. Thus it appears there is no connexion between the effects of vitamin D and the parathyroid, though their actions are in some instances similar.

*The endocrine glands.* Some of these have profound effects upon the blood Ca level, but others have but little action. Thus insulin very probably increases the blood Ca, but secondarily to its effect of lowering the inorganic P level. The thyroid has a considerable effect upon Ca excretion and in thyrotoxicosis there is an increased loss of Ca. There seems to be no unanimity about the level of blood Ca in this condition. Albright *et al.* [30] considered it to be slightly raised, but Wade [31] found it to be lowered. Robertson [32] reported that it was lowered from an average figure of 10.39 to 9.71 mg. per 100 ml., the difference proving to be statistically significant. In myxoedema the level is unchanged.

*Oestrogens.* As stated above, the blood level is raised in birds during the egg-laying cycle, and in non-ovulating birds by the administration of oestradiol or diethylstilboestrol, this being accompanied by the formation of medullary bone (see p. 106). This was still observed in birds after parathyroidectomy or hypophysectomy, so that the action of oestrogens was not through these glands [Riddle and McDonald, 33].

*Pituitary.* The reports in the literature are somewhat conflicting. Anderson and Oastler [34] in well-controlled experiments with rats found that hypophysectomy had no effect upon the blood Ca level, and assumed that the existence of a parathyrotropic hormone in the pituitary was not proved, which is still the consensus of opinion. Since the effect of the



anterior lobe of the pituitary on bone is directed more toward matrix formation than calcification, it might be expected that the blood Ca level would not be affected.

*Adrenal glands.* Cortisone causes a greatly increased excretion of Ca in the urine and faeces, but in otherwise normal animals both cortisone and adrenocorticotrophic hormone cause little change in the blood Ca or, if anything, a slight decrease [Pincus *et al.*, 35]. If, however, the animal is nephrectomized, cortisone causes an increase of serum Ca to toxic levels [Grollman, 36]. Adrenalin administration is followed by a rise in serum Ca [Pincus *et al.*, 35].

*Parathyroid glands.* These glands have an important controlling influence upon Ca metabolism. They are sensitive to the blood Ca level, since there is an inverse relationship between this level and the volume of the gland [Stoerk and Carnes, 37]. An active extract was prepared in 1925, which could increase the blood Ca level and produce all the effects seen in hyperparathyroidism. The exact mode of action of this hormone is still undecided, there being two schools of thought: one that the effect is directly upon bone, the other that the action is on the kidney, bone being secondarily affected.

The original view was that the hormone acted directly upon bone, but it was soon challenged by Albright and Ellsworth [38], who considered that this effect was secondary to renal excretory changes, the tubular reabsorption of P being decreased, leading to increased P excretion. Since then much evidence has been produced supporting this concept. Harrison and Harrison [39] reported parathyroid hormone to decrease tubular P reabsorption, and Tweedy *et al.* [40] in particular found the urinary excretion of  $P^{32}$  in parathyroidectomized animals to be most sensitive to small doses of parathyroid extract and judged it must act directly on the kidney. Deane and Gordon [41] likewise found that the serum P of parathyroidectomized rats could be easily reduced by doses which hardly affected normal animals.

Talmage and his colleagues have done a considerable amount of work, and seem to subscribe to both views. St

ing the metabolism of both  $\text{Ca}^{45}$  and  $\text{P}^{32}$  they considered that the hormone acted primarily on bone, this effect being enhanced by an additional action on the kidney, whereby the excretion of P was increased [42]. They pointed out that parathyroid extract raised the blood P, even though the renal excretion of P was ten times the normal level, so that P was entering the blood faster than it could be removed and presumably must have come from bone.

In a subsequent paper [43] they stated that the renal excretion of P was markedly reduced one hour after parathyroidectomy and considered that one action of the gland was to control renal excretion of P, the renal thresholds being altered in such a way that that for P was increased, while that for Ca was diminished. The parathyroid kept the blood Ca at 11 mg. per 100 ml. serum, but in its absence the figure fell to 6-7 mg. [44]. They seem to feel that while renal phosphate excretion is not the primary means by which the parathyroids influence Ca and P metabolism, none the less in the rat they do directly modify the excretion of both elements.

The pure renal theory has lost a good deal of ground, and even Albright and his colleagues now agree that this is not the only action of the hormone [45], since they found evidence of osteoclastic resorption in bone of nephrectomized animals injected with parathyroid extract. Stoerk [46] reported that the parathyroid hormone could still cause hypercalcaemia and even osteitis fibrosa after nephrectomy. Monahan and Freeman [47] stated that the serum Ca was still depressed in parathyroidectomized dogs after nephrectomy. Other workers have produced similar evidence that parathyroid hypercalcaemia can still be obtained after nephrectomy [e.g. Grollman, 48] or that, under appropriate conditions, the renal effect on P excretion is absent [Hogben and Bollman, 49] or much lessened if the urinary P was reduced [Coxon, 50]. Munson [51] in a recent paper stated that the fall in serum Ca after parathyroidectomy was not dependent upon a prior rise in serum inorganic P and that injection of the hormone into parathyroidectomized rats raised both the serum Ca and P. He



concluded there was a direct action of the parathyroid upon skeletal tissue independent of whatever effect it might have on the kidney tubule. Both Barnicot [52] and Chang [53] have reported that when fragments of bone and of parathyroid gland are grafted together subperiosteally or intracerebrally, bone resorption with an osteoclastic response occurs, the effect being specific and not caused by other glands and tissues tested.

Munson raised the interesting suggestion, for which there is increasing evidence, that there are two hormones in parathyroid extract, Ca mobilizing and phosphaturic. Handler *et al.* [54] found different effects if the hormone was given subcutaneously or intravenously, and Handler and Cohn [55] suggested from their findings that there were both hypercalcaemic and haemodynamic factors, the latter increasing the renal plasma flow and glomerular filtration rate, the former factor not being essential for hypercalcaemia. Stewart and Bowen [56], using the Tepperman technique for the preparation of parathyroid extract [57], got a substance from the thymus and spleen which stimulated P excretion by the kidney and considered the phosphaturic effect of parathyroid extract to be an artefact resulting from the extraction process, since formalin treatment caused a loss of the hypercalcaemic power but not of the renal factor. Davies and Gordon [41] found two factors in parathyroid extract, a dialysable one affecting the renal P excretion, and a non-dialysable one which affected both urinary P and also the serum Ca; more of the latter fraction had to be given to obtain the serum Ca action. Kenyon *et al.* [58] reported that there were discrepancies in the ratio of hypercalcaemic to phosphaturic activities of different parathyroid preparations.

It thus seems clear from this, that the disagreement in the literature on the mechanism of parathyroid action may well be due to the existence of two factors and the different proportions of these in different preparations. None the less, the present writer feels, on the balance of the evidence, that the primary action of the gland is on bone, and the bone changes are not secondary to alterations in renal P excretion.

Parathyroidectomized animals will develop rickets like unoperated controls and their cartilage will calcify *in vitro* [Heimer *et al.*, 59] and the evolution of hypervitaminosis A in rats is unaffected by parathyroidectomy [Cohen *et al.*, 60]. As stated above, parathyroidectomy does not prevent the hypercalcaemia caused by oestrogens [33].

## REFERENCES

1. GREENBERG, D. M., BOELTER, M. D. D. and KNOPF, B. W. *Amer. J. Physiol.*, **137**, 459 (1942)
2. BENJAMIN, H. R. and HESS, A. F. *J. biol. Chem.*, **100**, 27 (1933)
3. GREENE, C. H. and POWER, M. H. *J. biol. Chem.*, **91**, 183 (1931)
4. GREENBERG, D. M. and LARSON, C. E. *J. biol. Chem.*, **109**, 105 (1935)
5. MCLEAN, F. C. and HASTINGS, A. B. *Amer. J. med. Sci.*, **189**, 601 (1935)
6. MCLEAN, F. C. and HASTINGS, A. B. *J. biol. Chem.*, **108**, 285 (1935)
7. MARTIN, N. H. and PERKINS, D. J. *Biochem. J.*, **47**, 323 (1950)
8. MARTIN, N. H. and PERKINS, D. J. *Biochem. J.*, **54**, 643 (1953)
9. CARR, C. W. *Arch. Biochem. Biophys.*, **43**, 147 (1953)
10. ARMSTRONG, W. D., JOHNSON, J. A., SINGER, L., LIENKE, R. I. and PREMER, M. L. *Amer. J. Physiol.*, **171**, 641 (1952)
11. HOPKINS, T., HOWARD, J. E. and EISENBERG, H. *Bull. Johns Hopkins Hosp.*, **91**, 1 (1952)
12. HOWARD, J. E. Trans. 5th Conf. metab. Interr. New York, Josiah Macy Jr. Found., p. 11 (1953)
13. MCLEAN, F. C. and HINRICHS, M. A. *Amer. J. Physiol.*, **121**, 580 (1938)
- GERSH, I. *Amer. J. Physiol.*, **121**, 589 (1938)

14. RIDDLE, O. and REINHART, W. H. *Amer. J. Physiol.*, **660** (1926)
15. CORRELL, J. T. and HUGHES, J. S. *J. biol. Chem.*, **1** 511 (1933)
16. CLEGG, R. E. and HEIN, R. E. *Poultry Sci.*, **32**, (1953)
17. ERICSON, A. T., CLEGG, R. E. and HEIN, R. E. *Scien* **122**, 199 (1955)
18. GREENBERG, D. M. and MILLER, W. D. *J. Nutr.*, **22** (1941)
19. IRVING, J. T. *J. Physiol.*, **105**, 16 (1946)
20. KRAMER, B., SHEAR, M. J. and SIEGEL, J. *J. biol. Che* **91**, 271 (1931)  
IRVING, J. T. and NIENABER, M. W. P. *J. dent. R* **25**, 327 (1946)
21. POPOVICI, A., GESHICKTER, C. F., REINOVSKY, A. and RUBIN, M. *Proc. Soc. exp. Biol. N.Y.*, **74**, 415 (1950)
22. MCLEAN, F. C. *J. Amer. med. Ass.*, **117**, 609 (1941)
23. MCCHESENEY, E. W. and MESSER, F. *Amer. J. Physiol.*, **1** 577 (1942)
24. COLLIP, J. B. and CLARK, E. P. *J. biol. Chem.*, **64**, (1925)
25. HARRISON, H. E. and HARRISON, H. C. *Amer. J. Physiol* **137**, 171 (1942)
26. ALBRIGHT, F., BLOOMBERG, E., DRAKE, T. and SULZ WITCH, H. W. *J. clin. Invest.*, **17**, 317 (1938)
27. TAYLOR, N. B., WELD, C. B., BRANNION, H. D. and KAY, H. D. *J. Canad. med. Ass.*, **24**, 763; **25**, 20 (1931)
28. DALE, H. H., MARBLE, A. and MARKS, H. P. *Proc. R Soc., B.*, **111**, 522 (1932)
29. TWEEDY, W. R., TEMPLETON, R. D., PATRAS, M. and MCJUNKIN, F. A. and MCNAMARA, E. W. *J. biol. Che* **128**, 407, (1939)
30. ALBRIGHT, F., BAUER, W. and AUB, J. C. *J. clin. Inve* **10**, 187 (1931)
31. WADE, P. A. *Amer. J. med. Sci.*, **177**, 790 (1929)
32. ROBERTSON, J. D. *Lancet*, **2**, 129 (1941)

3. RIDDLE, O. and McDONALD, M. R. *Endocrinology*, **36**, 48 (1945)
4. ANDERSON, A. B. and OASTLER, E. G. *J. Physiol.*, **92**, 124 (1938)
5. PINCUS, J. B., NATELSON, S. and LUGOVOY, J. K. *Proc. Soc. exp. Biol. N.Y.*, **78**, 24 (1951)
6. GROLLMAN, A. *Proc. Soc. exp. Biol. N.Y.*, **85**, 582 (1954)
7. STOERK, H. C. and CARNES, W. H. *J. Nutr.*, **29**, 43 (1945)
8. ALBRIGHT, F. and ELSWORTH, R. J. *J. clin. Invest.*, **7**, 183 (1929)
9. HARRISON, H. E. and HARRISON, H. C. *J. clin. Invest.*, **20**, 47 (1941)
0. TWEEDY, W. R., CHILCOTE, M. E. and PATRAS, M. C. *J. biol. Chem.*, **168**, 597 (1947)
1. DAVIES, B. M. A. and GORDON, A. H. *Nature, Lond.*, **171**, 1122 (1953)
2. TALMAGE, R. V., LOTZ, W. E. and COMAR, C. L. *Proc. Soc. exp. Biol. N.Y.*, **84**, 578 (1953)
3. TALMAGE, R. V. and KRAINTZ, F. W. *Proc. Soc. exp. Biol. N.Y.*, **85**, 416 (1954)
4. TALMAGE, R. V., KRAINTZ, F. and BUCHANAN, G. D. *Proc. Soc. exp. Biol. N.Y.*, **88**, 600 (1955)
5. INGALLS, T. H., DONALDSON, G. A. and ALBRIGHT, E. *J. clin. Invest.*, **22**, 603 (1943)
6. STOERK, H. C. *Proc. Soc. exp. Biol. N.Y.*, **54**, 50 (1943)
7. MONAHAN, E. P. and FREEMAN, S. *Fed. Proc.*, **3**, 33 (1944)
8. GROLLMAN, A. *Endocrinology*, **55**, 166 (1954)
9. HOGBEN, C. A. M. and BOLLMAN, J. L. *Amer. J. Physiol.*, **164**, 670 (1951)
0. COXON, R. V. *J. Physiol.*, **124**, 38 P (1954)
1. MUNSON, P. L. *Ann. N.Y. Acad. Sci.*, **60**, art. 5, 776 (1955)
2. BARNICOT, N. A. *J. Anat. London*, **82**, 233 (1948)
3. CHANG, H-Y. *Anat. Rec.*, **111**, 23 (1951)
4. HANDLER, P., COHN, D. V. and DE MARIA, W. J. A. *Amer. J. Physiol.*, **165**, 434 (1951)
5. HANDLER, P. and COHN, D. V. *Amer. J. Physiol.*, **169**, 188 (1952)

56. STEWART, G. S. and BOWEN, H. F. *Endocrinology*, **80** (1952)
57. TEPPERMAN, H. M., L'HEUREUX, M. V. and WILFONG, A. E. *J. biol. Chem.*, **168**, 151 (1947)
58. KENNY, A. D., VINE, B. C. and COOKE, R. E. *Fed. Proc.*, **13**, 241 (1954)
59. HEIMER, C. B., MASLOW, H., SOBEL, A. E. and GRANT, D. M. *Proc. Soc. exp. Biol. N.Y.*, **87**, 13 (1954)
60. COHEN, J., MADDOCK, C. L. and WOLBACH, S. B. *Path.*, **59**, 723 (1955)



## CHAPTER X

# Bone Formation and the Influence of Various Factors upon this

A SECTION on histology may seem somewhat out of place in a monograph primarily biochemical. But there is a close correlation between the structure of bone, especially during development, and its chemical changes, and a brief account of the microscopic appearances must be given. Furthermore, certain parts of the bone, especially the rachitic cartilage, have been used in enzyme studies. The reader who wants a more complete description is referred to such works as that of Weinmann and Sicher [1].

There are two types of osteogenesis—endochondral and intramembranous. In both, specialized cells are concerned with bone formation and removal. The former cells, osteoblasts, are differentiated mesenchyme cells; they can be seen to take on this function especially in the early stages of intramembranous bone formation. They are usually cubical or flattened cells, with cytoplasm staining strongly with basic dyes, and lie in a row along the face of the bone they are forming. The 'bone removing' cells, or osteoclasts, are still one of the mysteries of osteogenesis. They are large cells with many nuclei, usually about six, and the cytoplasm takes up acidic stains in a characteristic way. They also lie along the bone face and often in little niches called Howship's lacunae. In spite of dogmatic statements by various authors, the present writer does not believe that the functions of these cells is yet settled. They are always present where bone resorption is occurring and they can appear and disappear with considerable

speed. No mitoses are seen in their development, and it has been suggested that they may form by fusion of osteoblasts. Compared with osteoblasts which line the bone face in a continuous row, osteoclasts are seen less frequently but have been shown to be motile [Hancox, 2]. Whether they actively resorb bone is not proved, but it is possible that they ingest the products of bone resorption since Bailie and Irving [3] found silver staining granules in them. They are macrophages in the accepted sense as they will not take acid dyes like trypan blue. They appear to have a fibrillary matrix next to the bone that they touch which some workers have associated with bone resorption, but Ham [4] has suggested that this is due to the fibrils of bone becoming visible during resorption. An excellent review of all the possibilities has been written by Hancox [5]. Heller *et al.* [6] studied the bone cellular changes in hyperparathyroidism, from which it appears that a kind of dynamic equilibrium exists between the processes associated with bone formation and destruction, so that changes from one to another can take place with ease. While this concept is a great advance, it still does not explain the detailed influences causing changes in this equilibrium nor how particular parts of a given bone are handled in a particular way.

In resorption of bone, removal of matrix and the calcified elements proceed at the same time. Removal of lime salts leaving behind the uncalcified matrix, a process called histolysis, is not considered to occur [McLean and Bloom, 7].

In intramembranous bone formation, certain of the mesenchyme cells take on the functions of osteoblasts. In the growing animal, one finds the appositional side of the bone lined with osteoblasts, and the resorptive side with osteoclasts. As the osteoblasts lay down bone, they become enclosed in lacunae as osteocytes or bone cells, with long processes extending through the matrix. Whether the osteoblasts merely calcify preformed matrix or actually make it as well is not known, but thick collagenous fibres can usually be seen extending from the tissue round the bone, between the osteoblasts and into the bone substance [Irving, 8], rather like coarse Korff fibres associ-

with dentin formation. Uncalcified bone matrix is called osteoid and normally this is rarely seen since matrix formation and calcification proceed at equal speeds. Only when bone formation is very rapid does one see uncalcified margins of osteoid. Calcified matrix differs from osteoid in its histological staining, being strongly basiphilic while osteoid is eosinophilic. Osteoid margins are seen typically in rickets when calcification lags behind matrix formation, and even after histological decalcification the difference in staining is still present. Weinmann and Sicher [1] suggest that some chemical change occurs in the organic cementing substance of osteoid before it can be calcified, and that this causes the difference in staining reaction.

Endochondral ossification is seen usually at the ends of long bones. Here a cartilage matrix is first used as a scaffold for the formation of the primary spongiosa, which is then remodelled in accordance with the stresses and strains imposed on the bone. Concurrently the shaft increases in width and thickness by processes of intramembranous ossification.

Dodds and Cameron [9] have given a very complete summary of the normal process and that occurring during rickets. They divide the epiphyseal cartilage into five zones: 1, reserve cells, a narrow layer; cells divide to produce daughter cells. 2, cell multiplication, a wide zone of up to 12 cells in thickness; cells divide to produce groups of 4 cells. 3, cell growth, the cells becoming square in profile; not a wide zone. 4, fully grown or hypertrophic cells; these cells look almost like plant cells, being large and translucent in appearance; they contain much glycogen; calcification starts in the longitudinal walls between the cell columns. 5, cartilage removal; by this time the cartilage cells are considered by many to be dead; narrow tongues of marrow, rich in capillaries, advance and remove the transverse cell walls and also some of the longitudinal walls between the cell columns. The remaining longitudinal walls are used as a foundation for bone deposition (primary calcification). Calcification is usually 3-4 cartilage cells ahead of cartilage removal.

A striking aspect of the whole picture is the regularity of the direction of the cartilage cells, which lie in columns of several lines from where they are first formed, through the various phases of their development, until they are finally eroded and removed.

At the site of primary calcification a honeycomb appearance is seen of calcification on the remaining cartilage (proliferative zone). As one progresses down the diaphysis, considerable remodelling occurs, stouter trabeculae of pure bone replace the more slender ones, and these new trabeculae, being laid sideways, make a firm connection with the shaft. Fragments of cartilage may still be seen in quite mature trabeculae and may persist for a long time. Below this secondary spongiosa most long bones form a tube which is filled with marrow.

The spongiosa forms a readily available store of Ca which can be easily withdrawn. This was shown by Bauer *et al.* using alizarin red which stains the most recently formed bone. Warnock and Duckworth [11] have also found the same.

With the cessation of growth, the epiphysis closes and the cartilage disappears. The formation of bone and exchange of bone salts do not stop however with the cessation of growth. Bone is a most labile tissue, and in spite of its apparent stability of structure, remodelling and replacement go on continuously. The osteoporosis of old age is due to breakdown being greater than new formation.

## EFFECT OF VARIOUS FACTORS UPON OSSIFICATION

### Dietary

*Ca, P and vitamin D.* Deficiency in any or all of these causes histologically very similar appearances, those of rickets. Experimentally it is possible to produce rickets in growing rats and rodents in general using diets with a Ca : P ratio of 4 or more (e.g. the Steenbock-Black diet), and a similar but not so marked condition is seen if the dietary Ca : P ratio is low, around 0.25. The requirement of vitamin D in rats is low [Irving, 12] and vitamin D free diets cause only



interference with calcification. Rickets is easily produced in puppies with diets low in vitamin D [Mellanby, 13] and in children vitamin D deficiency is the usual cause, though cases of rickets reported in South Africa, where considerable solar radiation should ensure adequate vitamin D, may be due to Ca deficiency.

The prime change in endochondral ossification in rickets is an inability to calcify and erode the hypertrophic cartilage cells. There is some disagreement over which of these processes stops first. Dodds and Cameron [9] consider failure of calcification to be the prime fault and Park also holds this view [14]. Shohl and Wolbach [15] consider that the cartilage cells do not degenerate and thus capillary invasion, which they regard as comparable to granulation tissue formation following tissue defect, does not occur. In the early stages calcification and cartilage removal are at the same level instead of being 3-4 cells apart. Later, calcification of the hypertrophic cells ceases completely and with it cartilage removal. The hypertrophic cartilage cell layer becomes greatly thickened and forms a new part of the bone structure called the metaphysis, which is visible radiologically as a widened translucent zone. Osteoid is often deposited on this cartilage. Later in the condition a lawless invasion and removal of cartilage in all directions, called 'resumed cartilage removal' by Dodds and Cameron, occurs, the eroded spaces being filled with twisted masses of osteoid. The rachitic metaphysis has been used by those studying the enzymatic changes in calcification, as it will recalcify *in vitro* if placed in appropriate media.

The changes in intramembranous bone during the onset of rickets have been recently described by Bailie and Irving [3]. The first change is a slowing of resorption, the bone gradually becoming wider. Ten days after being placed on the rachitogenic diet, an osteoid margin appears on the appositional side of the bone. This margin gradually becomes wider until by twenty-eight days the bone is entirely osteoid. Between the 10th and 28th day the osteoblast, while retaining the ability to make matrix, gradually loses the power to calcify it.



Resorption of the uncalcified matrix does not occur and the osteoclasts gradually disappear.

As a result of the large amount of osteoid which is formed, the proportion of calcified mass to total mass of bone is changed. This is usually estimated as the bone ash which may fall from the normal value of about 65 per cent of dry fat-free bone to less than half this figure. In the writer's opinion the bone ash figure alone is misleading in the diagnosis of rickets, since he regards rickets by definition as a histological change caused by biochemical factors.

Rickets can be caused by other factors precipitating phosphates in the gut, e.g. iron [16], strontium [17] and beryllium [18]. Be and Sr have in addition a direct action on calcification processes in bone.

Osteomalacia is a form of adult rickets. Since endochondral ossification has ceased, no epiphyseal changes are seen and the process of calcification of replaced bone is interfered with and osteoid is laid down instead. This condition is rare in Europe, but cases have been described in the Orient, especially in women after multiple pregnancies and lactation when the skeleton has collapsed and crippled the patient. Osteoporosis is a different condition and has a definite inheritance [Albright, 19], implying a deficient formation of bone which is normally calcified.

Rickets can be healed by giving vitamin D, starvation or a high Ca : P ratio diet is used (owing to mobilization of Ca from the soft tissues), or by adjusting the Ca : P ratio of the diet. With vitamin D the classical 'line' response is seen in the early stages, due to the deposition of lime salts in a line at the metaphysis, where they can be visualized with silver nitrate. Six stages of healing have been described by Dyer [20] and Coward [21] and they form the basis of the biological action of the vitamin. If the animal is given extra Ca in the diet, healing is usually so precipitous that no line is seen in the whole metaphysis filling with calcification. The site of the line and, in fact, the development of it depends, in the writer's experience, on the development of a high degree of

ickets and a great deal of 'resumed cartilage removal', since calcification occurs at the edge of the epiphyseal cartilage, however wide, and if the metaphysis is all cartilage the new calcification occurs at the junction with the diaphysis and no line is seen. It is sometimes stated that the new calcification occurs where it would have done normally. This is true in the sense that the edge of the epiphyseal cartilage is always the site of calcification, but geographically this can be anywhere in the metaphysis depending on the degree of 'resumed cartilage removal'. Exactly the same kind of calcification is seen *in vitro* if the end of the rachitic bone is incubated in a calcifying medium.

The action of vitamin D upon intestinal absorption has been described in an earlier chapter. There is a good deal of evidence that the vitamin has other actions. McGowan [22] some years ago suggested that it mobilized P inside the body, possibly from phospholipins. Irving [12] found in a study of the mental reactions to the vitamin that with small doses as much as eight days might elapse before new calcification began and considered this must be due to the time taken to mobilize from inside the body the elements needed for calcification, and more recently Underwood *et al.* [23] have reported that vitamin D mobilized P from organic sources.

Work with radio-Ca shows that vitamin D acts directly on, at any rate, rachitic bone. Greenberg [24] gave rachitic rats  $\text{Ca}^{45}$  by injection combined with vitamin D by mouth, the controls getting no vitamin. The percentage of the dose entering the skeleton in the controls was 28 per cent., and in those getting vitamin D, 45 per cent. Migicovsky and Emslie [25] could not confirm this in chicks and considered there was no direct action of vitamin D on bone; vitamin D had the same action in rats and chicks, but there was a difference in Ca metabolism and especially in the effect of vitamin D on the renal excretion of Ca and phosphate, the take-up of Ca by the rat bones being secondary to this [26]. However, Underwood *et al.* [23], in their turn, did not agree with Migicovsky and his colleagues and produced strong evidence for their

objection. When  $\text{Ca}^{45}$  but no vitamin D was given to or rachitic animals, there was a rise in blood Ca and bone  $\text{Ca}^{45}$  due to exchange. When vitamin D was given was a later second rise in bone  $\text{Ca}^{45}$  (especially in the rachitic animals) but not in the untreated animals, at a time when blood Ca was not rising, or was even falling. Work with radio-P has shown exactly the same [Morgareidge and Parfitt, 27], the bone  $\text{P}^{32}$  rising only in rachitic animals given vitamin D, while the blood  $\text{P}^{32}$  level was the same in these and control animals. Claasen and Wöstman [28] reported similar findings. Cohn and Greenberg [29] found that inorganic P of rachitic rat bone to be raised by vitamin D and they postulated that the vitamin facilitated the conversion of organic to inorganic P. Thus the evidence is that vitamin D has a direct action on bone, as well as indirect effects upon intestinal absorption.

An interesting action of vitamin D upon the internal metabolism of bone has been recently described by Tietzel and Patwardhan [30]. These writers found that cartilage from untreated rachitic rats could not oxidize pyruvic acid, but the cartilage from rachitic rats given vitamin D could. The oxidation of pyruvic acid by liver and kidney homogenates of rachitic rats was unimpaired.

The effects of excessive amounts of vitamin D on bone metabolism have been studied by Ham and Lewis [31]. The chief changes were overgrowth of matrix and the development of osteoid. Cortical bone is absorbed to a great extent and covered by large amounts of osteoid, the process being unaccompanied by an osteoclast response. Barnicot [32], using his implantation technique, also found that pieces of bone attached to small pellets of calciferol and grafted intracerebrally into immature mice, underwent diffuse resorption.

*Vitamin A.* In deficiency of this vitamin, a curious overgrowth of bone takes place. This was first described by Moore *et al.* [33] in calves and it was shown to be prevented by  $\beta$  carotene [Moore, 34]. Wolbach [35] considered that the normal processes of bone remodelling were accentuated, but

seems improbable since new bone development can take place on sites where normally resorption occurs. Mellanby [36] also analysed the changes and showed that the position of osteoblasts and osteoclasts could become reversed on certain bones, and this accounted for the development of new bone in abnormal situations. Irving [37] considered that the action of osteoblasts became uncontrolled in the absence of vitamin A so that they formed excessive bone of a primitive type. In areas where normally resorption occurred, but bone was laid down in avitaminosis A, the adjacent cells actively divided and became osteoblasts [Irving, 8]. In hypervitaminosis A, bone formation ceases and fractures occur [Moore and Wang, 38], possibly due to suppression of osteoblast activity, resorption continuing in a normal way [Irving, 37]. Barnicot [39] found that when bone implanted into the brain was in contact with a crystal of vitamin A, resorption occurred with many osteoclasts (this action being specific to vitamin A) and proposed that the action of the vitamin in excess was to cause an active resorption. Fell and Mellanby [40] using mouse foetal bones in tissue culture found that vitamin A in excess caused resorption of the bone but no great number of osteoclasts were present. In long bones which fracture, no particular cellular reaction is seen and it is possible that the fault lies in the organic matrix.

*Vitamins of the B group.* The changes in bone formation during deficiency of riboflavin, pantothenic acid and pyridoxine have been investigated. In riboflavin deficiency [Nelson *et al.*, 41] a gradual cessation of calcification occurs, the primary spongiosa disappearing, the epiphyseal cartilage becoming narrower and finally being sealed off with bone. Virtually the same was seen in pantothenic acid deficiency [Nelson *et al.*, 42]. Silberberg and Levy [43] reported similar findings in pyridoxine deficiency. This picture is presumably more due to interference with matrix formation than to diminution of calcification, and is not apparently specific to lack of these vitamins, as an identical picture is seen after inanition, protein deficiency or hypophysectomy [44].



*Ascorbic acid.* In scurvy a very characteristic picture emerges due to interference with the activity of the osteoblast, which modulates back to a fibroblast-like cell. A great deal of work has been done on this condition. Delf and Tozer [45] published an early account of experimental scurvy, and Wolbach and Howe [46] described the changes in great detail, as has been done by many writers since.

The chief change is an interference with ossification which leads to weakness of the bones, fractures and infraction of areas where growth is taking place. The subsequent transformation imposed on the original pathological process gives a complicated picture which is difficult to interpret. In addition, the changes vary in different bones and in different parts of the same bone, largely as a result of the rate of bone growth and the degree to which these bones are used. For these reasons the manifestations of scurvy are more severe in growing animals than in adults.

The first change is a cessation of bone apposition. The periosteum gradually thickens, the cells composing it become like fibroblasts. Resorption with accompanying osteoporosis continues unchanged. As a result, the shaft, especially the part at the metaphysis where remodelling normally takes place, becomes very thin and may disappear. Fracture occurs with considerable haemorrhage and damage to the cartilage trabeculae and cell columns, producing an area of debris, haemorrhage and fibrin deposition—the *trümmerfeld*. When this happens, the cartilage cell sequences are not so much disrupted as is capillary penetration, which is now very irregular. As the disease progresses, a mass of fibroblast-like cells develops in the metaphysis on the diaphyseal side of the *trümmerfeld*, occupying the area usually filled with the spongiosa. This zone is known as the *gerüstmark*. This mass of cells is probably made up of inactive osteoblasts which have accumulated where they would normally be calcifying the spongiosa [46]. This concept is supported by the finding of rapid new calcification in the *gerüstmark* after ascorbic acid administration [Menkin *et al.*, 47]. Stunted trabeculae



produced, but this may be due to the local trauma as the bone becomes weakened. Follis [48] found that if the limb was immobilized a cartilage lattice formed but was not calcified.

The fundamental cause of these changes is still a matter of dispute, but it would probably be generally agreed that they are due to the inability of the osteoblast to secrete a calcifiable matrix. The collagen content of bones and teeth is reduced during scurvy [Robertson, 49], but the blood Ca and P levels are unchanged [Todhunter and Brewer, 50].

*Vitamin E.* No change occurs in endochondral ossification in deficiency of this vitamin [Irving and Budtz-Olsen, 51].

### Endocrine factors

*Pituitary.* The effects of this organ have been widely studied, especially by the Becks school, using the recently available growth hormone. Hypophysectomy stops all endochondral bone sequences, the changes being like those in ageing normal animals [Becks *et al.*, 52; Walker *et al.*, 53]. On giving the growth hormone to normal animals, the growth sequences persist and the bones are larger and longer [Evans *et al.*, 54]. When the growth hormone is given to hypophysectomized animals, endochondral ossification is at once stimulated [Kibrick *et al.*, 55] and an almost normal histological picture is produced which is however continued for the whole experimental period [Becks *et al.*, 56] so that skeletal maturity, in the sense of closure of epiphyses, is not attained [53]. It may be that the growth hormone acts directly on the bone, since Blumenthal *et al.* [57] found, after administering it chorionallantoically to chicks during the last half of embryogenesis, that there was a great increase in the weight of bones and in mitotic activity. As is well known, overactivity of the gland produces clinically excessive bone growth in the conditions acromegaly and gigantism.

The interplay of hormones has also been studied. Thus testosterone mitigates to some extent the effects of hypophysectomy [Simpson *et al.*, 58]. When thyroxine is given to the hypophysectomized animal, the cartilage plate, sealed off by

bone as a result of lack of growth hormone, is now completely removed and the epiphysis closed [Asling *et al.*, 59].

*Thyroid.* Thyroidectomy causes a slowing in the rate of bone growth and differentiation. Endochondral ossification goes on, but at a very slow rate, so that the bone of a 72-day-old rat has the appearance of that of one 15–20 days old. Growth hormone stimulates growth but not differentiation of the tissues of the thyroidectomized animal, whereas if thyroxine is given, both growth and differentiation occur [Becks *et al.*, 60]. Long-continued overactivity of the gland produces osteoporosis, and the administration of the extract to normal animals causes an increased Ca excretion [Albright *et al.*, 61].

*Adrenals.* Adreno-corticotrophic hormone has been shown to retard chondrogenesis and osteogenesis in the normal rat [Becks *et al.*, 62] and also in the hypophysectomized rat given growth hormone [Becks *et al.*, 63]. It has hardly any further effect upon the bone changes in hypophysectomized animals. In Cushing's syndrome osteoporosis is common, and it is of interest that Albright [64] in his Harvey lecture predicted the existence of an anti-anabolic factor in the adrenal cortex some years before cortisone was isolated. When rats were both adrenalectomized and hypophysectomized, the bone picture was similar to that after hypophysectomy and the response to growth hormone was the same as if adrenalectomy had not been done—no increase in sensitivity was found [Simpson *et al.*, 65]. Follis [66] reported that cortisone, when given to growing rats, caused a dense zone of spicules of calcified cartilage matrix at the growing end of the bone which he thought was caused by a disturbance in the normal osteolytic activity, but in mice, rabbits and guinea-pigs no such action occurred. The reaction of the rat is thus in some respects anomalous.

*Oestrogens.* The action of oestrogens on bone is of great interest. It was first noted by Kyes and Potter in 1934 [67] that the female pigeon during the egg-laying period developed an excess of spongy bone in the marrow cavity of the long bones (medullary bone). When the Ca was needed for egg-

all formation the excess bone was rapidly removed. During this time, from pre-ovulation till the eggs were laid, the blood Ca rose to high figures. This effect could be reproduced in the pigeons and also in mice (female and male) by the administration of oestrogen [Gardner and Pfeiffer, 68]. Male hormones had no such effect and the removal of the pituitary parathyroids did not prevent the hypercalcaemia [Riddle and McDonald, 69]. The local application of oestradiol to chips of bone implanted in the brain of litter mate mice was not followed by any bone change [Barnicot, 32].

The blood Ca, P and phosphatase of mice do not change in level when oestrogens are given. Urist *et al.* [70] investigated the action of oestrogens in other animals, and of those they tested found young rats the only ones to be affected, but in a rather different way. Here the resorption of cartilage and new bone was prevented so that a very large spongiosa developed. In a later paper [Budy *et al.*, 71] it was found that the effect could be obtained in both males and females, that no effect was obtained in rachitic animals and that desoxycorticosterone and probably testosterone did not produce any bony changes. In humans it is stated that the long-continued administration of oestrogen is followed by hyperossification [Cooke, 72], and Albright *et al.* [73] have reported that defective bone growth and osteoporosis could be found in both sexes in primary gonadal insufficiency. It would appear, however, that the principal effect of oestrogens in bone formation is concerned with egg-laying in birds, and the reason for a somewhat similar action in some mammals is not very evident, as it appears to fulfil no physiological need. In pregnant mice neither the foetal nor maternal skeletons show any medullary bone formation.

*Parathyroid glands.* These organs have the most profound controlling influence upon Ca and P metabolism. The general effects are discussed in the section on the blood Ca, and here only the histological changes in bone will be considered. As far as the present writer knows, no description has been given of the effects of ablation of the glands on bone,

presumably because the experimental animals live so short a time after the operation.

In hyperparathyroidism changes are seen in bone which are in essence a destruction, an osteoclastic reaction predominating. Osteocytes and osteoblasts are destroyed in some animals. Snapper [74] summarized the bone changes in clinical cases as: proliferation of osteoclasts which erode bone with resulting decalcification, and proliferation of fibrous tissue in the Haversian canals; cyst formation, and formation of giant-cell tumours—osteoclastomas. Recently Heller *et al.* [6] have analysed the cell changes in rats after single toxic doses of parathyroid extract, and showed how, ranging round a central spindle cell, modulations (with little mitotic activity) could take place between all the cells concerned with bone formation and destruction. It is still undecided if parathormone acts directly on bone or if this dissolution is the result of changes elsewhere. Barnicot [75], using his implantation technique with parathyroid gland and bone, found that resorption and an osteoclastic response occurred in bone in contact with the gland; when transplants of pituitary, thyroid or adrenal gland were used instead, no change occurred.

## REFERENCES

1. WEINMANN, J. P. and SICHER, H. *Bone and bones*. St. Louis: The C. V. Mosby Company (1955)
2. HANCOX, N. M. *J. Physiol.* **110**, 205 (1949)
3. BAILIE, J. M. and IRVING, J. T. *Acta med. Scand.*, **152**, Supp. 306, 1 (1955)
4. HAM, A. W. *J. Bone Joint Surg.*, **34-A**, 701 (1952)
5. HANCOX, N. M. *Biol. Rev.*, **24**, 448 (1949)
6. HELLER, M., MCLEAN, F. C. and BLOOM, W. *Amer. J. Anat.*, **87**, 315 (1950)
7. MCLEAN, F. C. and BLOOM, W. *Anat. Rec.*, **78**, 333 (1940)
8. IRVING, J. T. *Med. Klin.*, **51**, 690 (1956)
9. DODDS, G. S. and CAMERON, H. C. *Amer. J. Anat.*, **55**, 135 (1934)



9. BAUER, W., AUB, J. C. and ALBRIGHT, F. *J. exp. Med.*, **49**, 145 (1929)
10. WARNOCK, G. M. and DUCKWORTH, J. *Biochem. J.*, **38**, 220 (1944)
11. IRVING, J. T. *J. Physiol.*, **103**, 9 (1944)
12. MELLANBY, E. Spec. Rep. Ser. med. Res. Coun. Lond., **61** (1921)
13. PARK, E. A. *Bull. N.Y. Acad. Med.*, **15**, 495 (1939)
14. SHOHL, A. T. and WOLBACH, S. B. *J. Nutr.*, **11**, 275 (1936)
15. BROCK, J. F. and DIAMOND, L. K. *J. Pediat.*, **4**, 442 (1934)
16. SOBEL, A. E., COHEN, J. and KRAMER, B. *Biochem. J.*, **29**, 2640 (1935)
17. GUYATT, B. L., KAY, H. D. and BRANION, H. D. *J. Nutr.*, **6**, 313 (1933)
18. ALBRIGHT, F. *Ann. intern. Med.*, **27**, 861 (1947)
19. DYER, F. J. *Quart. J. Pharm.*, **4**, 503 (1931)
20. COWARD, K. H. *The biological standardisation of the vitamins*. London: Baillière, Tindall and Cox (1938)
21. MCGOWAN, J. P. *Biochem. J.*, **27**, 943 (1933)
22. UNDERWOOD, E., FISCH, S. and HODGE, H. C. *Amer. J. Physiol.*, **166**, 387 (1951)
23. GREENBERG, D. M. *J. biol. Chem.*, **157**, 99 (1945)
24. MIGICOVSKY, B. B. and EMSLIE, A. R. G. *Arch. Biochem.*, **28**, 324 (1950)
25. MIGICOVSKY, B. B. and NIELSON, A. M. *Arch. Biochem. Biophys.*, **34**, 105 (1951)
26. MORGAREIDGE, K. and MANLEY, M. L. *J. Nutr.*, **18**, 411 (1939)
27. CLAASEN, V. and WÖSTMAN, B. S. J. *Biochim. biophys. Acta*, **12**, 557 (1953)
28. COLIN, W. E. and GREENBERG, D. M. *J. biol. Chem.*, **130**, 625 (1939)
29. TULPOLE, P. G. and PATWARDHAN, V. N. *Biochem. J.*, **58**, 61 (1954)
30. HAM, A. W. and LEWIS, M. D. *Brit. J. exp. Path.*, **15**, 228 (1934)
31. BARNICOT, N. A. *J. Anat. Lond.*, **85**, 120 (1951)



33. MOORE, L. A., HUFFMAN, C. F. and DUNCAN, C. W. *J. Nutr.*, **9**, 533 (1935)
34. MOORE, L. A. *J. Nutr.*, **17**, 443 (1939)
35. WOLBACH, S. B. *Proc. Inst. Med. Chicago*, **16**, 118 (1946)
36. MELLANBY, E. *J. Physiol.*, **105**, 352 (1947)
37. IRVING, J. T. *J. Physiol.*, **108**, 92 (1949)
38. MOORE, T. and WANG, Y. L. *Biochem. J.*, **39**, 222 (1945)
39. BARNICOT, N. A. *J. Anat. Lond.*, **84**, 374 (1950)
40. FELL, H. B. and MELLANBY, E. *J. Physiol.*, **116**, 320 (1952)
41. NELSON, M. M., SULON, E., BECKS, H. and EVANS, H. M. *Proc. Soc. exp. Biol.*, N.Y. **66**, 631 (1947)
42. NELSON, M. M., SULON, E., BECKS, H., WAINWRIGHT, W. W. and EVANS, H. M. *Proc. Soc. exp. Biol.* N.Y., **73**, 31 (1950)
43. SILBERBERG, R. and LEVY, B. M. *Proc. Soc. exp. Biol.* N.Y., **67**, 259 (1948)
44. FOLLIS, R. H. Trans. 2nd Conf. metab. Interr. New York, Josiah Macy Jr. Foundation, p. 221 (1950)  
FRANDSEN, A. M., NELSON, M. M., SULON, E., BECKS, H. and EVANS, H. M. *Anat. Rec.*, **119**, 247 (1954)
45. DELF, E. M. and TOZER, F. M. *Biochem. J.*, **12**, 416 (1918)
46. WOLBACH, S. B. and HOWE, P. R. *Arch. Path.*, **1**, 1 (1926)
47. MENKIN, V., WOLBACH, S. B. and MENKIN, M. F. *Amer. J. Path.*, **10**, 569 (1934)
48. FOLLIS, R. H. *Arch. Path.* **35**, 579 (1943)
49. ROBERTSON, W. V. B. *J. biol. Chem.*, **187**, 673 (1950)
50. TODHUNTER, E. N. and BREWER, W. *Amer. J. Physiol.*, **130**, 310 (1940)
51. IRVING, J. T. and BUDTZ-OLSEN, O. E. *Brit. J. Nutr.*, **9**, 301 (1955)
52. BECKS, H., SIMPSON, M. E. and EVANS, H. M. *Anat. Rec.*, **92**, 121 (1945)
53. WALKER, D. G., ASLING, C. W., SIMPSON, M. E., LI, C. H. and EVANS, H. M. *Anat. Rec.*, **114**, 19 (1952)
54. EVANS, H. M., BECKS, H., ASLING, C. W., SIMPSON, M. E. and LI, C. H. *Growth*, **12**, 43 (1948)

5. KIBRICK, E. A., BECKS, H., MARX, W. and EVANS, H. M. *Growth*, **5**, 437 (1941)
6. BECKS, H., ASLING, C. W., SIMPSON, M. E., LI, C. H. and EVANS, H. M. *Growth*, **13**, 175 (1949)
7. BLUMENTHAL, H. T., HSIEH, K-M and WANG, T-W. *Amer. J. Path.*, **30**, 771 (1954)
8. SIMPSON, M. E., MARX, W., BECKS, H. and EVANS, H. M. *Endocrinology*, **35**, 309 (1944)
9. ASLING, C. W., BECKS, H., SIMPSON, M. E. and EVANS, H. M. *Anat. Rec.*, **104**, 255 (1949)
10. BECKS, H., SCOW, R. O., SIMPSON, M. E., ASLING, C. W., LI, C. H. and EVANS, H. M. *Anat. Rec.*, **107**, 299 (1950)
11. ALBRIGHT, F., BAUER, W. and AUB, J. C. *J. clin. Invest.*, **10**, 187 (1931)
12. BECKS, H., SIMPSON, M. E., LI, C. H. and EVANS, H. M. *Endocrinology*, **34**, 305 (1944)
13. BECKS, H., SIMPSON, M. E., MARX, W., LI, C. H. and EVANS, H. M. *Endocrinology*, **34**, 311 (1944)
14. ALBRIGHT, F. *Harvey Lect.*, **38**, 123 (1942-3)
15. SIMPSON, M. E., MARX, W., BECKS, H. and EVANS, H. M. *Endocrinology*, **35**, 234 (1944)
16. FOLLIS, R. H. *Proc. Soc. exp. Biol. N.Y.*, **76**, 722 (1951); **78**, 723 (1951)
17. KYES, P. and POTTER, T. S. *Anat. Rec.*, **60**, 377 (1934)
18. GARDNER, W. U. and PFEIFFER, C. A. *Anat. Rec.*, **73**, Supp. 21 (1939)
19. RIDDLE, O. and MCDONALD, M. R. *Endocrinology*, **36**, 48 (1945)
20. URIST, M. R., BUDY, A. M. and MCLEAN, F. C. *Proc. Soc. exp. Biol. N.Y.*, **68**, 324 (1948)
21. BUDY, A. M., URIST, M. R. and MCLEAN, F. C. *Amer. J. Path.*, **28**, 1143 (1952)
22. COOKE, A. M. *Lancet*, **1**, 877, 929 (1955)
23. ALBRIGHT, F., BLOOMBERG, E. and SMITH, P. H. *Trans. Ass. Amer. Phycns.*, **55**, 298 (1940)
24. SNAPPER, I. *Acta med. Scand.*, **103**, 321 (1940)
25. BARNICOT, N. A. *J. Anat. Lond.*, **82**, 233 (1948)

## CHAPTER XI

# Bone – Chemistry and Physics

No attempt is made in this chapter to deal exhaustively with the chemistry and physics of bone. This would involve a great deal more space than available and has in any case already been done by a number of writers [Neuman and Neuman, 1; McLean and Urist, 2; Dallemagne, 3; Carlström, 4]. Only salient points which seem pertinent to the physiology of Ca metabolism will be discussed and the writer has in general followed these authors in his presentation.

Bone consists of an organic matrix, collagen, which is fibrillar in nature, and an amorphous ground substance which contains a mucopolysaccharide, probably chondroitin sulphate; in this organic framework are branching bone cells or osteocytes. About one-third of the bone mass is composed of minute crystals embedded in the organic matrix; these crystals contain chiefly Ca, phosphate and carbonate and confer on bone its hardness.

The organic matrix of bone is similar in general to that of other connective tissue and it may be asked why calcification is restricted to bone. This restriction is at times overruled, but under normal conditions the presence of bone-forming cells or osteoblasts is peculiar to bone only. In addition, the 'local factor' is postulated as conferring calcifiability on bone tissues.

The crystals of bone are microscopic in size and can barely be seen with the electron microscope. Authors agree that they are 25–30 Å thick, approximately 400 Å long and nearly as wide [Robinson and Watson, 5]. They are found in close association with the collagen, lying on the characteristic doublet banding of the fibres with the long crystal axis

parallel to the longitudinal direction of the fibre. The surface area of the crystals is enormous, about  $100 \text{ m.}^2$  per g., which would mean a total surface area of over 100 acres for the bone crystals of a 70-kg. man. This gives strong support to the conclusion that the available bone and plasma are in dynamic equilibrium.

The composition of these crystals has proved very hard to determine, although X-ray analysis undoubtedly shows that the basic lattice structure of the bone salt is an apatite, but this is not to be regarded as a compound but more as an arrangement in space of the ions found in the mineral. The crystal of the bone salt is roughly hexagonal, with the  $c$  axis of the unit cells oriented in its long dimension. This unit cell is a conceptual configuration and has no independent existence. It is the simplest arrangement of the atoms, treated as points, that embraces all the different ions found in the crystal, in the same ratios (with the smallest common multiple) and in the same spatial relationships in which they are present in the crystal' [McLean and Urist]. When these units are outlined by imaginary lines and extended through the crystal structure, they give the three-dimensional crystal lattice. The unit in a cross-section of the crystal is a two-dimensional parallelogram with four equal sides of  $9.4 \text{ \AA}$  in length, the  $a$  axis of the cell. In three dimensions, the unit cell is a six-sided right prism and the other dimension or  $c$  axis measures  $6.9 \text{ \AA}$ . The  $c$  axis is oriented in the long dimension of the bone salt crystal.

The crystallites of bone salt are roughly hexagonal in shape with columns of Ca and oxygen arranged at the intersections of the hexagons and of  $\text{OH}^-$  ions running in columns down the middle of the hexagon, the hexagons also being packed with phosphate. 47.3 per cent. of the unit cells are on the surface of the crystal (of dimensions as stated above), and about 50 per cent. of the units have two faces exposed.

The nature of the bone salt has been much investigated and is still a matter of dispute. One great trouble is that the particles being so small, adsorption can easily occur and this



alters their chemical composition. Furthermore, freeing the bone of its organic components changes the nature of the solid phase. Even water will do this. A favourite method is to use KOH in boiling anhydrous glycerol, but here the water in bone will give an alkaline solution which will change tricalcium phosphate hydrate (which Dallemagne and his colleagues think is the solid phase of bone) into hydroxyapatite. Calcination, also often used, causes the crystals to grow and may induce reactions between apatite and ions outside the lattice. Thus it will be seen that the problem is no easy one.

Armstrong [6] has given a good summary of the possible concepts. These are that bone salt is: 1, carbonate hydroxyapatite, 2, tri-calcium phosphate hydrate, or 3, hydroxyapatite.

1. Carlström has put forward evidence that the concept of carbonate apatite with carbonate present in the crystal lattice is not correct and that the carbonate is present as a surface component or a separate phase, the former more probable. Neuman and Neuman have summarized the literature which is against this hypothesis. They prefer to talk of carbon dioxide and consider that this is located at the crystal surface and not present as a separate phase.

2. The tricalcium phosphate hydrate hypothesis is largely that of Dallemagne and his school and they have brought forward a great deal of evidence in favour of it [7]. It implies that the carbonate is present as a separate phase as  $\text{CaCO}_3$ , which is still unproved. The chief reason why they favour tricalcium phosphate hydrate is that hydroxyapatite has a molar Ca:P ratio of 1.66, whereas that of bone is usually lower, about 1.5, but varies with the dietary intake of the minerals. The molar Ca:P ratio of tertiary phosphate is 1.50. However, it seems doubtful if the methods used by Dallemagne are applicable with the accuracy he assumed. Thus all poorly crystalline apatite-like precipitates, irrespective of their composition, give indistinguishable X-ray diffraction patterns, and refractive indices of precipitates of crystallites of colloidal dimensions are of no value. Neuman has proposed that hydrogen ions exchange for Ca ions in the surface layer of the

apatite unit cell and that this and also  $\text{Na}^+$  exchange for  $\text{Ca}^{++}$  account for changes in the Ca : P ratio.

3. It is generally agreed that the basic lattice structure of bone is an apatite and hydroxyapatite is taken as the prototype. It is quite clear that bone salt is not simply this, but much work has now been done showing that surface phenomena, due to the very large surface presented, can explain many of the figures which might be considered discrepant.

The bone crystal can be pictured as consisting of 'a surface hydration shell containing non-specific boundary anions in rapid equilibrium with the surrounding medium; an inner, crystal surface containing more or less specific cations and anions also in equilibrium with the solution (or the hydration water); and interior ions with a slow but measurable equilibration with the outer layers (recrystallization)' [Neuman and Neuman.] Carlström pictures it as 'built up from a core of hydroxyapatite covered by a shell of different ions and water'. An ion can be in one of four positions in relation to hydroxyapatite. It may be: 1. in an interior cell of the crystal lattice and thus in a relatively stable position, 2. in a surface position of the crystal lattice, 3. adsorbed on the surface of the crystal, 4. in solution in the hydration shell.

Surface adsorption can account for a good deal of variation in the Ca : P ratio of the mineral, but there are limits beyond which it is not possible to go. Thus a Ca : P ratio of 1.33 is not possible as the crystals are not small enough to adsorb so much P, one ion for every four in the crystal, and likewise high Ca : P ratios cannot occur.

Surface exchange accounts for a great deal of the rapid uptake of ions. Ionic exchange is now used by many chemists using synthetic resins, clays, &c. In the case of bone, two types of exchange occur, isoionic and heteroionic. Isoionic exchange has been demonstrated with radio-isotopes *in vitro* using dentin, enamel and bone, especially by Hodge and his school. Crystal size, temperature, pH and many other factors determine the final distribution of the isotope.

In heteroionic exchange an ion on the crystal lattice is

displaced by a different ion from the solution. Thus uranyl, strontium, sodium and hydronium ions will displace Ca and carbonate ions will displace surface phosphate. This displacement is to some extent specific.  $\text{Na}^+$  and  $\text{H}_3\text{O}^+$  whose ionic radii are close to that of  $\text{Ca}^{++}$  can displace it mole for mole, but  $\text{K}^+$  cannot as its ionic radius is greater.  $\text{Sr}^{++}$  has an ionic radius almost the same as  $\text{Ca}^{++}$  and exchanges with it on an equivalent basis. A mole of uranyl ion displaces two of  $\text{Ca}^{++}$ . The exchange of  $\text{F}^-$  is unusual as it appears to be irreversible and more is taken up than can be explained by surface considerations. The citrate ion is peculiar in that 70 per cent of the citric acid in the body is found in bone, and it may account for as much as 1 per cent. of the fresh weight of bone. The size of the molecule excludes it from the apatite lattice. Exactly where the citrate is located is not very clear, but a considerable amount must be held on the surface of the apatite crystal. However, only portions of citrate are readily dissolved from powdered bone so that some must be held on entrapped surfaces.

The hydration layer is a true water of hydration and may be of considerable magnitude, as much as 80 per cent. in the case of synthetic hydroxyapatite. In fresh untreated bone the hydration is not so high, suggesting that some of the compact bone may be isolated from the circulating fluids.

Recrystallization of apatite crystals and of bone crystals occurs in an aqueous medium. Labelled Ca and P slowly equilibrate with the bulk of the mineral phase, far more slowly than in the process of exchange, and there is no release of these isotopes in short-term exposures to non-labelled buffer. Newly deposited bone salt recrystallizes much faster than old bone. As will be seen, both this process and ionic exchange play important roles in bone physiology *in vivo*.

### Calcification

Attempts to explain this by solubility product relationships have not been successful. On ordinary physico-chemical considerations, for precipitation to occur, some solubility product

not be exceeded. This was the basis of the original phosphate theory of Robison. However, as Hodge has pointed out, no solubility product has ever been demonstrated. Knowing what we do now of the complexity of the composition of the crystals, this is not surprising. Levinskas [9] working with synthetic hydroxyapatite found the determination of a solubility product impossible.

Relationships do, however, seem to exist between calcification and the solubility product of  $\text{CaHPO}_4$ . Howland and Mermer [10] many years ago found that the presence or absence of rickets in children had a relationship to the  $\text{Ca} \times \text{P}$  product of the plasma, the figures being for total Ca and P, because at that time no reliable method for determining Ca and P ions existed. More recently the findings have been more related to the ion product  $[\text{Ca}^{++}] \times [\text{HPO}_4^{--}]$  than to any other combination. McLean and Urist [2] have drawn a graph of the solubility product constants of  $\text{CaHPO}_4$  and  $\text{Ca}_3(\text{PO}_4)_2$  as related to pH and the P content of the solution, and have superimposed the minimum ion concentrations at which calcification of hypertrophic rachitic cartilage occurs *in vitro*. At pH's above 7.3, calcification is correlated with the solubility product of  $\text{CaHPO}_4$  and not with that of  $\text{Ca}_3(\text{PO}_4)_2$  and thus the ion product  $[\text{Ca}^{++}] \times [\text{HPO}_4^{--}]$  appears to be the determining factor in the process. This product defines the upper solubility limit of Ca and phosphate ions in solution, but calcification can still occur (as in adults) at lower ion concentrations. The total ion concentration determines the speed of calcification but not its occurrence, the speed being determined by the collision frequency. Thus it can be seen how Howland and Mermer's empirical formula,  $\text{Ca} \times \text{P}$ , is as reliable as the ion product itself, since the ratios of  $\text{Ca}^{++}$  and  $\text{HPO}_4^{--}$  to the total Ca and total P are relatively constant.

Neuman and Neuman [1] stress our ignorance of the calcification process, but point out that from what little we do know, the process is not one of precipitation. The ion product of Ca and phosphate ions in the serum at levels of 10 and 10 mg. per 100 ml. respectively, is far less than the solubility



product constant for  $\text{CaHPO}_4$ , yet calcification occurs from such serum without difficulty. Apatite crystals can grow at C and phosphate levels well below the solubility product constant of  $\text{CaHPO}_4$ , and further it is hard to see how precipitation can result in the formation of a highly organized and oriented mineral pattern. A process of 'seeding' must be considered instead in which possibly the organic matrix of bone can bind Ca or phosphate ions in the proper space relationships for the apatite lattice, and on to these would be added the other ions making up the complete crystal. As we shall see there is evidence that such a process does initiate calcification.

The exchange of ions *in vitro* and *in vivo* has been investigated by a number of workers. Much of this has been done using  $\text{P}^{32}$ , but the handling of this element in bone and the conclusions drawn apply in most cases equally well to Ca. Thus Falkenheim *et al.* [11] found that one-fifth of the P of bone could be exchanged *in vitro*, using  $\text{P}^{32}$ , and considered that this must represent P on crystal surfaces. In a later paper [12] the same was reported for Ca using  $\text{Ca}^{45}$ . Dawson [13] stated that 33 per cent. of the Ca of unashed bone was exchangeable.

The exchange of Ca and P *in vivo* has also been investigated with radio isotopes. Neuman and Riley [14] using  $\text{P}^{32}$  realized that the rapid uptake *in vivo* was not due to normal growth but to 'exchange-adsorption' between the P of the plasma and that on the surface of the bone crystal, and was no index of growth changes or P deposition. However, if  $\text{P}^{32}$  was given *in vivo* and the animal kept for longer times before sacrifice, the  $\text{P}^{32}$  became less desorbable, indicating that recrystallization had occurred as well [Neuman and Mulryan, 15]. The same was found by Singer and Armstrong [16] and Tomlin *et al.* [17], using  $\text{Ca}^{45}$ . The latter workers reported that the interstitial replacement of Ca in previously deposited mineral was very slow and that there was a non-exchangeable accretion involving about 1 per cent. of the total cortical bone per week. Arnold and Jee [18] found that  $\text{Ca}^{45}$  was rapidly removed from non-exchangeable positions by crystallization and recrystallization.

tion, where it was not available for removal by ion-exchange techniques, half of the Ca being so dealt with after the hour.

Thus one has to picture the two processes of exchange and recrystallization occurring side by side to account for bone growth and maintenance. In adult animals both processes probably account for the appearance of isotopes in bone, but in young animals the formation of new crystals is quantitatively very important. The site of deposition of isotope is governed by hydration, density and age of crystals, and fluid change. The greatest concentrations of radioactivity are found in young bone and in the most recently deposited material, such as round Haversian canals [Arnold, 19]. The same is seen in sections *in vitro* and is thus largely due to the active physico-chemical processes of exchange and recrystallization. Rachitic bone, in which skeletal maturation does not occur, likewise exhibits the high reactivity seen in growing bone. In older bone, the uniform diffuse distribution seen in radio-autographs of young animals does not occur. In established bone the circulation is poor and limited sometimes to small canaliculi connecting osteocytes. In addition, the hydration of older bone is less, though it is not known through what agencies the hydration shell becomes reduced. Thus a smaller and smaller fraction of bone, as the skeleton matures, is in equilibrium with the body fluids. While nearly 100 per cent. of the skeleton of the young rat is available, this falls to 30 per cent. as the animals become adult. Kornberg [quoted by Neuman and Neuman, 11], using  $\text{Na}^{24}$ , found the available fraction of the adult human skeleton to be 30 per cent. However, it must be realized that in one small area of bone the rate of recrystallization and exchange may vary tremendously, due to the various factors mentioned above.

Changes in the composition of bone can be quite satisfactorily accounted for by the large crystal area which can carry an excess of ions. Carlström [20] considered that the bone crystallite surface could carry an excess of 1 to 3 Ca ions and 2 phosphate groups, and this could account for all Ca : P

ratios found by analysis; chemical analysis of bone was of very restricted value in the interpretation of crystal structure. It is now well known that the inorganic composition of bone can be greatly altered by changes in the diet or blood chemistry, the most effective way being to alter the animal's acid-base balance. The bone represents a great reserve of alkalinity which supplements the buffers of the blood in maintaining normal pH. In acidotic states the bone has reduced  $\text{CO}_2$  content, often the residual Ca:P ratio (after allowing for  $\text{CO}_2$  loss) is unchanged but the total Ca:P ratio may be altered.

Shear and Kramer [21] found many years ago that newly calcified bone was low in  $\text{CO}_2$ , and in such areas the Ca:P ratio was also low. In rats at the age of about twenty days the Ca:P ratio of the bone rose and at the same time the  $\text{CO}_2$  began to increase to the adult level [Neuman and Neuman]. Sobel and his colleagues in a series of important investigations have studied this matter further; their work has been recently summarized [22]. They estimated particularly the  $\text{PO}_4$ : $\text{CO}_3$  ratio in bone, and found [23] that high Ca:P ratio diets caused higher  $\text{CO}_3$ : $\text{PO}_4$  bone ratios than in animals on low Ca:P ratio diets. The  $\text{PO}_4$ : $\text{CO}_3$  ratio in bone was related to the P: $\text{CO}_2$  ratio in serum and the conclusion was drawn that the composition of the diet was reflected in that of the blood serum and so in that of the bone [24]. The mechanism of this process is probably at least in part an exchange of carbonate for surface phosphate, the process described as heterionic exchange by Neuman and Neuman [Hodge, 25]. Posner and Stephenson [26] in confirmation of the work of Sobel *et al.* consider that 6 per cent. of the phosphate ion is an inclusion in the entrapped surfaces of the hydroxyapatite crystal and that such inclusions depend on the individual concentrations of phosphate and  $\text{CO}_2$  in the serum.

Dallemagne [27] has postulated further that the very low Ca:P ratios in young bone may be due to phosphate ions attaching themselves to preosseous organic matrix.

It was mentioned above that bone has a buffer-like action in maintaining the composition of the blood. This is true on

thin certain limits, and as far as Ca is concerned acts in concert with certain biological reactions such as osteoblast and osteoclast activity and renal action. However, it has been shown that the injection or removal of large amounts of Ca from the circulation causes but little change in blood Ca concentration [28]. As pointed out by Neuman and Neuman, this is due to the constant dynamic equilibrium between the bone crystals and the blood, and to the fact that the surface mineral exceeds the mineral content of body fluid. The blood of a 70 kg. man contains 0.25 g. Ca, while the bone surface Ca, calculated as 10 per cent. of the available Ca, calculated in its turn as 20 per cent. of the available skeleton, is 42 g. Thus the physico-chemical reserve of surface Ca is nearly 200 times the amount in the circulating blood.

## REFERENCES

- NEUMAN, W. F. and NEUMAN, M. W. *Chem. Rev.*, **53**, 1 (1953)
- MCLEAN, F. C. and URIST, M. R. *Bone. An introduction to the physiology of skeletal tissue*. Chicago: Univ. of Chicago Press (1955)
- DALLEMAGNE, M. J. *J. Physiol. Path. gén.*, **43**, 425 (1951)
- CARLSTRÖM, D. *Acta radiol., Stockh., Supp.* **121** (1955)
- ROBINSON, R. A. and WATSON, M. L. *Anat. Rec.*, **114**, 383 (1952)
- ARMSTRONG, W. D. *Trans. 2nd Conf. metab. Interr.* New York, Josiah Macy Jr. Found., p. 11 (1950)
- DALLEMAGNE, M. J. *Données récentes sur la nature et le métabolisme de l'os*. Actualités biochimiques, No. 2. Paris: Masson et Cie (1945)
- HODGE, H. C. *Trans. 3rd Conf. metab. Interr.* New York, Josiah Macy Jr. Found., p. 194 (1951)
- LEVINSKAS, G. J. *Univ of Rochester Reps.* No. 273 (1953)
- HOWLAND, J. and KRAMER, B. *Mschr. Kinderheilk.*, **25**, 279 (1923)



11. FALKENHEIM, M., NEUMAN, W. F. and HODGE, H. C. *J. biol. Chem.*, **169**, 713 (1947)
12. FALKENHEIM, M., UNDERWOOD, E. E. and HODGE, H. C. *J. biol. Chem.*, **188**, 805 (1951)
13. DAWSON, K. B. *Biochem. J.*, **60**, 389 (1955)
14. NEUMAN, W. F. and RILEY, R. F. *J. biol. Chem.*, **168**, 545 (1947)
15. NEUMAN, W. F. and MULRYAN, B. J. *J. biol. Chem.*, **195**, 843 (1952)
16. SINGER, L. and ARMSTRONG, W. D. *Proc. Soc. exp. Biol. N.Y.*, **76**, 229 (1951)
17. TOMLIN, D. H., HENRY, K. M. and KON, S. K. *Brit. J. Nutr.*, **9**, 144 (1955)
18. ARNOLD, J. S. and JEE, W. S. S. *Proc. Soc. exp. Biol. N.Y.*, **85**, 658 (1954)
19. ARNOLD, J. S. *Amer. J. Physiol.*, **167**, 765 (1951)
20. CARLSTRÖM, D. *Biochim. biophys. Acta*, **17**, 603 (1955)
21. SHEAR, M. J. and KRAMER, B. J. *Biol. Chem.*, **79**, 12 (1928)
22. SOBEL, A. E. *Ann. N.Y. Acad. Sci.*, **60**, Art. 5, 713 (1955)
23. SOBEL, A. E., ROCKENMACHER, M. and KRAMER, B. *Fea Proc.*, **2**, 70 (1943)
24. SOBEL, A. E., ROCKENMACHER, M. and KRAMER, B. *J. biol. Chem.*, **159**, 159 (1945)
25. HODGE, H. C. *Ann. N.Y. Acad. Sci.*, **60**, Art. 5, 800 (1955)
26. POSNER, A. S. and STEPHENSON, S. R. *J. dent. Res.*, **31**, 371 (1952)
27. DALLEMAGNE, M. J. *Nature, Lond.*, **161**, 115 (1948)
28. HASTINGS, A. B. and HUGGINS, C. B. *Proc. Soc. exp. Biol. N.Y.*, **30**, 458 (1932-3)

## CHAPTER XII

# Enzymes and Bone Formation

In this chapter the emphasis may appear to be more on P than on Ca, but since the two are intimately related it is clear that P deposition also involves Ca. The intervention of enzymes in bone formation was first postulated by Robison in 1923 [1] when he found phosphatase in bone. There are at least two phosphatases in the body, designated from their optimum pH's, acid and alkaline phosphatase. The former is found in the prostate gland and has little to do with calcification. Alkaline phosphatase, found in bone and other tissues, acts on monoesters of phosphoric acid at an optimal pH of 9.4, liberating inorganic P. Robison considered that this enzyme acted on a suitable substrate, the solubility product of calcium phosphate was exceeded and this salt was deposited in the process of calcification.

This theory as originally stated was soon found to be unsatisfactory. As already mentioned, calcification of bone does not occur by the exceeding of a solubility product and precipitation from supersaturated solutions. Apart from that, suitable substrates for the enzyme are none too plentiful and those in blood are in the corpuscles. The optimum pH is very much on the alkaline side. Besides this, phosphatase is found in many other tissues which do not normally calcify.

It is worth while at this point to consider phosphatase in a little more detail. There seems no doubt that phosphatase is more associated with matrix formation than with calcification. Important accounts of phosphatase distribution in bone have been given by Lorch and Siffert. Lorch [2] found no phosphatase in bone matrix or small-celled cartilage, but

present in hypertrophic cartilage (in rats not kittens), in osteocytes, Haversian canals, endosteum, inner lining of periosteum, and the linings of vascular and marrow spaces. Osteoblasts contained it, but marrow cells did likewise, so that she did not think that only the osteoblasts secreted phosphatase. Extra-cellular phosphatase was always found where ground substance was being deposited, but was absent in areas of resorption. Newly deposited bone salts were never found in areas where there was not some extra-cellular phosphatase, though the converse was not necessarily true.

Siffert [3] found that phosphatase activity and free phosphate localization did not invariably coincide and that phosphatase was more intimately related to preosseous cellular metabolism and to the elaboration of a matrix that was chemically calcifiable. The elaboration of bone matrix was always associated with phosphatase activity but could and did occur in the absence of calcification as in rickets. Calcification might occur later, in the absence of the enzyme. 'Left over' phosphatase, in cartilage remnants in the metaphysis, was still active but had no physiological significance. Bevelander and Johnson [4], in a study of the developing mandible, reported that phosphatase disappeared from the bone matrix as mineralization occurred. Morse and Greep [5] found in the normal epiphyseal cartilage that the content of phosphatase increased as the cells became more mature, the hypertrophic cell layer containing most, but the layer of primary calcification virtually none. In rickets the same was found in the cartilage cells, and osteoid also contained the enzyme.

Wislocki and Sognnaes [6] investigated the developing tooth and found the enzyme in sites of proliferation such as the developing dental pulp, tissue of the dental sac, periodontal membrane and gingival tissues. They also found phosphatase in small amounts in the predentin and dentinal fibres. In studies of amelogenesis Bevelander and Johnson [7] found phosphatase to develop in the ameloblasts just before matrix formation occurred and to remain in them during matrix formation and subsequent calcification. They also found [8]

at ribonucleic acid and phosphatase coexisted in terms of time and localization in the odontoblasts and ameloblasts and suggested that ribonucleic acid might be implicated in some phase of the calcifying mechanism.

The behaviour of phosphatase in a number of conditions is instructive. Danielli *et al.* [9] studied wound healing in normal and scorbutic guinea-pigs. In normal animals there were two phases of phosphatase activity in the healing tissue, one due to leucocyte invasion, the second probably to the differentiation of collagenous fibrous tissue. But if the animal had scurvy there was no second peak and collagen formation was absent or greatly reduced. Gould and Schwachman [10] found in scorbutic guinea-pigs that the phosphatase content of serum and bone rose and fell in parallel, that of the other tissues not being significantly altered. It will be recalled that the osteoblasts become inactive during scurvy forming the *gerüstmark*. During healing from scurvy the bone and blood phosphatases rise together, and Gould and Schwachman considered the blood phosphatase to be derived from osteoblasts. Norris and John [11] found after injection of Ra that the alkaline phosphatase of both bone and serum fell, but that of soft tissues was not affected. *In vitro* no effect on phosphatase was seen but that the phosphatase-forming cells were affected by the Ra. During rickets the blood phosphatase rises considerably as was shown many years ago by Kay. In this condition there is much matrix formation but no calcification. Incidentally it is difficult to correlate these findings with the theory of Zetterström that vitamin D is a coenzyme for phosphatase, since during rickets when calcification is in abeyance due to vitamin D lack, phosphatase is most active especially in blood, while in scurvy, when there is no lack of vitamin D, phosphatase may disappear from bone completely.

It may thus be concluded that phosphatase is concerned, certainly in bone and probably elsewhere in the body, with matrix formation and its association with calcification in bone probably incidental.

Some other quite different roles have been assigned to



phosphatase. Thus Neuman *et al.* [12] found that calcification *in vitro* was inhibited by phosphoric esters if phosphatase was inactivated; they considered that phosphatase removed ester P which acted normally as an inhibitor of calcification. Di Stephano and Neuman [13] reported that adenosine triphosphate was an inhibiting agent to calcification *in vitro* since Ca formed a relatively undissociated complex with it; they attributed this inhibiting action to the fact that phosphatase acted very slowly on ATP. While these may seem rather far-fetched theories, it is true that after the enzymes of bone have been destroyed, *in vitro* calcification will not take place in media containing organic sources of P, but only if the P is inorganic [Waldman, 14].

Robison was led to postulate a second mechanism on top of the phosphatase one when he found that certain procedures stopped calcification of hypertrophic cartilage *in vitro* but had no effect upon phosphatase activity [Robison *et al.*, 15]. The researches into this second mechanism have taken two courses: one into the enzyme aspect and the other a search for a specific calcifiable substance in epiphyseal cartilage (the local factor).

It must be explained at this point how calcification *in vitro* has been studied. The rachitic epiphyseal cartilage of the rat has been used; when removed and incubated in appropriate media, rapid calcification occurs in a manner that appears identical with natural healing *in vivo*. Thus it is not ossification that is being observed but calcification of cartilage. So far it has not proved possible to cause rachitic osteoid to calcify *in vitro* [Bloom, 16], and Bailie and Irving [17] give reasons for supposing that such calcification could not occur save in tissue culture.

### Enzyme systems in calcification

The role of glycogen in calcification appears to be significant. Only the hypertrophic cartilage cells contain it and they correspond closely with those that calcify. If the glycogen is removed with saliva, calcification will not occur, but can be restarted with glucose-1-phosphate [Marks *et al.*, Marks and

orr, 18]. Marks *et al.* think that glycogen has to do with the preparatory stages of inorganic salt deposition in cartilage and the influence of glucose-1-phosphate is consistent with its position in the glycolytic cycle. Cobb [19] confirmed the findings of Marks *et al.* with regard to glycogen, in that the cells showing the most rapid formation of calcifiable matrix contained phosphorylase and had contained glycogen; the glycogen might be concerned with the production of bone matrix or osteoid or the alteration of cartilage matrix prior to calcification. Follis [20] also found that the calcification of rachitic cartilage occurred in the matrix of cells containing glycogen. Libaum *et al.* [21] have found adenosine triphosphate (ATP) in epiphyseal cartilage and that Ca prevented its disappearance from incubated cartilage slices. These authors think it possible that ATP is the substrate for phosphatase postulated by Robison and that ATP, rather than glycogen, is the limiting factor in calcification, but this does not appear to be in line with the findings of DiStephano and Neuman [13].

Turning to the enzymes involved in calcification, here the position is by no means clear, since calcification goes on quite well in certain media when all enzymes have been inactivated, though again in certain other media, enzymes are needed. Putman and Yu [22] investigated the actions of various enzymic inhibitors. They stated that in the presence of phloridzin, calcification from inorganic salts was stopped. Phloridzin inhibits phosphorylase and prevents the formation of glucose-1-phosphate. The addition of glucose-1-phosphate or any other factors concerned with subsequent stages of the glycolytic cycle would allow calcification to proceed in the presence of phloridzin. Iodoacetate inhibits 1-3-diphosphoglyceric aldehyde dehydrogenase and blocked the calcifying power of all factors higher in the cycle and of inorganic P, but not that of 3-phosphoglycerate. Fluoride, which blocks enolase, inhibited calcification with inorganic P, 3-phosphoglycerate and all factors higher up in the cycle. From this it might appear that calcification chemistry follows the same pathway as found in other tissues. However, no substrate for phosphatase is

provided at the end of the cycle, the P reappears as inorganic P and, as McLean and Urist state, the system runs into a dead end. Although when P is supplied as  $\alpha$  or  $\beta$  glycerophosphate, phenyl phosphate or creatine phosphate, there is good calcification in the presence of all these inhibitors (and these esters are all acted on by phosphatase) they are not in the main pathway of glycogenolysis.

Furthermore, calcification can still occur when enzymes are destroyed. Waldman [14], using a variety of methods which inactivated all enzymes, found that calcification would still proceed from inorganic phosphate media, but no longer from organic ester sources. He therefore concluded that enzyme systems were not necessary for calcification. Hiatt *et al.* [23] also found that certain inhibitors inactivated calcification from organic but not inorganic sources (though heat prevented calcification from inorganic phosphate) and were puzzled by the role of inorganic P which, while appearing to traverse the glycolytic cycle, could still cause calcification when the cycle was inactivated. However, the inhibitors used would, in higher concentration, also stop calcification from inorganic sources and they concluded that under the condition of their experiments calcification was dependent on an enzyme system. Goldenberg and Sobel [24], arguing from the results of F and cyanide inhibition, also considered the mechanism of calcification from both inorganic and organic media to be the same and not two discrete processes. If the glycolytic enzyme system does play a part in calcification, which it may well do in the living organism, it may be concerned with moving phosphate across cell membranes, or against concentration gradients, and the high-energy phosphate bonds formed may play some important role in the process. This is at present pure speculation, and only further research will give us the answer.

Ca has not been mentioned as playing a part in the above processes. It is, however, involved with the 'local factor' which is supposed to be present in calcifiable cartilage.

*The local factor.* There is now strong evidence that the substance which takes up Ca is a mucopolysaccharide and prob-



poly chondroitin sulphate. Thus Boyd and Neuman [25] considered that the binding of Ca by cartilage was connected with its sulphate content, implicating chondroitin sulphate. They further stated that phosphate was taken up only by cartilage which contained an appreciable amount of Ca. Bélanger [26], on auto-radiographic evidence, suggested that there was a relationship between the local distribution of Ca in cartilage calcified after decalcification and that of labelled sulphate.

Sobel and his colleagues have done much work, especially on the reversibility of calcification. They found that many ions such as Sr inactivated calcification *in vitro* [Sobel *et al.*, Sobel and Hanok, 27], but that this inactivation could be reversed by shaking the cartilage in Ca-containing solutions. They considered there was a competition for the constituent necessary for calcification which in their opinion was chondroitin sulphate and that the first step in calcification was a combination with Ca. In a later paper [Sobel and Burger, 28] they used a synthetic collagen chondroitin sulphate complex. This took up Ca like rachitic cartilage to the extent of an ash content of 50 per cent. dry weight and the deposited salt had an apatite diffraction pattern. They considered that a Ca complex first formed which reacted with phosphate to form an apatite salt and the process was then repeated. This view was supported by the finding that mineralization only occurred if the complex was shaken first with Ca salts and then with phosphate, but not in the reverse order.

DiStephano *et al.* [29], using chromatographic analysis, found a new spot in material from rachitic rat cartilage which appeared to be a stable ester of hexosamine. Procedures which stopped calcification caused the disappearance of this substance which they considered acted as a template for crystallization of the inorganic material during calcification.

Evidence of another kind also supports this concept. Histologically, mucopolysaccharides show metachromasia and there is much evidence that this staining reaction is strong where calcification is going to occur. Sylvén [30] and Levine *et al.* [31] considered the metachromatic and basiphilic staining material



in osteoid, bone and cartilage to be acid mucopolysaccharide, almost certainly chondroitin sulphate. Rubin and Howard [32] found the metachromasia to increase in areas of cartilage about to become calcified and in areas about to become bone. This suggests either the formation of a new mucopolysaccharide or a change in that already there, conferring calcifiability on the matrix. Howard [33] showed that the disappearance of metachromasia and of the periodic acid-Schiff reaction with calcification was due to masking with bone salts, and these reactions could be made to reappear if careful decalcification was carried out. Wislocki and Sognnaes [6] also found metachromasia and basiphilic staining in dentin and enamel, indicative of the presence of an acid mucopolysaccharide.

As far as the specificity of these staining reactions in bone is concerned, Heller-Steinberg [34] has sounded a note of caution, as minor variations in technique could change the sites of metachromasia or intensify them. All acid radicles in a tissue can be made to react metachromatically and decalcification increases metachromasia. She does not feel that under these circumstances metachromasia necessarily indicates the presence of chondroitin sulphate.

Thus in the case of bone, the possibility of a template action of chondroitin sulphate is not yet certain. The sulphate content of the organic matter of cartilage is at least six to eight times that of bone [Logan, 35], and Rogers [36] reported very small amounts of mucopolysaccharide in ox cortical bone. It is, however, of interest that Sobel *et al.* [37] have found after demineralization of bone with versene that the bone can be recalcified by shaking first in a  $\text{CaCl}_2$  solution and then with phosphate.

Another type of evidence supports the concept of acid mucopolysaccharide acting as a template in cartilage. Miller *et al.* [38] found that preliminary exposure of the cartilage to metachromatic or other basic dyes inhibited subsequent calcification in a calcifying medium. But if both Ca and phosphate were present with the dye, no inhibition occurred; if Ca or P only were present, then the dye inhibited. They concluded

at the dye reacted with an acid mucopolysaccharide essential for calcification, probably chondroitin sulphate. Incidentally, while the dye inhibited calcification from inorganic media, it did not stop calcification from organic sources. Sobel and Burger [39] have also reported the inhibitory nature of these dyes.

Exactly how all these results fit in with the enzymic system at present not clear. The fact that the dye inhibition does not affect calcification from organic media seems to suggest even more strongly that calcification from inorganic and organic sources are by means of different processes, or that calcification caused by the glycolytic cycle does not involve chondroitin sulphate. In spite of all these queries, it must be admitted that there has been a great advance since Robison's original work, and much of the calcification mechanism is now understood.

A final word should be said about the citrate in bone. Dickens in 1941 [40] showed that over two-thirds of the citrate in the body was in bone. In the previous chapter mention was made of its possible position in the apatite crystal. What exact role citrate plays in bone is not known. Dixon and Perkins [41] found that enzymes existed in bone capable of causing a high citrate content. The amount of citrate in different parts of bone was in inverse ratio to the metabolic activity of these parts. Parathyroidectomy caused a slow fall in bone citrate, but had no effect upon the bone citrate content, and the lowering of the blood Ca was not due to changes in the enzyme activity of the bone [42]. Steenbock and Bellin [43] reported that physiological doses of vitamin D increased blood and bone citrate (and that in several other tissues); bone citrate was not reduced in rickets, but seemed to have some relation with the Ca status of the animal. Freeman and Chang [44] found that vitamin D in large doses produced a parallel change in plasma Ca and citric acid, suggesting it influences the metabolism of both substances. It has also been reported that parathyroid hormone mobilizes citrate together with Ca. Thus citrate appears to play some active part in bone metabolism.

## REFERENCES

1. ROBISON, R. *Biochem. J.*, **17**, 286 (1923)
2. LORCH, I. J. *Quart. J. micr. Sci.*, **88**, 367 (1947)
3. SIFFERT, R. S. *J. exp. Med.*, **93**, 415 (1951)
4. BEVELANDER, G. and JOHNSON, P. L. *J. dent. Res.*, **29**, 665 (1950)
5. MORSE, A. and GREEP, R. O. *Anat. Rec.*, **111**, 193 (1950)
6. WISLOCKI, G. B. and SOGNAES, R. F. *Amer. J. Anat.*, **87**, 239 (1950)
7. BEVELANDER, G. and JOHNSON, P. L. *Anat. Rec.*, **107**, 125 (1949)
8. JOHNSON, P. L. and BEVELANDER, G. *J. dent. Res.*, **33**, 128 (1954)
9. DANIELLI, J. F., FELL, H. B., and KODICEK, E. *Brit. J. exp. Path.*, **26**, 367 (1945)
10. GOULD, B. S. and SHWACHMAN, H. *Amer. J. Physiol.*, **135**, 485 (1942)
11. NORRIS, W. P. and COHN, S. H. *J. biol. Chem.*, **196**, 257 (1952)
12. NEUMAN, W. F., DISTEPHANO, V. and MULRYAN, B. *J. biol. Chem.*, **193**, 227 (1951)
13. DISTEPHANO, V. and NEUMAN, W. F. *J. biol. Chem.*, **200**, 759 (1953)
14. WALDMAN, J. *Proc. Soc. exp. Biol. N.Y.*, **69**, 262 (1949)
15. ROBISON, R., MCLEOD, M. and ROSENHEIM, A. H. *Biochem. J.*, **24**, 1927 (1930)
16. BLOOM, W. Trans. 2nd Conf. metab. Interr. New York Josiah Macy Jr. Found., p. 216 (1950)
17. BAILIE, J. M. and IRVING, J. T. *Acta med. Scand.*, **153**, Supp. 306, 1 (1955)
18. MARKS, P. A., HIATT, H. H. and SHORR, E. *J. biol. Chem.*, **204**, 175 (1953)
- MARKS, P. A. and SHORR, E. *Science*, **112**, 752 (1950)
19. COBB, J. D. *Arch. Path.*, **55**, 496 (1953)
20. FOLLIS, R. H. *Proc. Soc. exp. Biol. N.Y.*, **71**, 572 (1949)

- ALBAUM, H. G., HIRSHFELD, A. and SOBEL, A. E. *Proc. Soc. exp. Biol. N.Y.*, **79**, 238 (1952)
- GUTMAN, A. B. and YU, T. B. Trans. 2nd Conf. metab. Interr. New York, Josiah Macy Jr. Found. (1950)
- HIATT, H. H., MARKS, P. A. and SHORR, E. *J. biol. Chem.*, **204**, 187 (1953)
- GOLDENBERG, H. and SOBEL, A. E. *Proc. Soc. exp. Biol. N.Y.*, **85**, 275 (1954)
- BOYD, E. S. and NEUMAN, W. F. *J. biol. Chem.*, **193**, 243 (1951)
- BÉLANGER, L. F. *Proc. Soc. exp. Biol. N.Y.*, **88**, 150 (1955)
- SOBEL, A. E., NOBEL, S. and HANOK, A. *Proc. Soc. exp. Biol. N.Y.*, **72**, 68 (1949)
- SOBEL, A. E. and HANOK, A. *J. biol. Chem.*, **197**, 669 (1952)
- SOBEL, A. E. and BURGER, M. *Proc. Soc. exp. Biol. N.Y.*, **87**, 7 (1954)
- DISTEPHANO, V., NEUMAN, W. F. and ROUSER, G. *Arch. Biochem. Biophys.*, **47**, 218 (1953)
- SYLVÉN, J. *J. Bone Joint Surg.*, **29**, 973 (1947)
- LEVINE, M. D., RUBIN, P. S., FOLLIS, R. H. and HOWARD, J. E. Trans. 1st Conf. metab. Interr. New York, Josiah Macy Jr. Found., p. 41 (1949)
- RUBIN, P. S. and HOWARD, J. E. Trans. 2nd Conf. metab. Interr. New York, Josiah Macy Jr. Found., p. 155 (1950)
- HOWARD, J. E. *Bull. New York Acad. Med.*, **27**, 24 (1951)
- HELLER-STEINBERG, M. *Amer. J. Anat.*, **89**, 347 (1951)
- LOGAN, M. A. *J. biol. Chem.*, **110**, 375 (1935)
- ROGERS, H. J. *Biochem. J.*, **49**, xii (1951)
- SOBEL, A. E., SAMACHSON, J. and SLOVEK, N. *Fed. Proc.*, **14**, 283 (1955)
- MILLER, Z. B., WALDMAN, J. and MCLEAN, F. C. *J. exp. Med.*, **95**, 497 (1952)
- SOBEL, A. E. and BURGER, M. *Proc. Soc. exp. Biol. N.Y.*, **87**, 7 (1954)
- DICKENS, F. *Biochem. J.*, **35**, 1011 (1941)



41. DIXON, T. F. and PERKINS, H. R. *Biochem. J.*, **52**, 26 (1952)
42. PERKINS, H. R. and DIXON, T. F. *Science*, **118**, 139 (1953)
43. STEENBOCK, H. and BELLIN, S. A. *J. biol. Chem.*, **20**, 985 (1953)
44. FREEMAN, S. and CHANG, T. S. *Amer. J. Physiol.*, **16**, 341 (1950)

## CHAPTER XIII

# The Calcium Metabolism of the Teeth

THE teeth consist of three different calcified tissues: enamel, dentin and cementum. The bulk of the tooth consists of dentin, the crown is covered by enamel, and the roots by cementum.

Enamel is the hardest tissue in the body, containing 1.2 to 2.0 per cent. water, 0.2 to 0.8 per cent. organic matter and 95 to 97 per cent. inorganic matter [1]. One analysis gives the Ca content of the ash to be 39.98 per cent. and the P content 4.42 per cent. [Murray, 2]. Sobel *et al.* [3] and Trautz *et al.* [4] find that the calcified elements show an X-ray diffraction pattern of apatite. The protein of enamel is enamel-keratin [Hess *et al.*, 5].

Dentin is a more elastic structure. It contains 10.8 to 15.7 per cent. water, 20.3 to 22.4 per cent. organic matter and 61 to 65 per cent. inorganic matter [1]. Murray gives the Ca and P contents of the ash as 39.83 and 19.04 per cent., respectively. The mineral matter has the apatite pattern. The protein of dentin is a collagen [Hess *et al.*, 6]. Burnett and Scherp [7] tested the 'accessibility' of the protein by coupling it with diazonium compounds and found the protein unreactive till calcification was begun, when the amount of azo-protein formed was proportional to the weight loss.

Cementum is chemically and physiologically like bone. Zipkin and Piez [8] found citrate in both enamel and dentin; the dentin contained around 0.9 per cent., about eight times as much as in enamel; very insoluble in both tissues.

The formation of enamel and dentin during development of the tooth are quite different processes. The enamel is formed

by the enamel organ, a complicated structure in which the active cells are the ameloblasts. An organic matrix is laid down first, and is gradually thickened by increments to its final width. As the matrix becomes mature it is calcified, and while the Ca and P increase the moisture and organic part decrease [Weinmann *et al.*, 9]. Deakins [10] found that Ca, P and CO<sub>2</sub> contents increased linearly and in a constant ratio as the enamel matured, a five times increase in inorganic content occurring from softest to hardest enamel. Water and inorganic material exchanged in equal proportions and the final calcification of enamel was not just a 'drying up'. As a result of the high degree of mineralization of enamel, the fully calcified tissue is lost in decalcified histological preparations unless special precautions are taken to preserve the fine organic matrix. The processes of matrix formation and calcification appear to be closely linked, even at an early stage, since interference with calcification stops matrix formation (Irving, 11) which is not true of bone or dentin. Some workers [e.g. Nuckolls *et al.*, 12] have divided the calcification process into primary and secondary stages, but this seems to the present writer unnecessarily complicated.

How the enamel is transformed into its final form is still not understood, but it appears certain that this must be carried out by the enamel organ only and not partly by the pulp as claimed by some. If the ameloblasts are damaged the matrix does not mature properly [Chase, 13; Weinmann, 14; Irving, 11] and it remains acid insoluble in histological preparations. Thus the enamel organ has probably one of the most diverse roles of any collection of cells in the body: it forms matrix, calcifies it, removes water and organic material, and in the case of some rodents, finally lays down a pigment layer on the incisor tooth.

Dentin is formed by cells lining the pulp, the odontoblasts. They probably change the material of the Korff fibres, formed in the pulp, which pass into the dentin between them, into a collagenous matrix, which they then calcify. Calcification is normally delayed for a definite period, twenty-four hours in

In the case of the rat incisor, the uncalcified matrix being called predentin, staining with eosin in decalcified sections, the fully calcified dentin taking up haematoxylin. This change in staining reactions which accompanies calcification has, as in the case of bone, been attributed to some chemical change in the matrix [Weinmann and Sicher, 15].

Three types of material have been studied: fully erupted teeth, teeth before eruption, and continually erupting teeth of animals such as rats. The methods have been chemical, physical, using X-ray (Grenz) analysis, and histological, both conventional and radio-autographic, with  $\text{Ca}^{45}$ ,  $\text{P}^{32}$ , and other isotopes. Ground sections are often employed to avoid enamel loss during decalcification, but do not show the finer detail in cellular and matrix structure. Although this monograph is primarily about Ca, it is impossible to avoid mentioning results obtained with P, since the two elements are so similarly treated.

It should be appreciated that the fully formed and erupted tooth is outside almost all vital influences of the body (with the exception of a few processes such as the formation of secondary dentin), and thus metabolically should be very inactive, this applying especially to enamel. Experimental results support this supposition. Thus Hevesy *et al.* [16] calculated that the human tooth exchanged about 1 per cent. of the dietary P every 250 days. Hevesy and Armstrong [17] likewise reported that the enamel of the permanent and erupted teeth of cats took up little injected radio-P, about one-tenth of that of dentin. Volker and Sognnaes [18] gave radio-P to an adult rat by stomach tube; they detected 4.4 per cent. of the dose in the teeth of which 91 per cent. was in the dentin and 2.3 per cent. in the enamel. Barnum and Armstrong found the same [19] and considered that pathways existed for  $\text{P}^{32}$  to go from the saliva to the dentin via the enamel, and from the blood to the enamel via the dentin. The main source of  $\text{P}^{32}$  to the teeth was from the blood-supply to the pulp. They found that the P of rat molars was not completely removed by exchange after 116 days. In confirmation of the importance of the blood-supply, Gilda *et al.* [20] reported pulpless teeth



to take up much less  $P^{32}$ . Sognnaes and Shaw [21] have studied the uptake of injected radio-P in enamel and dentin of the erupted teeth of monkeys and found that there was a gradient in concentration of  $P^{32}$ , in the case of enamel falling from the exterior to the interior parts and rising from the internal to the external (nearest the pulp) dentin. They attributed these uptakes to salivary  $P^{32}$  in the case of enamel and to the blood radio-P in the case of dentin. They also studied unerupted enamel [22] and considered that the very large uptake by this tissue was due to a high rate of exchange plus an active accretion of P accompanying the final growth of submicroscopic crystals.

*In vitro* work has shown that, as with bone, powdered dentin and enamel will adsorb  $P^{32}$ . Enamel has a slower rate than dentin and both have less adsorbing power than bone, possibly due to differences in size of the mineral crystal [Hodge *et al.* 23]. Thus in the erupted tooth the uptake of  $P^{32}$  must, as in the case of fully mature bone, be largely a process of exchange and possibly recrystallization.

Much work has been done using the continually erupting incisor tooth of the rat. This, as would be expected from a continually calcifying structure, takes up radio-Ca and P with great readiness. Manly *et al.* [24] considered that there was a labile part of the tooth in equilibrium with blood  $P^{32}$  and a stable fraction building up radio-P incrementally. In the rat incisor, 73 per cent. of the  $P^{32}$  was in the labile fraction after one day, and 3 per cent. at twenty days after dosage. Carlsson [25], using  $Ca^{45}$ , found that the content of the isotope in the incisor rose continually up to twenty-four days after administration. But this interstitial metabolism is, in the case of dentin, very low compared to that of bone, although the  $Ca^{45}$  returned to the circulation with the resorption of bone giving a persistent activity for up to ten weeks [Tomlin *et al.*, 26]. Armstrong [27] found in mature rats that the incisal uptake of radio-Ca and P given together was about equal in enamel and dentin.

The microscopic details of the incorporation of  $P^{32}$  into

Enamel and dentin have been studied by Bélanger using radioautographs [28]. In the case of enamel, a wide band of radio-P was seen at the dentino-enamel junction at the formative end of the tooth (in the transitional enamel stage), but not where the enamel was first deposited, showing that newly formed matrix has to reach a certain stage of maturation before it is receptive to mineral deposition, this stage being reached in a short time after formation. During the process of general mineralization, the enamel took up less and less  $P^{32}$ , and finally, after optimal saturation, no more. In dentin, a thin radioactive line appeared near the pulp and slowly approached the dentino-enamel junction, and eventually disappeared, leaving behind a diffuse distribution of radioactivity. Mineralization of dentin occurred in a series of waves, and the  $P^{32}$  became incorporated, by dissolution or reconstruction of existing crystals, into more stable crystals.

The relationship between mucopolysaccharides and calcification appears to be similar in teeth to that in bone. Pincus [29] found chondroitin sulphate in dentin, and Rogers [30] stated that there was 0.1 per cent. hexosamine and 3.5 per cent. of total reducing substances in dentin. Hess and Lee [31] reported 64 per cent. of chondroitin sulphate in dentin, analytically the same as in cartilage. Thus there is little doubt of the existence of this substance in dentin. Engel [32] found histologically a glycoprotein in many parts of the enamel organ, and also in ameloblasts and odontoblasts in the early stages of tooth development prior to the formation of enamel and dentin. Wislocki and Sognnaes [33] found an acid mucopolysaccharide in enamel and dentin. Both Burstone [34] and Cleveland and Johnson [35] reported a relationship between metachromasia and calcification in dentin, the latter workers stating that metachromasia gradually disappeared as calcification proceeded. Greulich and Le Blond [36], using  $C^{14}$ , detected what was probably a carbohydrate which appeared at the predentin-dentin junction at the same time as mineral crystals. They thought the crystals were bound to the collagen matrix by means of this carbohydrate. Bélanger [37], working

with radio-S, found this isotope in the ameloblasts and new enamel during the growth period. This, he stated, was synthesized to an acid mucopolysaccharide which appeared in the matrix before keratin and disappeared progressively. Thus there is a good deal of evidence that, as with bone, acid mucopolysaccharides are involved in the calcification of enamel and dentin, and they may act as a template for the crystallization of the apatite structure. It is also of interest that Engel [32] found glycogen in many parts of the developing tooth and associated structures.

The studies of dental enzymes have centred chiefly round alkaline phosphatase which is found in many parts of the developing tooth. In view of what has been said above about the functions of this enzyme, it is doubtful, at any rate in the opinion of the present writer, whether it has any direct connexion with the calcification of enamel and dentin, though it has been found in ameloblasts and odontoblasts while enamel and dentin were being elaborated [Johnson and Bevelander, 38]. It has also been demonstrated in dentinal fibres and enamel prisms and very abundant in the growing pulp [Wislocki and Sognnaes, 33]. Symons [39] reported the appearance of phosphatase in areas where enamel would appear, but the odontoblasts had less than the adjacent pulp cells. The finding of phosphatase in the pulp, where the Korff fibres are formed lends support to the view that the enzyme is more concerned with matrix formation.

### **The influence of diet and endocrine factors upon the teeth**

Leaving aside the production of caries, which is undoubtedly dietary in origin, the fully erupted tooth is outside the influence of dietary factors, but the developing tooth and the continually erupting incisors of rodents are very sensitive to dietary changes. Sobel and Hanok [40], working with rat incisors, have reported that, as with bone, the  $\text{PO}_4 : \text{CO}_2$  ratios of enamel and dentin are related to those of the blood serum and to those of the diet.

Ca, P or vitamin D deficiency, or parathyroidectomy, al

have similar effects on dentin formation in continuously erupting incisors, and Erdheim in 1906 was the first to report the changes after parathyroidectomy. Histologically, the predentin becomes wider, often with vascular inclusions, and the calcified dentin is irregular and arranged in small calcospherites (interglobular dentin). Becks and Ryder [41] were among the earliest to report this condition in low P rickets. The Ca : P ratio of the diet is important in this respect. Gaunt and Irving [42] found that diets of high Ca : P ratio affected the bones more than the teeth, but if the ratio was low, the reverse occurred, and the dentin ash was reduced. When vitamin D is given to rats with low P rickets, the newly formed predentin is properly calcified, but that existing before the vitamin was given remains unchanged [Irving, 43]. This illustrates another difference between bones and teeth, the latter having no reparative power. They also have no resorptive mechanism and in Ca or P deficiency the pre-existing dental tissue is unaffected.

Several factors causing sudden changes in dentin calcification produce a similar histological picture which has been analysed by Irving and Weinmann [44] and called the calcio-traumatic response. The last formed dentin becomes hypercalcified, the existing predentin is not calcified at all, and the next formed dentin is hypercalcified for a short time. This is sometimes seen in the deciduous teeth of children caused by the adventure of birth, and called the neonatal line [Rushton, Schour, 45]. It is also caused by injections of fluorine [Schour and Smith, 46], large doses of calciferol or parathyroid hormone [Schour and Ham, 47; Schour *et al.*, 48], strontium injections [Irving and Weinmann, 44] and nephrectomy [Irving *et al.*, 49]. This response appears to be a non-specific reaction to changes in the Ca and P environment of the tooth. Applebaum [50] has shown by Grenz-ray analysis that the areas appearing histologically hypo- and hypercalcified are in fact radiolucent and radiopaque. The reasons for these changes are not understood, but they may be due to alterations in the calcifiability of the matrix.



Hypoplasia of the enamel in humans is usually caused by systemic disturbances during enamel matrix formation, too little enamel being formed [Diamond and Weinmann, 51] which may or may not be hypocalcified. In amelogenesis imperfecta, a rare congenital condition, the enamel matrix forms but calcifies incompletely.

Several vitamins besides vitamin D have characteristic protective effects upon the tooth, but these probably affect matrix formation rather than calcification. In vitamin A deficiency dentin formation is upset, the odontoblasts do not regress as they should and excessive dentin is formed on the labial side of the incisor tooth [Wolbach and Howe, Schour *et al.*, 52]. Enamel formation is deranged and the incisor loses its pigment [Irving and Richards, 53]. The vitamins of the B group do not appear to influence tooth formation. In scurvy a characteristic change occurs, the formation of dentin ceasing and the last part formed being sealed off from the pulp by a hypercalcified zone [Fish and Harris, 54]. The odontoblasts lose their ability to make a calcifiable matrix and change to a structure like the *gerüstmark* of bone [Irving and Boyle, 35]. Both vitamin A and C thus affect tooth calcification as well as that of bone, and in a similar way, but little is known of the biochemical changes underlying these effects. Lysine or tryptophane deficiency causes a change in the dentin very similar to that seen in C or P lack [56].

Some of the endocrine glands besides the parathyroids affect tooth formation, causing changes in calcification, matrix formation or the growth sequences. Hypophysectomy produces a great retardation in the eruption rate, and as a result the rat's incisor has a folded appearance at the formative end [Becks *et al.*, 57], but dentin apposition continues so that the pulp cavity is almost obliterated [Schour and van Dyke, 58]. The calcification of the tooth is not changed. Thyroidectomy retards the calcification of dentin [Ziskin and Applebaum, 59] which becomes virtually uncalcified matrix, as shown by Grenz-ray analysis [Ziskin and Applebaum, 60]. Adrenalectomy and gonadectomy interfere with dentin calcification.

61, 62], but oestrogen administration has no effect on rat or mice teeth, which is of interest when its effects on bone are recalled [63].

One can conclude that while the fully formed and erupted tooth is outside influences exerted through the body, the forming tooth is very sensitive to changes in Ca and P metabolism and is under the control of several vitamins and most endocrine glands. It has been found that the dentin of rachitic rats responds exponentially to vitamin D dosage in the same way as does the rachitic epiphysis [Irving, 43], and also that the dental changes after vitamin D dosage could be detected after twenty-four hours, four days before any change could be seen in the bones. The teeth have been largely neglected by physiologists, which is to be regretted since they react so quickly and specifically to many diverse influences.

## REFERENCES

1. HODGE, H. C. Quoted in NOYES, F. B., SCHOUR, I. and NOYES, H. J. *A textbook of dental histology and embryology*. London: Henry Kimpton (1938)
2. MURRAY, M. M. *Biochem. J.*, **30**, 1567 (1936)
3. SOBEL, A. E., HANOK, A., KERSHNER, H. A. and FANKUCHEN, I. *J. biol. Chem.*, **179**, 205 (1949)
4. TRAUTZ, O. R., KLEIN, E., ADDELSTON, H. K. and FANKUCHEN, I. *J. dent. Res.*, **30**, 478 (1951)
5. HESS, W. C., LEE, C. Y. and NEIDIG, B. A. *J. dent. Res.*, **32**, 585 (1953)
6. HESS, W. C., LEE, C. Y. and NEIDIG, B. A. *J. dent. Res.*, **31**, 791 (1952)
7. BURNETT, G. W. and SCHERP, H. W. *J. dent. Res.*, **29**, 663 (1950)
8. ZIPKIN, I. and PIEZ, K. A. *J. dent. Res.*, **29**, 498 (1950)
9. WEINMANN, J. P., WESSINGER, G. D. and REED, G. *J. dent. Res.*, **20**, 244 (1941)
10. DEAKINS, M. and BURT, R. L. *J. biol. Chem.*, **156**, 77 (1944)  
DEAKINS, M. *J. dent. Res.*, **21**, 429 (1942)

11. IRVING, J. T. *Brit. J. exp. Path.*, **31**, 458 (1950)
12. NUCKOLLS, J., SAUNDERS, J. B. DE C. and FRISBIE, H. *J. dent. Res.*, **22**, 210 (1943)
13. CHASE, S. W. *Proc. dent. Centen. Celebr. (Baltimore)* p. 425 (1940)
14. WEINMANN, J. P. *J. Amer. dent. Ass.*, **30**, 874 (1943)
15. WEINMANN, J. P. and SICHER, H. *Bone and bones*. St. Louis: The C.V. Mosby Company (1955)
16. HEVESY, G., HOLST, J. J. and KROGH, A. *K. dansk vidensk. Selsk. Biol. Middel.*, **13**, 13 (1937)
17. HEVESY, G. VON and ARMSTRONG, W. D. *J. dent. Res.*, **19**, 318 (1940)
18. VOLKER, J. F. and SOGNAES, R. F. *J. dent. Res.*, **19**, 29 (1940)
19. BARNUM, C. P. and ARMSTRONG, W. D. *Amer. J. Physiol.*, **135**, 478 (1942)
20. GILDA, J. E., MCCAULEY, H. B. and JOHANSSON, E. *J. dent. Res.*, **22**, 200 (1943)
21. SOGNAES, R. F. and SHAW, J. H. *J. Amer. dent. Ass.*, **4**, 489 (1952)
22. SOGNAES, R. F., SHAW, J. H. and BOGOROCH, R. *Amer. J. Physiol.*, **180**, 408 (1955)
23. HODGE, H. C., VAN HUYSEN, G., BONNER, J. F. and VAN VOORHIS, S. N. *J. biol. Chem.*, **138**, 451 (1941)
24. MANLY, R. S., HODGE, H. C. and MANLY, M. L. *J. biol. Chem.*, **134**, 293 (1940)
25. CARLSSON, A. *Acta pharm. tox. Kbh.*, **7**, Supp. 1 (1951)
26. TOMLIN, D. H., HENRY, K. M. and KON, S. K. *Brit. Nutr.*, **9**, 144 (1955)
27. ARMSTRONG, W. D. *J. dent. Res.*, **24**, 192 (1945)
28. BÉLANGER, L. F. *Anat. Rec.*, **114**, 529 (1952)
29. PINCUS, P. *Proc 1st internat. Congr. Biochem.*, p. 206 (1949)
30. ROGERS, H. J. *Nature, Lond.*, **164**, 625 (1949)
31. HESS, W. C. and LEE, C. *J. dent. Res.*, **31**, 793 (1952)
32. ENGEL, M. A. *J. dent. Res.*, **27**, 681 (1948)
33. WISLOCKI, G. B. and SOGNAES, R. F. *Amer. J. Anat.*, **87**, 239 (1950)

4. BURSTONE, M. S. *J. dent. Res.*, **32**, 269 (1953)
5. BEVELANDER, G. and JOHNSON, P. L. *J. dent. Res.*, **34**, 123 (1955)
6. GREULICH, R. C. and LE BLOND, C. P. *J. dent. Res.*, **33**, 859 (1954)
7. BÉLANGER, L. F. *J. dent. Res.*, **34**, 20 (1955)
8. JOHNSON, P. L. and BEVELANDER, G. *J. dent. Res.*, **33**, 128 (1954)
9. SYMONS, N. B. B. *J. Anat. Lond.*, **89**, 238 (1955)
0. SOBEL, A. E. and HANOK, A. *J. biol. Chem.*, **176**, 1103 (1948)
1. BECKS, H. and RYDER, W. B. *Arch. Path.*, **12**, 358 (1931)
2. GAUNT, W. E. and IRVING, J. T. *J. Physiol.*, **99**, 18 (1940)
3. IRVING, J. T. *J. Physiol.*, **103**, 9 (1944)
4. IRVING, J. T. and WEINMANN, J. P. *J. dent. Res.*, **27**, 669 (1948)
5. RUSHTON, M. A. *Dent. Rec.*, **53**, 170 (1933)  
SCHOUR, I. *J. Amer. dent. Ass.*, **23**, 1946 (1936)
6. SCHOUR, I. and SMITH, M. C. *Univ. Arizona agric. exp. Stat. Tech. Bull.*, **52**, 69 (1934)
7. SCHOUR, I. and HAM, A. W. *Arch. Path.* **17**, 22 (1934)
8. SCHOUR, I., TWEEDY, W. R. and MCJUNKIN, F. A. *Amer. J. Path.*, **10**, 321 (1934)
9. IRVING, J. T., WEINMANN, J. P., SCHOUR, I. and TWEEDY, W. R. *J. dent. Res.*, **28**, 356 (1949)
0. APPLEBAUM, E. *J. dent. Res.*, **22**, 7 (1943)
1. DIAMOND, M. and WEINMANN, J. P. *The enamel of human teeth*. New York: Columbia University (1940)
2. WOLBACH, S. B. and HOWE, P. R. *J. exp. Med.*, **42**, 753 (1925)  
SCHOUR, I., HOFFMAN, M. M. and SMITH, M. C. *Amer. J. Path.*, **17**, 529 (1941)
3. IRVING, J. T. and RICHARDS, M. B. *Nature, Lond.*, **144**, 908 (1939)
4. FISH, E. W. and HARRIS, L. J. *Phil. Trans. Roy. Soc., B*, **223**, 489 (1934)



55. IRVING, J. T. and BOYLE, P. E. *J. dent. Res.*, **31**, 50 (1952)
56. BAVETTA, L. A. and BERNICK, S. *J. Amer. dent. Ass.*, **5**, 427 (1955)
- BAVETTA, L. A., BERNICK, S., GEIGER, E. and BERGREN, W. *J. dent. Res.*, **33**, 309 (1954)
57. BECKS, H., COLLINS, D. A., SIMPSON, M. E. and EVANS, H. M. *Arch. Path.*, **41**, 457 (1946)
58. SCHOUR, I. and VAN DYKE, H. B. *Amer. J. Anat.*, **50**, 39 (1932)
59. ZISKIN, D. E. and APPLEBAUM, E. *J. dent. Res.*, **19**, 30 (1940)
60. ZISKIN, D. E. and APPLEBAUM, E. *J. dent. Res.*, **20**, 2 (1941)
61. SCHOUR, I. and ROGOFF, J. M. *Amer. J. Physiol.*, **115**, 334 (1936)
62. SCHOUR, I. *Anat. Rec.*, **65**, 177 (1936)
63. STAHL, S. S., WEINMANN, J. P., SCHOUR, I. and BUDY, A. M. *Anat. Rec.*, **107**, 21 (1950)

## CHAPTER XIV

# The Excretion of Calcium

Ca is lost from the body by three ways: from the gut, the kidneys and the skin. The latter is the least important but a by no means insignificant route. Mitchell and Hamilton [1] found sweat to contain 2-7 mg. per 100 ml., and that even under minimal sweating conditions the daily loss could be 149 mg. Four to 14 mg. per hour could be lost at high temperatures [Johnston *et al.*, 2].

The mechanism of Ca excretion by the kidney has been investigated by a number of workers. The constant presence of Ca in the urine had been explained by Albright and Ellsworth [3] as due to the threshold for Ca being normally 8.5 mg. per 100 ml. serum, so that with a value of 10 mg. the threshold would be always exceeded. This explanation is doubted by Robertson [4], who found the usual level of Ca excretion in a patient with a serum Ca level of 5.5 mg. per 100 ml. Wolf and Ball [5] thought there was no renal threshold for Ca and that the plasma Ca level was not regulated by renal function. In a series of important papers, Chen [6] and Chen and Neuman [7] have reported findings on the handling of Ca by the renal tubule. The Ca percentage in the glomerular filtrate was almost the same as that in an ultrafiltrate of serum (the ionized Ca fraction), reabsorption was an active process and over 99 per cent. of that filtered was reabsorbed. The small but constant amount of Ca excreted was considered to be an ionized complex, since the Ca clearance was increased by the injection of phosphate or versene. Chang and Freeman [8] also found that the injection of citrate greatly increased urinary excretion. If this concept is true it suggests that the kidney

is very efficient at handling ionized Ca and that the Ca is in a form that the tubules cannot absorb.

The process of Ca reabsorption is different from that which acts on Na or K [Chen and Neuman, 7]. Ca reabsorption is depressed by phloridzin, dinitrophenol or Na azide, none of which affects Na or K reabsorption. On the other hand, diazotized urea (a carbonic anhydrase inhibitor) increases Na and K excretion but has no effect on that of Ca. Phloridzin and dinitrophenol inhibit phosphorylation reactions and the generation of high energy phosphate bonds, and both they and Na azide interfere with the secretory activity of the tubules. Thus some kind of metabolic energy is needed for the reabsorption of Ca.

The percentage of total Ca excretion which appears in the urine has been stated by some investigators to be under normal conditions fairly constant, about 24 per cent. [Bauer *et al.*, 3; Robertson, 4]. Nicolaysen *et al.* [10] in a long study of over 100 men and women found the Ca in the urine, as a percentage of the intake, to be between 20 and 25 per cent. Knapp [11] likewise reported an average figure of 25 per cent., but a great variation in his group of subjects. While the urinary Ca excretion was constant within individuals, it varied considerably from person to person [10], and even in the same individual periods of change could occur for no apparent reason. Altering the Ca intake might or might not affect the urinary output. Seeing that the handling of Ca by the kidney is under the control of a number of endocrine factors, this variability is not surprising. Furthermore, the pH of the urine is a ruling factor since urine cannot act as a vehicle for Ca excretion if it is alkaline, though in man this does not normally occur.

After hypophysectomy the urinary Ca excretion falls in humans [Luft, 12], but this may not be due to the loss of the direct action of the pituitary since adrenocorticotrophic hormone, cortisone and desoxycorticosterone cause a negative Ca balance [Luft *et al.*, 13] and cortisone increases the urinary and faecal Ca [Soffer *et al.*, Moehlig *et al.*, 14]. The thyroid has a marked action on Ca excretion in the urine and in hypothyroidism there is a considerable Ca loss [Aub *et al.*, 15].

Robertson [16] has proposed that the renal threshold for Ca be lowered, and that the excessive Ca loss causes by a *vis à* *vis* action a decalcification of bone. The parathyroids have been postulated to have a direct action on the kidney. This has been discussed in Chapter IX.

Before leaving this aspect of the subject, a word should be said on the renal action which has been claimed for vitamin D. Harrison and Harrison considered that the vitamin caused an increase in tubular reabsorption of P and a rise in serum Ca and P, A.T.10 and parathyroid hormone causing a fall in P absorption [17]. There seems to be some confusion about the effect of the reabsorption of P on the blood Ca level; if these results are true, both an increase and a decrease in this reabsorption accompany a rise in the blood Ca. Crawford *et al.* [18] consider that the results with vitamin D have been due to a suppression of parathyroid action since, if parathyroidectomy is carried out, vitamin D causes a decrease in tubular P reabsorption. This finding makes the renal action of vitamin D more consistent with that of the parathyroid. Cruickshank *et al.* [19] found, after giving a large dose of vitamin D, that a considerable amount was retained by the kidney, and they thought this might be connected with a tubular action.

The faecal Ca in man consists largely of unabsorbed food Ca. It was shown in Chapter III that, in addition, secretion of Ca into the gut undoubtedly occurs. Geissberger, Bellin and Szabo, and Staub [20] have injected  $\text{Ca}^{45}$  into humans and found the isotope to be rapidly eliminated in the faeces in amounts of from 2.5 to 15 per cent. of the dose. Geissberger stated that four days later there was more  $\text{Ca}^{45}$  in the faeces than in the urine.

From the evidence it seems legitimate to conclude that the human intestine can excrete Ca but may not do so always. The findings of Steggerda and Mitchell [21] are conclusive on this point. They gave citrate to men and reported that the faecal Ca might amount to over twice the Ca intake. It is true that in these experiments the conditions were unusual in that the urine might have been alkaline (the pH of the urines was not



determined), but none the less the fact was established that intestine could act as an excretory pathway. Whether it does this under normal conditions in any significant sense is possible but doubtful. McCance and Widdowson [22] gave humans gluconate intravenously and found no increase in faecal excretion. Johnson [23] and Welch *et al.* [24], studying patients with ileostomies and colostomies, found no evidence of Ca excretion in the gut.

Animal experimentation suggests that they too can excrete Ca in the faeces. Thus rats given  $\text{Ca}^{45}$  by injection excrete a considerable proportion of the dose in the faeces [Greenblatt, L'Heureux *et al.*, Wallace *et al.*, 25], the latter two groups considering that the small intestine played a major role. Rabbits have likewise been shown to excrete Ca in this way. Sjollesma [26] many years ago thought that Ca played a part in faeces formation in these animals. Cowell [27] caused uraemia in rabbits and as a result the faecal excretion exceeded the urinary plus food Ca. In herbivorous animals who habitually secrete an alkaline urine, faecal Ca excretion would be expected.

Other animals have been shown to excrete Ca by this route, e.g. cats [Stewart and Percival, 28] and dogs [Greenwald and Gross, 29].

From the evidence cited above it may be concluded that for man the kidney is the main excretory pathway, the Ca being lost possibly owing to the kidney tubule being unable to absorb unionized Ca complexes. The intestine is able to excrete Ca and forms a reserve mechanism for this purpose.

## REFERENCES

1. MITCHELL, H. H. and HAMILTON, T. S. *J. biol. Chem.*, **178**, 345 (1949)
2. JOHNSTON, F. A., MCMILLAN, J. T. and EVANS, E. *J. Nutr.*, **42**, 285 (1950)
3. ALBRIGHT, F. and ELLSWORTH, R. *J. clin. Invest.*, **7**, 1 (1929)

4. ROBERTSON, J. D. *Lancet*, **1**, 156 (1941)
5. WOLF, A. V. and BALL, S. M. *Amer. J. Physiol.*, **158**, 205 (1949)
6. CHEN, P. S. *Fed. Proc.*, **12**, 310 (1953)
7. CHEN, P. S. and NEUMAN, W. F. *Amer. J. Physiol.*, **180**, 623, 632 (1955)
8. CHANG, T. S. and FREEMAN, S. *Amer. J. Physiol.*, **160**, 330 (1950)
9. BAUER, W., AUB, J. C. and ALBRIGHT, F. *J. clin. Invest.*, **7**, 75 (1929)
10. NICOLAYSEN, R., EEG-LARSEN, N. and MALM, O. J. *Physiol. Rev.*, **33**, 424 (1953)
11. KNAPP, E. L. *J. clin. Invest.*, **26**, 182 (1947)
12. LUFT, R. *Recent Prog. Hormone Res.*, **10**, 460 (1954)
13. LUFT, R., SJÖGREN, B., IKKOS, D., LJUNGGREN, H. and TARUKOSKI, H. *Recent Prog. Hormone Res.*, **10**, 425 (1954)
14. SOFFER, L. J., GABRILOVE, J. L. and JAILER, J. W. *J. clin. Invest.*, **10**, 594 (1950)
15. MOEHLIG, R. C. and STEINBACH, A. L. *J. Amer. med. Ass.*, **154**, 42 (1954)
16. AUB, J. C., BAUER, W., HEATH, C. and ROPES, M. *J. clin. Invest.*, **7**, 97 (1929)
17. ROBERTSON, J. D. *Nature, Lond.*, **148**, 724 (1941)
18. HARRISON, H. E. and HARRISON, H. C. *Amer. J. Physiol.*, **137**, 171 (1942); *J. clin. Invest.*, **20**, 47 (1941)
19. CRAWFORD, J. D., GRIBETZ, D. and TALBOT, N. B. *Amer. J. Physiol.*, **180**, 156 (1955)
20. CRUICKSHANK, E. M., KODICEK, E. and ARMITAGE, P. *Biochem. J.*, **58**, 172 (1954)
21. GEISSBERGER, W. *Helv. med. Acta*, **18**, 461 (1951)
22. BELLIN, J. and LASZLO, D. *Science*, **117**, 331 (1953)
23. STAUB, H. *Med. Wschr. Schweiz.*, **84**, 499 (1954)
24. STEGGERDA, F. R. and MITCHELL, H. H. *J. Nutr.*, **31**, 423 (1946)
25. MCCANCE, R. A. and WIDDOWSON, E. M. *Biochem. J.*, **33**, 523 (1939)

23. JOHNSTON, R. M. *J. clin. Invest.*, **16**, 223 (1937)
24. WELCH, C. S., WAKEFIELD, E. G. and ADAMS, M. *Arch. int. Med.*, **58**, 1095 (1936)
25. GREENBERG, D. M. *J. biol. Chem.*, **157**, 99 (1945)  
L'HEUREUX, M. V., TWEEDY, W. R. and ZORN, E. M.  
*Proc. Soc. exp. Biol. N.Y.*, **71**, 729 (1949)  
WALLACE, H. D., SHIRLEY, R. L. and DAVIS, G. K.  
*J. Nutr.*, **43**, 469 (1951)
26. SJOLLEMA, B. *J. biol. Chem.*, **57**, 255 (1923)
27. COWELL, S. J. *Biochem. J.*, **31**, 848 (1937)
28. STEWART, C. P. and PERCIVAL, G. H. *Biochem. J.*, **21**, 301 (1927)
29. GREENWALD, I. and GROSS, G. *J. biol. Chem.*, **66**, 20 (1925)

## CHAPTER XV

# Extra-skeletal Functions of Calcium

IN the previous chapters the description of the metabolism of Ca has centred round its functions and participation in the formation and reactions of the hard tissues. Ninety-nine per cent. of the Ca of the body is in the skeleton and teeth, but the remaining 1 per cent. plays an important role in cellular and other bodily activities, which will be briefly reviewed.

Ca has a number of general actions which have been largely investigated in protozoa and simple metazoa. It has a protective action against the toxic actions of other ions, especially Na and K. The anaesthetic action of Mg can be reversed by small amounts of Ca. Even in small concentrations Ca will protect against the toxic action of distilled water, and solutions containing only Ca salts are not harmful to many lower organisms.

Ca governs cell permeability, usually by reducing it. This has been found with plant cells and human erythrocytes, in which the leaching of salts is retarded by Ca; the rate of intestinal absorption of sugar is lowered in parathyroidectomized rats. Ca also maintains the semipermeability for water of membranes such as those of sea-urchins and plant cells.

The absence of Ca from the environment is most injurious, largely owing to the unopposed effects of other ions. Muscle will not contract in the absence of Ca, this applying to smooth, heart and striated muscle. On the other hand, this tissue is sensitive to the concentration of Ca. Ringer showed many years ago that a solution with an excess of Ca perfused through the heart caused contraction to cease in systole, and as mentioned earlier, McLean and Hastings have used the frog heart



as a method of estimating the level of free Ca ions. Skeletal muscle is impermeable to Ca, but if it is cut so that Ca ions can enter, violent tetanic contractions occur.

Ca is apparently essential for clotting reactions, since it takes part in the coagulation of milk and blood. It has a marked effect on the integrity of intercellular cementing substance, which becomes weakened if Ca is absent from the surrounding medium. The blastomeres of sea-urchin eggs become separated in Ca-free sea-water, which is probably due to the absence of this clotting action. Ca will cause the plasmodium of amoebae to stiffen. In blood coagulation Ca is needed for the conversion of prothrombin to thrombin, but the plasma Ca has to fall considerably, to levels around 1 mg. per 100 ml. plasma, before any retardation of clotting occurs.

Nerve is very sensitive to the Ca content of its environment. If the Ca level is lowered, the nerve becomes hyperexcitable and may generate a spontaneous train of impulses. Not only the nerve, but also the sensitivity of ganglia to acetyl choline and in fact the excitability of the whole nervous system may be enhanced by Ca lack. This is seen clinically in tetany associated with hypocalcaemia. In the case of nerve, it has been suggested that 'accommodation', the process whereby nerve resists the action of the exciting stimulus, is depressed.

Ca accelerates the action of several enzymes, but there seems to be an optimal concentration for this to occur, excessive amounts inhibiting their activity. Blood clotting, which is basically enzymic, is prevented by excess Ca.

Heilbronn considers Ca to play a leading role in the development of bioelectric potentials, due to the existence of a Ca electrode in the Ca proteinate of the cell cortex. The present writer feels that the ability of Ca to combine with proteins to form compounds which occur almost everywhere in the body is probably the most significant biological property of this element, and accounts for its participation in so many physiological activities.

## Summary

IN the previous chapters the writer has given a survey of the literature and has attempted to weigh up the various findings as fairly as possible. This brief summary is based on his own conclusions from the published observations in the field.

Milk and milk products are the best dietary sources of Ca and pasteurization has little if any effect upon this. Certain vegetables form valuable supplementary sources. It is more important to know the availability of Ca to the body in these foods than the total amounts as there is not necessarily any correlation between the two. Water, even when very hard, is not a reliable source of Ca.

Work with Ca<sup>45</sup> shows that about half of the dietary Ca is absorbed into the body. A number of factors (e.g. reaction of the diet, fat, protein) have been stated to affect Ca absorption, but from the balance of the evidence it appears that in man the only important single factor is vitamin D. This vitamin also controls the Ca metabolism of bone and possibly renal P excretion. Phytic acid, present in high-extraction flour, has been claimed in England to be significant, in that it depresses Ca absorption in the intestine, but while this has been confirmed experimentally, it is doubtful if it plays such a role in practical dietetics.

In studying the question of Ca requirements, a number of factors have to be taken into account. The so-called endogenous loss, or maintenance requirement, due to processes other than growth, is a very real liability in adult man, and the Ca intake has to meet this to keep the individual in Ca equilibrium. In spite of statements to the contrary, the present writer holds strongly to the view that the previous Ca intake is, within wide limits, the most important controlling factor in

determining the level of Ca intake for equilibrium, owing to the ability of the body to adapt to low dietary levels of Ca. It therefore follows that the Ca requirement is a function of the previous dietary history. With our present dietary habits, certain figures have been arrived at, but they are relevant only to that Ca status. In many studies the significance of the previous Ca intake has not been realized, and as a result the findings for short period balance experiments have no practical meaning and must be discarded.

In growing animals there is no requirement for maintenance and the Ca in the diet is used solely for growth purposes. It should thus be easy to determine, from the daily Ca accretion, what the intake should be. Reliable data as to the composition of the body at different ages are so far lacking, but when such knowledge has been assembled, the problem should be largely solved.

The utilization of Ca in the body is high in many animals, but it is much lower in well-nourished children and adults, between 20 and 30 per cent. of that ingested. It may be much higher in individuals on a previously lower Ca intake. In old people an osteoporosis often develops, due to a combination of endocrine imbalance, inactivity and possibly Ca deficiency.

Pregnancy does not impose a strain on the Ca metabolism of the mother as the Ca content of the foetus at term is not high. Lactation, on the other hand, may lead to quite severe depletion unless extra Ca is given in the diet. Even so, the mother may find the Ca of her bones more available for her milk than that of her food.

The Ca level of the blood is remarkably constant; Ca is present in two fractions, ionized and non-diffusible, the latter being combined with protein. In man, virtually all the Ca is in the plasma. A large number of factors, both dietary and endocrine, control the blood Ca level, the most important endocrine influence coming from the parathyroids. Two views have been adumbrated, one that the gland acts on Ca mobilization from bone, and the other that it controls renal phosphate excretion. The balance of evidence supports the former view,

though recent work suggests that there are two factors, hypercalcaemic and phosphaturic, in parathyroid extracts.

The histology and biochemistry of bone are becoming increasingly interconnected, especially owing to the use of the rachitic epiphysis which will calcify under appropriate conditions *in vitro*. Using this technique, it has been shown that the glycolytic cycle may under certain conditions govern cartilage calcification, though it is possible for cartilage to calcify with all enzymes inactivated. Quite frankly, the present writer cannot see the solution to this discrepancy, but advances are so rapid in this field that the problem will doubtless soon be solved. Robison's alkaline phosphatase seems to be solely concerned with matrix formation, and not with calcification. There is strong and increasing evidence that mucopolysaccharides are intimately related to calcification, by acting as a template for crystallization.

The evidence is now unanimous that bone salt has an apatite structure, the differences in composition (e.g. Ca : P ratio) in bone being accounted for by surface adsorption. The total surface area of the bone crystals in an adult man has been calculated to be over 100 acres, which is in equilibrium with the blood-plasma, and accounts in part for the constancy of the blood Ca level. The process of mineralization of bone is a crystallization and not a precipitation from supersaturated solutions, and no solubility product has yet been demonstrated to be exceeded. However, calcification is in some ways correlated with the product  $[Ca^{++}] \times [HPO_4^{--}]$  which seems to determine the upper limit for Ca and P ions in solution. Bone formation is governed by almost all the endocrine glands and several of the vitamins. The most extreme change caused by dietary imbalance is rickets, due in man to lack of vitamin D, when the calcification sequences slow and finally cease.

Teeth differ from bones in having only a one-way process of calcification and no resorption can take place. The fully erupted tooth is metabolically very inactive compared with bone, but the unerupted tooth and the continually erupting rodent incisor have a high Ca metabolism. The mineral in both



enamel and dentin has an apatite structure. It seems likely that mucopolysaccharides are implicated in calcification as they are in bone. Several vitamins and endocrine glands control the formation of dental tissues.

The bulk of Ca lost from the body appears in the faeces, but most of this is unabsorbed dietary Ca. There seems no doubt that active Ca excretion from the body occurs via the gut in man, but it may be a reserve mechanism not usually employed to any extent. Active Ca excretion by this route also occurs in animals. Recent work suggests that the kidney tubule re-absorbs ionic Ca very efficiently, and the Ca lost in the urine is in the form of unionized complexes which the tubule cannot absorb. This is the most convincing explanation up to now of the constant Ca loss in the urine. Sweat forms a vehicle for Ca loss which though relatively small is not insignificant.

In conclusion it can be stated that there is hardly any physiological action which does not involve the intervention of Ca.

# Index

- (Unless otherwise stated, phosphatase refers to alkaline phosphatase)
- Accommodation in nerve and Ca, 154
- Acetyl choline, Ca on sensitivity of ganglia to, 154
- Achlorhydria and Ca deficiency, 1
- Acid-base balance of body and bone composition, 120
- Acid mucopolysaccharides  
bone and, 112, 130, 157  
Ca binding in bone by, 129  
dentin and, 130, 139-40, 158  
enamel and, 130, 139-40, 158  
metachromasia of, 129, 130  
metachromatic dyes and Ca binding by, 131
- Acidosis and  $\text{CaCl}_2$  ingestion, 17
- Acromegaly, bone changes in, 105
- Adaptation to low levels of Ca intake, 2, 54, 76, 156
- Adenosine triphosphate  
action of phosphatase on, 126  
in epiphyseal cartilage, 127  
inhibition of calcification by, 126
- Adrenal glands, 106  
intracerebral implantation with bone, 108  
removal on bone formation, 106  
removal on tooth formation, 142
- Adrenocorticotrophic hormone  
blood Ca and, 88  
bone formation and, 106  
Ca balance and, 148  
chondrogenesis and, 106
- Adsorption  
bone salt and, 113, 115  
P by teeth, 138
- Adults, recommended Ca standards for, 73, 74, 156
- Age  
influence on Ca content of  
aorta, 43  
body, 36, 40  
bone, 44  
trachea, 43
- Albumin, Ca binding by, 83
- Ameloblasts  
action of, 136  
glycoprotein in, 139  
phosphatase in, 124, 140  
and ribonucleic acid, 125  
radio-S in, 140
- Amelogenesis, 136  
phosphatase and, 124
- Amelogenesis imperfecta, 142
- Aorta, Ca content of, and age, 43
- Apatite (*see also* Hydroxyapatite)  
structure of bone crystals, 113, 115, 157  
dentin, 135  
enamel, 135
- Appetite and dietary Ca level, 16
- Ascorbic acid  
deficiency on bone formation, 104  
*gerüstmark* and, 104  
tooth formation and, 142
- Ash content of bone, 44, 45
- Balance metabolism data and Ca requirements, 75, 76
- Beryllium and rickets, 100
- Bioelectric potentials and Ca, 154
- Blood Ca (*see also* Plasma Ca)  
adrenal glands on, 88

**Blood Ca—*cont.***

- adrenalin on, 88
  - adrenocorticotrophic hormone on, 88
  - animals, 82
  - available skeletal Ca and, 121
  - blood coagulation and, 154
  - blood citric acid and, 131
  - bone citric acid and, 131
  - bone citrogenase and, 131
  - buffer-like action of bone and, 120
  - cortisone on, 88
  - diethylstilboestrol on, 84, 87
  - dihydotachysterol on, 86
  - egg-laying and, 84, 87
  - ethylenediamine tetra-acetic acid on, 85
  - hypophysectomy on, 87
  - insulin and, 87
  - ionized, 82-3, 156
  - lactation and, 64
  - lactose on, 57
  - man, 82
  - myxoedema and, 87
  - muscle Ca and, 44
  - non-diffusible, 82-4, 156
  - oestrogens and, 87, 107
  - parathyroids and, 86, 88-90, 156
  - P on, 84
  - P reabsorption in kidney tubules on, 149
  - pregnancy and, 62
  - removal on blood Ca level, 121
  - rickets and, 85, 117
  - scurvy and, 105
  - starvation and, 85
  - thyroid gland on, 87
  - vitamin D on, 86, 102, 131
- Blood coagulation and Ca, 154**
- phosphatase
    - changes during rickets, 125
    - scurvy, 125
  - radium on, 125

**Blood inorganic P**

- blood Ca relationship, 84
  - insulin action on, 87
  - oestrogen administration to mice on, 107
  - rickets and, 117
  - scurvy and, 105
  - starvation on, in rickets, 85
  - vitamin D and, 86, 102
- Body, Ca content of, 36-45**
- adults, 42
  - age on, 36, 40, 156
  - children, 39-41, 156
  - dietary factors on, in the rat, 37
  - human foetuses, 38, 77
  - lactation on, 62, 64
  - lactose on, 57
  - pregnancy on, 62, 63
- Bone (*see also* Skeleton)**
- acidotic states on composition of, 120
  - acromegaly and, 105
  - adrenocortrophic hormone on, 106
  - adsorption by bone salt, 113, 115
  - adsorption of Ca and P on crystals of, 118
  - ascorbic acid on, 104
  - ash content, 44-5
    - lactose on, 57
  - available Ca of, 121
  - buffer action of, 120
  - Ca and P deficiency on, 98
  - Ca in, 3
  - Ca : P ratio in young bone, 120
  - calcium phosphate hydrate in, 114
  - carbonate hydroxyapatite in, 114
  - chemistry, 112-21
  - chondroitin sulphate in, 112, 130
  - citrate in, 116, 131
    - vitamin D on, 131

Bone—*cont.*

- CO<sub>2</sub> in young bone, 120
- collagen in, 112
  - in scurvy, 105
- composition, 44, 45
  - of diet on, 120
- crystals, 112–16, 157
  - composition and structure, 113
  - unit cell of, 113
- desoxycorticosterone on, 107
- diaphysis of, 98
- dietary source of Ca, 1
- endochondral formation of, 97
  - primary spongiosa of, 97
- epiphyseal cartilage of, 97
- exchange of Ca and P on
  - crystals of, 115
- formation, 95–108
  - and enzymes, 123–8
- fractures in avitaminosis A, 103
- gigantism and, 105
- glycolytic cycle and calcification of, 127
- gonadal insufficiency on, 107
- heterionic exchange in, 115, 116, 120
- Howship's lacunae in, 95
- hydration layer of, 115, 116
  - and age, 119
- hydroxyapatite in, 114
- hyperparathyroidism on, 108
- hypophysectomy on, 103
- inanimation on, 103
- intracerebral implantation of
  - with calciferol, 102
  - with oestradiol, 107
  - with parathyroid, pituitary, thyroid and adrenal 108,
  - with vitamin A, 103
- intra-membranous formation of, 96
- isoionic exchange in, 115
- lactation on, 64
- 'local factor' in, 112, 128–31
- male hormone on, 107

Bone—*cont.*

- matrix of, 112
  - as 'seedling' agent in calcification, 118
- medullary bone formation in
  - egg laying, 106
- metabolic changes in, in rickets, 102
- metachromasia in, 130
- mucopolysaccharides in, 112, 130
- old bone, calcification of, 119
- oestrogens on, 106
- osteoblasts and, 95
- osteoclasts and, 95
- osteocytes in, 112
- osteoid in, 97
- osteomalacia in, 100
- pantothenic acid deficiency on, 103
- parathyroids on, 88–90, 156
  - after nephrectomy, 89
- phosphatase in, 123, 124
- physics, 112–21
- pituitary on, 105
- PO<sub>4</sub>: CO<sub>2</sub> ratio of, 120
- primary calcification in, 97
- protein deficiency on, 103
- pyridoxine deficiency on, 103
- recalcification *in vitro*, 130
- recrystallization of bone salt, 116
- remodelling, 98
- resorption, 95
  - and phosphatase, 124
- riboflavin deficiency on, 103
- rickets in, 98–102, 157
- salt, adsorption by, 115
  - composition of, 114
  - lability of, 114
- scorbutic changes and phosphatase, 125
- scurvy and, 104
- secondary spongiosa of, 98
- spongiosa as Ca store, 98
- staining reactions of, 97



**Bone—cont.**

- structure of, 112
- testosterone on, 107
  - and hypophysectomy, 105
- thyroid on, 106
- trabeculae of, 98
- vitamin A and, 102
- vitamin D action on, 98, 100-2, 157
- vitamin E and, 105
- young bone, calcification of, 119

Bone P, vitamin D and, 102

**Brain**

- Ca content of, 43
- rickets on, 44

Bread, Ca content of, 6

Bulk of diet and Ca absorption, 16

**Calcification, 3**

- adenosine triphosphate and, 126
- creatine phosphate and, 128
- cyanide on, 128
- dentin, 136, 138-9
- destruction of enzymes and, 128
- enamel, 136, 139
- epiphyseal cartilage *in vitro*, 126, 157
- fluoride on, 127, 128
- glucose-1-phosphate and, 126, 127
- glycerophosphate and, 128
- glycogen and, 126
- glycolytic cycle and, 127
- inactivation by strontium, 129
- inhibition of, and phosphatase, 126
  - by metachromatic dyes, 130
- iodoacetate and, 127
- 'local factor' in, 126
- metachromasia and, 129, 130
- mucopolysaccharides and, 129-131, 157
- osteoid, *in vitro*, 126
- P.A.S. reaction and, 130

**Calcification—cont.**

- phenyl phosphate and, 128
- phloridzin and, 127
- phosphatase and, 123-6
- phosphate, role of, in, 126-8
- phosphorylase and, 127
- 'seeding' as cause of, 118
- solubility products and, 116-17
- teeth and mucopolysaccharides, 139
- 3-phosphoglycerate and, 127
- young bone, 119

Calcination on apatite crystals, 114

Calciotraumatic response in teeth, 141

Calcium (*see also* Radio-Ca, Calcification, Bone)

- accommodation in nerve and, 154
- achlorhydria and, 1
- acidosis and, 17
- adaptation to low intakes of, 2, 54-6, 77, 156
- adsorption of, on bone crystals, 115, 119
- adults, recommended standards for, 73, 74
- aorta, content of, 43
- appetite and, 16
- balance metabolic data of, 75-6
- binding of, by albumin, 83
- bioelectric potentials and, 154
- blastomeres of sea-urchin eggs and, 154
- blood Ca (*see* Blood Ca), 82-91, 156
- body stores of, Ca on retention of, 56
  - adults, 42, 43
  - children, 39-42, 156
  - dietary factors on, 37, 38
  - foetuses, 38, 39
  - growth and, 39-42
- bone, citrate and, 131
  - Ca content of, 44-45

**Calcium, bone—*cont.***

- crystals and, 3, 113–15
  - as dietary source of Ca, 1
- brain, content of, 43
- bread, content of, 6
- cell permeability and, 153
- clearance, 147
  - citrate on, 147
  - phosphate on, 147
  - versene on, 147
- cheese, content of, 5
- children, recommended Ca standards for, 74, 79–80
- chondroitin sulphate and, 129
- clotting of blood and, 154
  - milk and, 154
- collagen and, 129
- complex with adenosine triphosphate, 126
- deficiency and achlorhydria, 1
- dentin, content of, 135
  - Ca deficiency and, 140, 141
  - uptake of Ca by, 138
- diethylstilboestrol, blood Ca and protein of chickens, 84, 87
- dihydrotachysterol on blood level of, 86
- distilled water and protective action of, 153
- electrode in cell, 154
- enamel, content of, 135–6
  - uptake by, 138
- endogenous loss of, 49, 155
  - adults, 51
  - faecal, 12, 52
  - infants, and children, 50–1, 156
- exchange in bone, 118
- excretion, 147–50
  - endocrine factors on, 148
  - faecal, 149–50, 158
  - kidney, 147–8, 158
  - thyroid gland and, 87, 106
- fat tissue, content of, 43
- fertility and, 64

**Calcium—*cont.***

- heart, content of, 44
- heterionic exchange of, in apatite crystals, 115–16
- infants, recommended standards for, 74, 78
- intake on renal excretion, 148
- intercellular cementing substance, and, 154
- intestinal absorption of, 1, 12–33, 155
  - dietary bulk and, 16
  - dietary fibre and, 16
  - dietary protein and, 16, 155
  - pH and, 1, 14, 155
  - phosphatase and, 33
  - phosphate on, 17
  - phytic acid on, 21–8, 155
  - soaps and, 15
  - vitamin D and, 30–3, 155
- kidney, content of, 44
  - percentage excreted by, 148
- lactation and, 64–6, 156
  - recommended standards during, 73, 77–8
- lactose on retention of, 56–7
- liver, content of, 43, 44
- magnesium and, 153
- maintenance requirement of, 49, 155
- metabolism
  - children, 60–2
  - lactation, 64–6, 156
  - oestrogen on, 59
  - old people, 58–9, 156
  - pregnancy, 62–4, 156
- milks, content of, 5, 155
  - availability of Ca of, 7
  - utilization of Ca of, 57
- mucopolysaccharides in cartilage and, 129
- muscle
  - Ca content of, 43
  - contraction and, 153
- nerve, conduction and, 154
  - ganglia and, 154

**Calcium—cont.**

- parathyroids and, 2
  - plasmagel of amoebae and, 154
  - potassium and, 153
  - pregnancy and, 62-4, 156
    - recommended standards during, 73, 77
  - protozoa, action on, 153
  - reabsorption by renal tubule, 147, 148, 158
  - requirements, 2, 53, 71-80, 155
  - retention of, 3, 49, 60-2
    - children, 60-2
    - infants, 60, 78
  - salts, availability of, 7
    - utilization of, 58
  - simple metazoa and, 153
  - skin content of, 43
  - sodium and, 153
  - spleen, content of, 43
  - state of, in body, 2
  - store in spongiosa, 98
  - sulphate in cartilage and, 129
  - sweat, loss in, 147
  - threshold for renal excretion, 147
  - tissues, content of, 43-5
  - trachea, content of, 43
  - utilization of, 3, 49, 57-8, 62, 156
    - children, 62
    - infants, 62
  - vegetables, 1
    - availability of Ca of, 7, 8, 155
    - content of, 5-6, 155
    - water, as source of, 9, 155
- CaHPO<sub>4</sub>**, solubility product of, and calcification, 117
- Ca : P ratio**
- adult body, 42
  - age and ratio of bone, 120
  - blood and PO<sub>4</sub> : CO<sub>2</sub> ratio in bone, 120
  - bone, 45, 114, 120
    - in acidotic states, 120
  - diet and Ca content of muscle, 44

**Ca : P ratio—cont.**

- diet and rickets, 32, 98
  - diet and tooth formation, 14
  - foetal body, 39
  - hydroxyapatite, 114, 115, 119
  - newly calcified bone, 120
  - tertiary phosphate, 114
  - young bone, 120
- Calcium phosphate (tertiary)**
- solubility product of, and calcification, 117
- Canadian Council on Nutrition**
- recommended dietary standards for Ca, 71
    - adults, 73, 76
    - children, 74
    - infants, 74
    - lactation, 73, 78
    - pregnancy, 73, 77
- Carbonate hydroxyapatite bone**, 114
- Carbonate in bone**, 114
- acidotic states and, 120
  - age and, 120
  - heterionic exchange by, 116
- Carbonate in enamel**, 136
- Cartilage (see also Epiphyseal cartilage)**
- sulphate content of, 130
- Cats, faecal excretion of Ca b**
- 150
- Cattle**
- faecal endogenous loss of, 52
  - plasma Ca of, 82
- Cementum of teeth**, 135
- Cheese, Ca content of**, 5
- Chickens**
- growth hormone on bones
  - embryos, 105
  - Vitamin D on bones of, 101
  - on kidney of, 101
- Children**
- Ca content of bodies of, 39-4
  - Ca metabolism of, 60-2
  - Ca retention by, 60-2
  - Ca utilization by, 62

ren—*cont.*

recommended Ca standards for, 74, 79-80, 156  
 rickets in, 99  
 vitamin D on Ca retention of, 61  
 1,25-dihydroxyvitamin D<sub>3</sub> (calciferol), 31  
 osteogenesis, adrenocorticotrophic hormone on, 106  
 chondroitin sulphate  
   ne and, 112  
   binding by, 129  
   collagen complex, 129  
   competition for, in calcification, 129  
   vitamin and, 139  
   metachromasia of, 130  
   metachromatic dyes and calcification, 130  
   apatite  
     apatite crystal and, 116  
     food citrate and parathyroidectomy, 131  
     and vitamin D, 131  
     ne and, 116, 131  
     ne and parathyroids, 131  
     and rickets, 131  
     clearance and, 147  
     fecal Ca excretion and, 149  
     eth, 135  
     phosphatase in bone, parathyroidectomy on, 131  
     retention of Ca, 147-8  
     rate on, 147  
     phosphate on, 147  
     arsene on, 147  
     metabolic reactions and Ca, 154  
     vitamin  
       binding and bone crystals, 112  
       ne and, 112  
       carbohydrate in dentin and, 139  
       changes in bone and teeth during scurvy, 105  
       chondroitin sulphate complex, 129  
       vitamin, 135  
       formation of, in, 136

Committee on Nutrition of the British Medical Association  
 recommended dietary standards  
   for Ca, 71  
   adults, 73, 76  
   children, 74  
   infants, 74  
   lactation, 73, 78  
   pregnancy, 73, 77  
 Compensation dialysis of plasma Ca, 82  
 Cooking on Ca in vegetables, 6  
 Cortisone  
   blood Ca and, 88  
   after nephrectomy, 88  
   bone formation and, 106  
   excretion of Ca and, 88, 148  
   urinary Ca and, 148  
 Cows  
   Ca metabolism of, in lactation, 64  
   Ca storage by, during pregnancy, 63  
 Creatine phosphate and calcification, 128  
 Cushing's syndrome, bone formation in, 106  
 Cyanide and calcification, 128  
 Dentin  
   acid mucopolysaccharides in, 130, 139-40  
   adrenalectomy on, 142  
   adsorption of P by, 138  
   apatite in, 135, 158  
   Ca deficiency on, 140  
   uptake by, 138  
   calcification of, 136  
   calciotraumatic response of, 141  
   carbohydrate and calcification of, 139  
   chondroitin sulphate in, 139  
   citrate in, 135  
   collagen in, 135  
   composition of, 135



**Dentin—cont.**

- formation in vitamin A deficiency, 142
  - gonadectomy on, 142
  - hexosamine in, 139
  - hypophysectomy on, 142
  - interglobular, 141
  - isoionic exchange in apatite crystals of, 115, 138
  - lysine deficiency on, 142
  - metachromasia of, 130, 139
  - P deficiency on, 140
    - uptake by, 137, 138
  - phosphatase in, 124, 140
  - radioautographs of  $P^{32}$  uptake by, 139
  - rickets and, 141
  - scurvy and, 142
  - staining reactions of, 137
  - thyroidectomy and, 142
  - tryptophane deficiency on, 142
  - vitamin D and, 143
- Desoxycorticosterone** on bone formation, 107
- Diamox** on Ca reabsorption by renal tubules, 148
- Diethylstilboestrol** on blood Ca, 84, 87
- and protein in chickens, 84
- Dihydrotachysterol (A.T.10)**
- blood Ca and, 86, 149
  - bone and, 86
  - intestinal absorption of Ca and, 86
  - P reabsorption in kidney and, 149
  - renal function and, 86
- Dinitrophenol** on Ca absorption by renal tubules, 148
- Distilled water**, toxic action of, and Ca, 153
- Dogs**, faecal excretion of Ca by, 150
- Dutch Nutrition Council**
- recommended dietary standards for Ca, 72

- Dutch Nutrition Council—cont.**
- recommended dietary standards for Ca, adults, 73, 76
  - children, 74
  - infants, 74
  - lactation, 73, 78
  - pregnancy, 73, 77

**Egg laying**

- blood Ca level and, 84, 87
- medullary bone formation and, 106

**Enamel**

- acid mucopolysaccharides in, 130, 139
- apatite structure of, 135, 158
- carbonate in, 136
- Ca uptake by, 138
- citrate in, 135
- composition, 135
- eu-keratin in, 135
- hypoplasia of, 142
- isoionic exchange in, 115
- metachromasia of, 130
- phosphatase in, 140
- P uptake by, 138
- radioautographs of, after  $P^{32}$  administration, 139
- radio-S in, 140
- vitamin A deficiency on, 142

**Enamel organ**

- functions of, 136
  - glycoprotein in, 139
- Endochondral ossification**, 95, 97
- bone trabeculae in, 98
  - epiphyseal cartilage in, 97
  - hypophysectomy and, 105
  - metaphysis formation in, 99
  - osteoblasts and, 95
  - primary spongiosa in, 98
  - rickets and, 98–9
  - scurvy and, 104
  - secondary spongiosa in, 98
  - thyroidectomy and, 106
- Endogenous loss of Ca**, 49
- adults, 50, 51–3, 155

- Endogenous loss of Ca—*cont.*  
  faecal, 52  
  infants and children, 50-1, 156
- Enzymes  
  bone formation and, 123-31, 126  
  Ca action on, 154  
  destruction of, and calcification, 128, 157  
  tooth formation and, 140
- Epiphyseal cartilage  
  adenosine triphosphate in, 127  
  calcification of, *in vitro*, 32, 126, 157  
  closure of, 98  
  glycogen in, 126  
  hexosamine in, 129  
  hypophysectomy and, 103, 105  
  inanition and, 103  
  metachromasia of, 130  
  metachromatic dyes and calcification of, 130  
  mucopolysaccharides in, and Ca binding, 129, 130  
  P.A.S. reaction in, and calcification, 130  
  pantothenic acid deficiency and, 103  
  phosphatase in, 124  
  phosphorylase in, 127  
  protein deficiency and, 103  
  P uptake by, 129  
  pyridoxine deficiency and, 103  
  response to vitamin D, 100, 143  
  riboflavin deficiency and, 103  
  rickets and, 99  
  scurvy and, 104  
  structure of, 97  
  thyroxine on, after hypophysectomy, 105
- Ergocalciferol, 30  
  poisoning and liver and brain Ca, 44
- Erythrocytes, permeability of and Ca, 153
- Ethylenediamine tetra-acetic acid (versene)  
  blood Ca level and, 85  
  bone calcification and, 130  
  Ca clearance and, 147
- Eu-keratin in enamel, 135
- Excretion of Ca, 2, 147-50, 158  
  adrenal glands and, 88  
  Ca intake on renal excretion of, 148  
  endocrine factors on, 148  
  faecal, 2, 52, 58, 65, 88, 149-50, 158  
  kidney and, 147-9  
  percentage excreted by kidney, 148  
  reabsorption of Ca in renal tubules, 147, 148  
  renal, 2, 59, 86, 88, 147-9, 158  
  skin and, 147  
  thyroid gland and, 87, 106
- Faecal excretion of Ca, 2, 149-50, 158  
  animals, 150  
  citrate on, 149  
  cortisone on, 88, 148  
  endogenous, 52, 149, 150  
  lactation and, 65  
  man, 149  
  variations in, 58
- Fats and soaps and intestinal Ca absorption, 15
- Fatty tissue, Ca content of, 43
- Fertility, Ca on, 64
- Fibre in diet and Ca absorption, 16
- Fluoride  
  calcification and, 127, 128  
  calciotraumatic response in teeth and, 141  
  heterionic exchange of, in apatite crystal, 116  
  tetany and, 85
- Foetus, Ca content of body of, 38-9, 77, 156

- Food and Nutrition Board,  
National Research Council  
of the U.S.A.  
recommended dietary standards  
of Ca, 71, 72  
adults, 73, 76  
children, 74  
infants, 74  
lactation, 73, 78  
pregnancy, 73, 77
- Ganglia, 154
- Gerüstmark*, 104  
ascorbic acid on, 104  
bone and, 104, 125  
teeth and, 142
- Gigantism, bone changes in, 105
- Glucose-1-phosphate and calci-  
fication, 126, 127
- Glycerophosphate and calcifica-  
tion, 128
- Glycogen  
calcification and, 126-7  
epiphyseal cartilage and, 126  
teeth and, 140
- Glycolytic cycle and calcification,  
127-8
- Glycoprotein in enamel organ,  
139
- Gonadectomy on dentin forma-  
tion, 142
- Grenz-ray analysis of teeth, 137  
calciotraumatic response and,  
141  
thyroidectomy and, 142
- Growth  
Ca content of diet on, in rats, 37  
endogenous loss of Ca during,  
51  
scurbutic changes in bone and,  
104
- Growth hormone  
adrenalectomy on bone and,  
106  
adrenocortrophic hormone on  
bone formation and, 106
- Growth hormone—*cont.*  
bone formation and, 105  
in hypophysectomized ani-  
mals, 105  
bones of chick embryos and,  
105  
epiphyseal cartilage and, 105
- Guinea pigs, cortisone on bone  
formation of, 106
- Halisteresis of bone, 96
- Haversian canals  
calcification round, in young  
bone, 119  
hyperparathyroidism and, 108  
phosphatase in, 124
- Heart  
Ca in, 44  
determination of plasma ionic  
Ca with, 83, 153  
sensitivity to Ca, 153
- Hens  
diethylstilboestrol on blood Ca  
and protein of, 84  
plasma Ca level in during egg-  
laying period, 84
- Heterionic exchange  
apatite crystals and, 115, 116  
carbonate ions and, 116, 120  
fluoride ions and, 116  
hydronium ions and, 116  
phosphate ions by carbonate  
ions, 120  
potassium ions and, 116  
sodium ions, 116  
uranyl ions, 116
- Hexosamine  
rachitic cartilage and, 129  
teeth and, 139
- Hydration layer  
bone salt and, 115, 116, 119  
calcification in young bone and,  
119  
older bone and, 119  
synthetic hydroxapatite and,  
116

- Hydronium ions in apatite, 114, 116
- Hydroxyapatite (*see also* Apatite)  
bone and, 114  
formation from tricalcium phosphate hydrate, 114  
heterionic exchange in, 115-16  
isoionic exchange in, 115  
phosphate inclusion in, 120  
position of ions in, 115  
possible Ca : P ratios of crystals of, 115  
recrystallization of, 116
- Hydroxyl ions in bone crystals, 113
- Hypophysectomy  
adrenocorticotrophic hormone on bone formation and, 106  
blood Ca level and, 87  
bone formation and, 103, 105  
oestrogen action and, 107  
renal excretion of Ca and, 148  
testosterone on bone formation and, 105  
thyroxine on bone formation and, 105  
tooth formation and, 142
- Hypoplasia of enamel, 142
- Inanition on bone formation, 103
- Infants  
recommended Ca standards for, 74, 78-9  
retention of Ca by, 60, 78, 79
- Insulin and blood Ca and P, 87
- Intercellular cementing substance and Ca, 154
- Interglobular dentin, 141
- Intestine  
Ca absorption in, 1, 12-33, 155  
dihydrotachysterol on, 86  
parathyroid on, 86  
phytic acid on, 21-8, 155  
vitamin D on, 30-3, 155  
formation of vitamin D precursor in, 32  
secretion of Ca in, 1, 12
- Intramembranous ossification, 95, 96  
osteoblasts and, 95, 96  
osteoclasts and, 96  
rickets in, 99
- Iodoacetate, effect on calcification, 127
- Ions  
concentration of Ca and P on calcification, 117  
exchange of, in apatite, 115-16  
position of, in hydroxyapatite, 115
- Iron and rickets, 100
- Irradiation on vitamin D content of skin fat, 31
- Isoionic exchange  
bone and, 115  
Ca and P in bone, 118  
dentin and, 115  
enamel and, 115  
young bone and, 119
- Isotope dilution method, 12, 52
- Kidney (*see also* Renal), Ca content of, 44
- Korff fibres of dental pulp, 136
- Lactation, 62, 64-6  
Ca content of body and, 62  
Ca metabolism of women during, 64-6, 77, 78, 156  
recommended Ca standards during, 73, 77, 78
- Lactose  
Ca retention and, 56-7  
intestinal pH and, 56, 57  
tetany and, 56
- League of Nations Health Organization, recommended dietary standards for Ca, 71
- 'Line test'  
estimation of vitamin D by, 100  
healing of rickets and, 100, 101
- Liver  
Ca content of, 43



- Liver, Ca content of—*cont.*  
 after Ca administration, 44  
 in rickets, 44  
 'Local factor' in bone calcification, 112, 126, 128-31  
 Low levels of Ca intake, adaptation to, 54-6  
 Lysine deficiency on teeth, 142
- Magnesium, anaesthetic action and Ca, 153  
 Maintenance requirement of Ca, 49-53, 155, 156  
 Male hormone on bone formation, 107  
 Mandible, calcification of, and phosphatase, 124  
 Matrix of bone, phosphatase and formation of, 123, 124, 125  
 Medullary bone formation and oestrogens, 106  
 Metabolic balance experiments, 2  
 Metachromasia  
 calcification and, 129  
 chondroitin sulphate and, 130  
 dentin and calcification, 139  
 metachromatic dyes and calcification, 130  
 mucopolysaccharides and, 129  
 Metaphysis  
 enzyme studies and, 99  
 formation in rickets, 99  
*in vitro* calcification of, 32, 101, 126  
 'line test' in, 100  
 scurvy and, 104  
 Metazoa, simple, Ca on, 153  
 Mice  
 Ca content of, during growth, 37  
 cortisone on bone formation in, 106  
 oestrogen on blood Ca, P and phosphatase of, 107  
 bone formation in, 107  
 medullary bone in and pregnancy, 107
- Milk, 155  
 Ca content of, 5, 78  
 utilization of Ca of, by adult, 57, 58, 75  
 Mineral oil and intestinal vitamin D action, 32  
 Mucopolysaccharides, *see* Acid mucopolysaccharides  
 Muscle  
 Ca content of, 43  
 Ca : P ratio of diet and, 44  
 parathyroid and, 44  
 contraction and Ca, 153  
 Myxoedema, blood Ca level in, 8
- Neonatal line in teeth, 141  
 Nephrectomy  
 calciotraumatic response in teeth and, 141  
 cortisone on blood Ca after, 88  
 parathyroid extract on bone after, 89  
 blood Ca after, 89  
 Nerve, Ca on, 154
- Odontoblasts, 136  
 glycoprotein in, 139  
 phosphatase in, 125, 140  
 and ribonucleic acid in, 125  
 scurvy and, 142  
 vitamin A deficiency and, 142  
 Oestradiol  
 blood Ca level and, 87  
 after hypophysectomy, 87  
 after parathyroidectomy, 87  
 bone changes after intracerebral implantation of, 107  
 Oestrogens  
 blood Ca level in birds and, 87  
 91, 107  
 Ca, P and phosphatase in mice and, 107  
 bone formation in rats and, 107  
 man and, 107  
 hypophysectomy on blood Ca level and, 107

- Oestrogens—*cont.*  
  medullary bone formation and, 87, 106-7  
  therapy in osteoporosis, 59  
  tooth formation and, 143
- Old people, Ca metabolism of, 58-9
- Osteoblasts, 95, 112  
  blood Ca level and, 121  
  hyperparathyroidism and, 96  
  intramembranous bone formation and, 96  
  phosphatase in, 124  
  rickets and, 99  
  scurvy and, 104-5, 125  
  vitamin A and, 103
- Osteoclasts, 95-6  
  blood Ca level and, 121  
  hyperparathyroidism and, 96, 108  
  hypervitaminosis D and, 102  
  intramembranous bone formation and, 96  
  osteoclastomas and hyperparathyroidism, 108  
  parathyroid glands on, 90, 108  
  rickets and, 100  
  scurvy and, 104  
  vitamin A and, 103
- Osteocytes, 112  
  phosphatase in, 124
- Osteoid tissue, 97  
  calcification of, *in vitro*, 126  
  in hypervitaminosis D, 102  
  metachromasia of, 130  
  metaphysis and, 99  
  rickets and, 99
- Osteomalacia, 59, 100
- Osteoporosis, 100  
  Cushing's syndrome and, 106  
  immobility and, 59  
  oestrogen therapy and, 59  
  oestrogens in man and, 107  
  post-menopausal, 59  
  senile, 58, 98, 156  
  thyroid gland and, 106  
  X-ray diagnosis of, 59
- Oxygen in bone crystals, 113
- Pantothenic acid and bone formation, 103
- Parathyroid gland and hormone  
  blood Ca and, 86, 88-91, 156  
  and nephrectomy, 89  
  and volume of gland, 88  
  blood P and, 88, 89  
  bone and, 89, 90, 107-8, 156  
  cell changes in, 96, 108  
  citrate and Ca, 131  
  Ca binding by albumin and, 83, 84  
  calciotraumatic response in teeth and, 141  
  Ca metabolism and, 2  
  citrate and, 131  
  intestinal absorption of Ca and, 86  
  intracerebral implantation with bone, 108  
  muscle Ca and, 44  
  oestrogen on bone and, 87  
  plasma Ca and, 83, 149  
  reabsorption of P in kidney and, 149, 156  
  renal function and, 88, 149, 156  
  two hormones in, 90, 157  
  vitamin D and, 87
- Parathyroidectomy  
  action of vitamin A and, 91  
  blood citric acid and, 131  
  bone changes after, 107  
  bone citrogenase and, 131  
  intestinal absorption of sugar and, 153  
  oestrogen action on blood Ca level and, 87, 91, 107  
  renal excretion of P and, 88  
  rickets and, 91  
  tooth formation and, 141  
  vitamin D action on kidney and, 149
- Parathyrotrophic hormone, 87
- Pasteurization of milk, 8

- Periodic acid-Schiff reaction and calcification, 130
- Periosteum, phosphatase in, 124
- Phenyl phosphate and calcification, 128
- Phloridzin  
calcification of rachitic cartilage and, 127  
reabsorption of Ca by renal tubules and, 149
- Phosphatase, acid, 123
- Phosphatase, alkaline, 3, 123-6, 127, 128  
adenosine triphosphate and, 126, 127  
ameloblasts and, 124, 140  
bone matrix formation and, 123, 124, 125, 157  
calcification and, 117, 123-6, 157  
dentin and, 124, 140  
enamel and, 140  
extracellular, in bone, 124  
Haversian canals and, 124  
intestinal absorption of Ca and, 33  
odontoblasts and, 124, 140  
oestrogen action on blood and, 107  
optimum pH of, 123  
osteoblasts and, 124  
osteocytes and, 124  
periosteum and, 124  
radium on, 125  
rickets and, 124, 125  
scurvy and, 125  
bone content of during, 125  
serum content of during, 125  
wound healing during, 125  
substrates for, 127, 128  
tooth formation and, 124, 140  
vitamin D and, 33, 125
- Phosphate  
adsorption on bone crystals, 115, 119, 138  
dentin and enamel, 138,  
Phosphate—*cont.*  
Ca clearance and, 147  
exchange of, on bone crystals, 118, 120  
in teeth, 137, 138  
high-energy bonds and calcification, 128  
plasma and rickets, 117  
recrystallization of, in bone *in vivo*, 118  
role of, in calcification, 126-8  
tubular reabsorption of, 149  
and vitamin D, A.T.10 and parathyroid hormone, 149  
uptake by cartilage, 129  
teeth, 137-9
- Phosphate : carbonate ratio in bone, 120  
Ca : P ratio of diet on, 120  
teeth, 140
- Phosphorus  
deficiency on dentin, 140  
dentin content of, 135  
dietary, on Ca content of body, 38  
enamel content of, 135, 136  
parathyroid on renal excretion of, 88-90  
vitamin D and mobilization of, 101
- Phosphorylase  
calcification and, 127  
epiphyseal cartilage and, 127
- Phytase in flour, 25  
yeast, 25
- Phytic acid  
Ca metabolism and, 21-8, 155  
flour, content of, 22  
hydrolysis of, in intestine, 24  
by phytase, 25
- Pig  
Ca content of during growth, 37  
Ca on fertility of, 64
- Pigeons  
medullary bone formation in, 106  
oestrogens on, 107

- Pituitary (*see also* Hypophysectomy)  
   bone formation and, 103, 105  
   intracerebral implantation with bone, 108  
 Placenta, Ca content of, 39  
 Plant cells, permeability of, and Ca, 153  
 Plasma Ca (*see also* Blood Ca)  
   compensation dialysis of, 82  
   content in man, 82  
   animals, 82  
   frog-heart method of determination, 83  
   ionized, 82-3, 156  
   non-diffusible, 82, 83-5, 156  
   protein and, 83-5  
   ultra-filtration of, 83  
   vividiffusion of, 83  
 Plasmagel of amoebae and Ca, 154  
 Potassium  
   exchange in apatite crystal, 116  
   toxic action and Ca, 153  
 Predentin  
   Ca deficiency on, 140, 141  
   calciotraumatic response in, 141  
   formation of, 137  
   parathyroidectomy on, 140, 141  
   P deficiency on, 140, 141  
   rickets and, 141  
   vitamin D deficiency on, 140, 141  
 Pregnancy  
   Ca content of body and, 62  
   Ca metabolism during, 62-4, 156  
   Ca storage during, 63  
   recommended Ca standards during, 73, 77  
 Protein  
   chicken blood protein during egg-laying, 84, 85  
   diethylstilboestrol on, 84, 85  
   deficiency on bone formation, 103  
   Protein—*cont.*  
     dietary and Ca absorption, 16  
     plasma and Ca, 83-5  
   Prothrombin, 154  
   Protozoa, Ca on, 153  
   Puppies, rickets in, 99  
   Pyridoxine deficiency on bone formation, 103  
 Rabbit  
   Ca content of, during growth 37  
   cortisone on bone formation in 106  
   faecal excretion of Ca by, 150  
   plasma Ca of, 82  
 Rachitic cartilage (*see also* Epiphyseal cartilage)  
   calcification of, 32, 100-1, 117, 157  
   phosphatase and, 124  
 Radioactive isotopes, 2  
 Radio-Ca (*see also* Ca)  
   endogenous Ca and, 52  
   faecal excretion of, 149, 150  
   foetal bone and, 64  
   movement across placenta, 64  
   recrystallization of bone and apatite and, 116, 118  
   specific activity of, 52  
   teeth and, 138  
   transcapillary movement of, 84  
   utilization of, by adults, 58  
   vitamin D on bone and, 101, 102  
   intestinal absorption of, 33  
 Radio-Na  
   availability of that of skeleton, 119  
   transcapillary movement of, 84  
 Radium on bone and blood phosphatase, 125  
 Rat  
   Ca content of  
     dietary factors on, 37  
     fertility and, 64



- Rat, Ca content of—*cont.*  
   growth and, 36  
   lactation and, 64  
   cortisone on bone formation in, 106  
   faecal excretion of Ca by, 150  
   oestrogens on bone formation in, 107  
   retention of Ca by, 62  
   rickets in, 98  
   storage of Ca during pregnancy by, 63
- Reaction of intestinal contents and Ca absorption, 14, 155  
 of urine and Ca excretion, 148
- Recrystallization  
   apatite crystals and, 116  
   bone crystals and, 116  
   Ca in bone *in vivo*, 118  
   old bone and, 119  
   P in bone *in vivo*, 118  
   teeth and, 138, 139  
   young bone and, 119
- Renal excretion of Ca, 2, 147–9, 158  
   adrenocorticotrophic hormone on, 148  
   blood Ca level and, 121  
   Ca clearance, 147  
     citrate, phosphate and ver-sene on, 147  
   Ca in glomerular filtrate, 147  
   Ca intake on, 148  
   Ca reabsorption by tubules, 147, 148  
   Ca threshold for, 147  
   cortisone on, 88, 148  
   desoxycorticosterone on, 148  
   hypophysectomy on, 148  
   immobility and, 59  
   percentage of total excretion, 148  
   pH of urine and, 148  
   thyroid on, 148
- Renal excretion of P  
   parathyroids on, 88, 89, 156
- Renal excretion of P—*cont.*  
   vitamin D action on, 86, 149, 155
- Renal function, dihydrotachysterol on, 86
- Requirements of Ca, 2, 53, 71–80, 155
- Resorption of bone, 95, 96  
   halisteresis and, 96  
   osteoclasts and, 95, 96
- Retention of Ca, 1, 3, 49  
   body Ca stores on, 56  
   children and, 60–2  
   infants and, 60, 78  
   lactation and,  
     in cows, 64  
     in women, 64–6  
   previous dietary level on, 53–4
- Reversibility of calcification, 129
- Riboflavin on bone formation, 103
- Ribonucleic acid and phosphatase in ameloblasts and odontoblasts, 125
- Rickets, 32, 98–100  
   beryllium and, 100  
   blood Ca changes in, 85  
     starvation on, 85  
     vitamin D on, 86  
   blood phosphatase and, 125  
   bone ash during, 100  
     citrate and, 131  
     metabolic changes during, 102  
     phosphatase and, 124  
   Ca content of brain, liver and muscle during, 44  
   Ca  $\times$  P product of plasma and, 117  
   epiphyseal cartilage changes in, 99  
   healing of, 100–1  
   in Eire, 26  
   intramembranous bone and, 99  
   iron and, 100  
   ‘line test’ and, 100–1

**Rickets—*cont.***

- oestrogens and, 107
- parathyroidectomy and, 91
- 'resumed cartilage removal' during, 99
- strontium and, 100
- sunshine and, 31
- tooth changes during, 140-1
- vitamin D and, 32, 86, 100-2

**Saliva as source of P for enamel,**  
137, 138

**Scurvy**

- blood Ca and P levels during, 105
- bone changes in, 104-5, 125
- collagen content of bones and teeth during, 105
- osteoblasts and, 104, 125
- phosphatase during, 125
- tooth changes in, 142

**Sea urchin**

- blastomeres of, and Ca, 154
- permeability of, and Ca, 153

**Serum (*see also* Plasma)**

- diet on composition of, 120
- P : CO<sub>2</sub> ratio of, on bone, 120
- teeth, 140
- phosphatase in, during scurvy and rickets, 125

**Sheep, plasma Ca of,** 82

**Skeleton (*see also* Bone)**

- availability of, and age, 119
- composition, 44-5
- percentage of body-weight in adults, 42
- in children, 40

**Skin**

- Ca content of, 43
- Ca excretion by, 147, 158
- vitamin D precursors in, 31

**Sodium**

- exchange for Ca in apatite, 115, 116
- toxic action and Ca, 153

**Sodium azide on Ca reabsorption by renal tubules,** 148

**Solubility products and calcification,** 116-17, 123, 157

**South African Nutrition Council, recommended dietary standards of Ca,** 72

adults, 73, 76

children, 74, 80

infants, 74

lactation, 73, 78

pregnancy, 73, 77

**Spleen, Ca content of,** 43

phosphaturic factor in, 90

**Standards of Ca intake, 71-80**

adults, 73, 74-7

children, 74, 79-80

infants, 74, 78-9

lactation, 73, 77-8

pregnancy, 73, 77

rationale of, 72-3

**Starvation**

blood Ca level and, 85

during rickets, 85

healing of rickets and, 100

**Strontium**

calcification inactivation by, 129

calciotraumatic response in teeth and, 141

heterionic exchange in apatite by, 115-16

rickets and, 100

**Sulphate**

content of cartilage, 130

epiphyseal cartilage and Ca binding, 129

radio-S in ameloblasts and enamel, 140

**Sunlight, anti-rachitic effect of,** 31-2

**Surface exchange in bone crystals,** 115

**Sweat, Ca loss in,** 147, 158

**Teeth (*see also* Enamel and Dentin), 3, 135-43, 157**

**Teeth—cont.**

- calcio-traumatic response in, 141
- Ca uptake in, 138
- chondroitin sulphate in, 139
- collagen in during scurvy, 105
- composition, 135
- dietary factors and, 140–2
- glycogen in, 140
- hypophysectomy on, 142
- methods of studying, 137
- mucopolysaccharides in, 139–140, 158
- neonatal line in, 141
- oestrogens on, 143
- parathyroidectomy on, 140–1
- P exchange in, 137–9
- phosphatase and, 124, 140
- phosphate : carbonate ratio of, 140
- pigment layer on, 136
- scurvy and, 142
- structure of, 135
- vitamin A deficiency and, 142
- Testosterone, bone formation and, 107
  - after hypophysectomy, 105
- Tetany, 2, 55, 82
  - blood Ca and, 82
  - lactose and, 56
  - starvation and, 85
- Thrombin, 154
- Thymus, phosphaturic factor in, 90
- Thyroid gland
  - blood Ca level and, 87
  - bone formation and, 106
  - Ca excretion and, 87, 106, 148
  - intracerebral implantation with bone, 108
  - osteoporosis and, 106
  - urinary Ca and, 148
- Thyroidectomy
  - bone changes after, 106
  - dentin formation and, 142
  - thyroxine on bone changes after, 106

- Trachea, Ca content of, 43
- Tricalcium phosphate hydrate, Ca : P ratio of, 114
  - in bone, 114
- Trümmerfeld* in bone during scurvy, 104
- Tryptophane deficiency on teeth, 142
- Ultrafiltration of plasma Ca, 83
- Unit cell of bone crystal, 113
- Uranyl ions, exchange of, in apatite, 116
- Urine (*see* Renal excretion of Ca)
  - renal excretion of Ca in, 147–9
  - as index of intestinal absorption, 13
- Utilization of Ca, 3, 49, 156
  - adults and, 57–8
  - Ca salts by adults, 57–8
  - calculation of, 50–1
  - children and, 62
  - lactation and, 64–5
  - low Ca intake on, 58, 156
  - milk Ca by adults, 57–8, 75
  - pregnancy and, 77
- Vegetables
  - Ca content of, 5–6
  - source of dietary Ca, 1, 5–8, 155
- Versene, *see* Ethylenediamine tetra-acetic acid
- Vitamin A
  - bone and, 102–3
    - after parathyroidectomy, 91
  - deficiency on bone, 102–3
  - teeth, 142
  - hypervitaminosis on bone, 103
- Vitamin C (*see* Ascorbic acid and Scurvy), 104–5
- Vitamin D
  - activator of alkaline phosphatase, 33, 125
  - biological estimation of, 100
  - blood Ca and, 85–6, 149
  - blood P and, 86, 149

**Vitamin D—cont.**

bone and, 101-2, 155  
calciotraumatic response in  
teeth, and, 141

Ca retention in children and, 61

chemistry of, 30-1

citrate, blood and bone and, 131

deficiency on teeth, 140-1

dosage on teeth, 143

formation in skin, 31-2

hypervitaminosis on bone, 102

intestinal absorption of Ca and,  
1, 30-3, 155

kidney and, 86, 101, 149, 155  
after parathyroidectomy, 149

'line test' and, 100

metabolic bone changes in  
rickets and, 102

**Vitamin D—cont.**

mobilization of P by, 101

osteomalacia and, 59

parathyroids and, 87, 149

precursor in skin, 31-2

preen gland oil and, 31

rickets and, 32, 85-6, 98-100,  
157

**Vitamin E**

bone formation and, 105

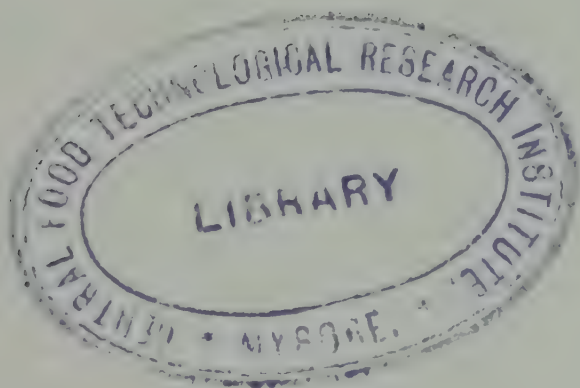
Ca storage in pregnancy and, 63

Vitamins of B group, bone and,  
103

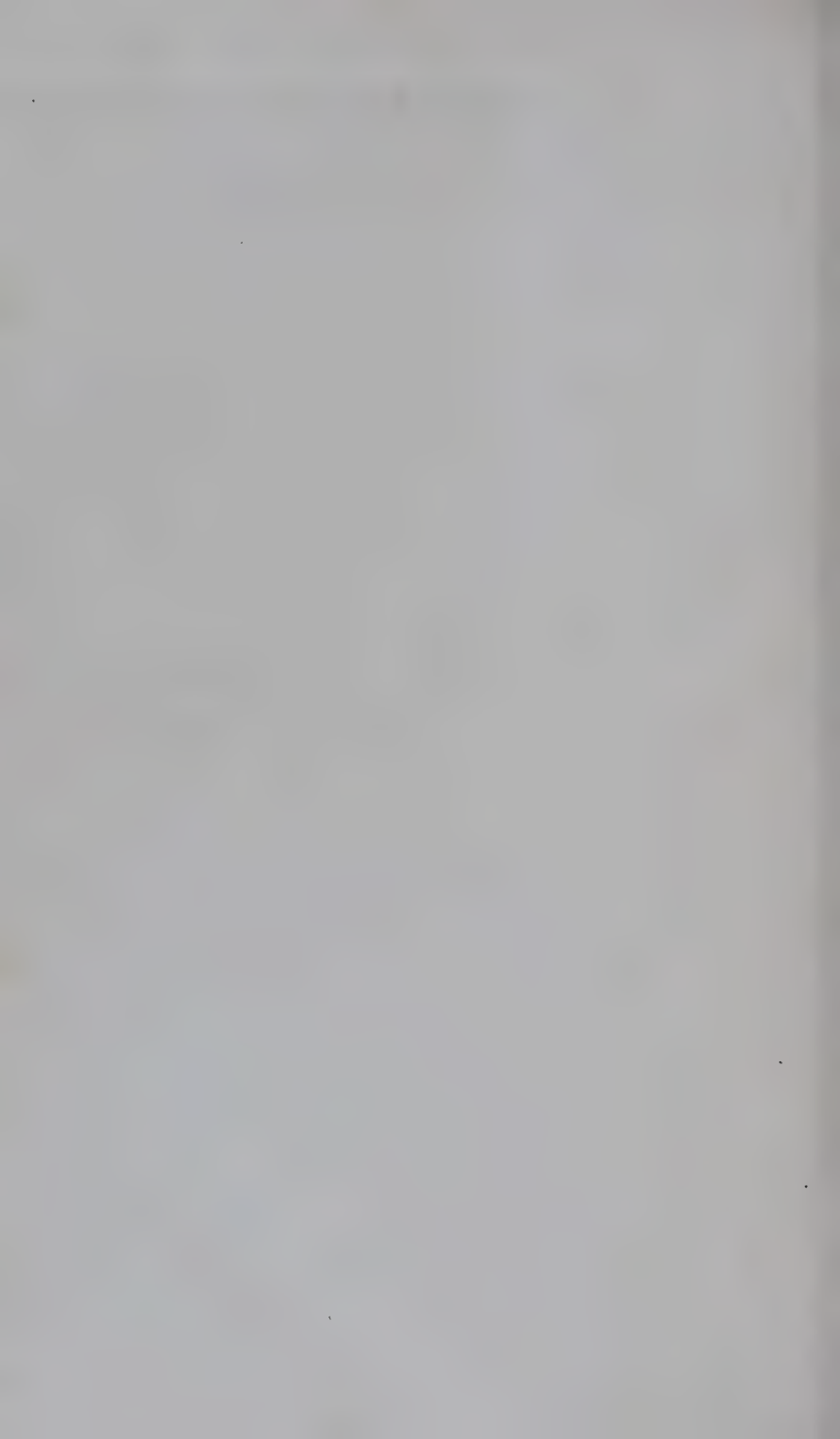
Vividdiffusion and plasma Ca, 83

Water, as source of Ca, 9, 155

Wound healing, phosphatase dur-  
ing scurvy and, 125















✓ C. F. T. R. I. LIBRARY, MYSORE.

✓ 6/8/80

CHECKED

5.5.97

CHECKED

2008

VERIFIED

2013



CFTRI-MYSORE



4045

Calcium metaboli..

